

Package ‘gsDesign’

July 20, 2026

Version 3.10.1

Title Group Sequential Design

Description Derives group sequential clinical trial designs and describes their properties. Particular focus on time-to-event, binary, and continuous outcomes. Largely based on methods described in Jennison, Christopher and Turnbull, Bruce W., 2000, ``Group Sequential Methods with Applications to Clinical Trials" ISBN: 0-8493-0316-8.

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URL <https://keaven.github.io/gsDesign/>,
<https://github.com/keaven/gsDesign>

BugReports <https://github.com/keaven/gsDesign/issues>

Encoding UTF-8

Depends R (>= 4.1.0)

Imports dplyr (>= 1.1.0), ggplot2 (>= 3.1.1), graphics, gt, methods, r2rtf, rlang, stats, tibble, tidyr, tools, xtable

Suggests covr, data.table, gridExtra, kableExtra, knitr, mvtnorm, rmarkdown, rpact, scales, testthat, utils, vdiff

VignetteBuilder knitr

Config/roxygen2/version 8.0.0

NeedsCompilation yes

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Repository CRAN

Date/Publication 2026-07-20 03:10:02 UTC

Contents

as_gt	3
as_rtf	4
as_table	7
binomialPowerTable	8
checkLengths	10
ciBinomial	12
condPower	18
eEvents	25
gsBinomialExact	27
gsBound	33
gsBoundCP	35
gsCP	37
gsDensity	41
gsDesign	44
gsProbability	51
gsSurvCalendar	54
gsSurvPower	58
nNormal	64
normalGrid	67
plot.gsDesign	69
print.nSurvival	72
repeatedPValueBinomialExact	77
sequentialPValue	78
sequentialPValueBinomialExact	80
sfExponential	82
sfHSD	84
sfLDOF	85
sfLinear	88
sfLogistic	91
sfPoints	94
sfPower	96
sfTDist	98
sfTruncated	100
sfXG1	103
simBinomialSeasonalExact	106
summary.gsDesign	108
summary.spendfn	114
tEventsIA	116
toBinomialExact	128
toInteger	130
xtable	132

Index

133

as_gt	<i>Convert a summary table object to a gt object</i>
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Description

Convert a summary table object created with [as_table](#) to a gt_tbl object; currently only implemented for [gsBinomialExact](#).

Usage

```
as_gt(x, ...)

## S3 method for class 'gsBinomialExactTable'
as_gt(
  x,
  ...,
  title = "Operating Characteristics for the Truncated SPRT Design",
  subtitle = "Assumes trial evaluated sequentially after each response",
  theta_label = html("Underlying<br>response rate"),
  bound_label = c("Futility bound", "Efficacy bound"),
  prob_decimals = 2,
  en_decimals = 1,
  rr_decimals = 0
)
```

Arguments

x	Object to be converted.
...	Other parameters that may be specific to the object.
title	Table title.
subtitle	Table subtitle.
theta_label	Label for theta.
bound_label	Label for bounds.
prob_decimals	Number of decimal places for probability of crossing.
en_decimals	Number of decimal places for expected number of observations when bound is crossed or when trial ends without crossing.
rr_decimals	Number of decimal places for response rates.

Details

Currently only implemented for [gsBinomialExact](#) objects. Creates a table to summarize an object. For [gsBinomialExact](#), this summarized operating characteristics across a range of effect sizes.

Value

A gt_tbl object that may be extended by overloaded versions of [as_gt](#).

See Also

```
vignette("binomialSPRTExample")
```

Examples

```
safety_design <- binomialSPRT(
  p0 = .04, p1 = .1, alpha = .04, beta = .2, minn = 4, maxn = 75
)
safety_power <- gsBinomialExact(
  k = length(safety_design$n.I),
  theta = seq(.02, .16, .02),
  n.I = safety_design$n.I,
  a = safety_design$lower$bound,
  b = safety_design$upper$bound
)
safety_power |>
  as_table() |>
  as_gt(
    theta_label = gt::html("Underlying<br>AE rate"),
    prob_decimals = 3,
    bound_label = c("low rate", "high rate")
  )
```

as_rtf

Save a summary table object as an RTF file

Description

Convert and save the summary table object created with [as_table](#) to an RTF file using `r2rtf`; currently only implemented for [gsBinomialExact](#).

Usage

```
as_rtf(x, ...)
```

```
## S3 method for class 'gsBinomialExactTable'
as_rtf(
  x,
  file,
  ...,
  title = paste("Operating Characteristics by Underlying Response Rate for",
    "Exact Binomial Group Sequential Design"),
  theta_label = "Underlying Response Rate",
  response_outcome = TRUE,
  bound_label = if (response_outcome) c("Futility Bound", "Efficacy Bound") else
    c("Efficacy Bound", "Futility Bound"),
  en_label = "Expected Sample Sizes",
  prob_decimals = 2,
```

```

        en_decimals = 1,
        rr_decimals = 0
    )

## S3 method for class 'gsBoundSummary'
as_rtf(
    x,
    file,
    ...,
    title = "Boundary Characteristics for Group Sequential Design",
    footnote_p_onesided = "one-side p-value for experimental better than control",
    footnote_appx_effect_at_bound = NULL,
    footnote_p_cross_h0 = "Cumulative type I error assuming binding futility bound",
    footnote_p_cross_h1 = "Cumulative power under the alternate hypothesis",
    footnote_specify = NULL,
    footnote_text = NULL
)

```

Arguments

x	Object to be saved as RTF file.
...	Other parameters that may be specific to the object.
file	File path for the output.
title	Title of the report.
theta_label	Label for theta.
response_outcome	Logical values indicating if the outcome is response rate (TRUE) or failure rate (FALSE). The default value is TRUE.
bound_label	Label for bounds. If the outcome is response rate, then the label is "Futility bound" and "Efficacy bound". If the outcome is failure rate, then the label is "Efficacy bound" and "Futility bound".
en_label	Label for expected number.
prob_decimals	Number of decimal places for probability of crossing.
en_decimals	Number of decimal places for expected number of observations when bound is crossed or when trial ends without crossing.
rr_decimals	Number of decimal places for response rates.
footnote_p_onesided	Footnote for one-side p-value.
footnote_appx_effect_at_bound	Footnote for approximate effect treatment at bound.
footnote_p_cross_h0	Footnote for cumulative type I error.
footnote_p_cross_h1	Footnote for cumulative power under the alternate hypothesis.
footnote_specify	Vector of string to put footnote super script.
footnote_text	Vector of string of footnote text.

Details

Currently only implemented for `gsBinomialExact` objects. Creates a table to summarize an object. For `gsBinomialExact`, this summarized operating characteristics across a range of effect sizes.

Value

`as_rtf()` returns the input `x` invisibly.

See Also

`vignette("binomialSPRTexample")`

Examples

```
# as_rtf for gsBinomialExact
zz <- gsBinomialExact(
  k = 3, theta = seq(0.1, 0.9, 0.1), n.I = c(12, 24, 36),
  a = c(-1, 0, 11), b = c(5, 9, 12)
)
zz |>
  as_table() |>
  as_rtf(
    file = tempfile(fileext = ".rtf"),
    title = "Power/Type I Error and Expected Sample Size for a Group Sequential Design"
  )

safety_design <- binomialSPRT(
  p0 = .04, p1 = .1, alpha = .04, beta = .2, minn = 4, maxn = 75
)
safety_power <- gsBinomialExact(
  k = length(safety_design$n.I),
  theta = seq(.02, .16, .02),
  n.I = safety_design$n.I,
  a = safety_design$lower$bound,
  b = safety_design$upper$bound
)
safety_power |>
  as_table() |>
  as_rtf(
    file = tempfile(fileext = ".rtf"),
    theta_label = "Underlying\nAE Rate",
    prob_decimals = 3,
    bound_label = c("Low Rate", "High Rate")
  )

# as_rtf for gsBoundSummary
xgs <- gsSurv(lambdaC = .2, hr = .5, eta = .1, T = 2, minfup = 1.5)
gsBoundSummary(xgs, timename = "Year", tdigits = 1) |> as_rtf(file = tempfile(fileext = ".rtf"))

ss <- nSurvival(
  lambda1 = .2, lambda2 = .1, eta = .1, Ts = 2, Tr = .5,
  sided = 1, alpha = .025, ratio = 2
)
```

```

xs <- gsDesign(nFixSurv = ss$n, n.fix = ss$nEvents, delta1 = log(ss$lambda2 / ss$lambda1))
gsBoundSummary(xs, logdelta = TRUE, ratio = ss$ratio) |> as_rtf(file = tempfile(fileext = ".rtf"))

xs <- gsDesign(nFixSurv = ss$n, n.fix = ss$nEvents, delta1 = log(ss$lambda2 / ss$lambda1))
gsBoundSummary(xs, logdelta = TRUE, ratio = ss$ratio) |>
  as_rtf(file = tempfile(fileext = ".rtf"),
    footnote_specify = "Z",
    footnote_text = "Z-Score")

```

as_table

*Create a summary table***Description**

Create a tibble to summarize an object; currently only implemented for [gsBinomialExact](#).

Usage

```

as_table(x, ...)

## S3 method for class 'gsBinomialExact'
as_table(x, ...)

```

Arguments

x Object to be summarized.

... Other parameters that may be specific to the object.

Details

Currently only implemented for [gsBinomialExact](#) objects. Creates a table to summarize an object. For [gsBinomialExact](#), this summarized operating characteristics across a range of effect sizes.

Value

A tibble which may have an extended class to enable further output processing.

See Also

```
vignette("binomialSPRTExample")
```

Examples

```

b <- binomialSPRT(p0 = .1, p1 = .35, alpha = .08, beta = .2, minn = 10, maxn = 25)
b_power <- gsBinomialExact(
  k = length(b$n.I), theta = seq(.1, .45, .05), n.I = b$n.I,
  a = b$lower$bound, b = b$upper$bound
)
b_power |>

```

```
as_table() |>
as_gt()
```

binomialPowerTable	<i>Power Table for Binomial Tests</i>
--------------------	---------------------------------------

Description

Creates a power table for binomial tests with various control group response rates and treatment effects. The function can compute power and Type I error either analytically or through simulation. With large simulations, the function is still fast and can produce exact power values to within simulation error.

Usage

```
binomialPowerTable(
  pC = c(0.8, 0.9, 0.95),
  delta = seq(-0.05, 0.05, 0.025),
  n = 70,
  ratio = 1,
  alpha = 0.025,
  delta0 = 0,
  scale = "Difference",
  failureEndpoint = TRUE,
  simulation = FALSE,
  nsim = 1e+06,
  adj = 0,
  chisq = 0
)
```

Arguments

pC	Vector of control group response rates.
delta	Vector of treatment effects (differences in response rates).
n	Total sample size.
ratio	Ratio of experimental to control sample size.
alpha	Type I error rate.
delta0	Non-inferiority margin.
scale	Scale for the test ("Difference", "RR", or "OR").
failureEndpoint	Logical indicating if the endpoint is a failure (TRUE) or success (FALSE).
simulation	Logical indicating whether to use simulation (TRUE) or analytical (FALSE) power calculation.
nsim	Number of simulations to run when simulation = TRUE.
adj	Use continuity correction for the testing (default is 0; only used if simulation = TRUE).
chisq	Chi-squared value for the test (default is 0; only used if simulation = TRUE).

Details

The function `binomialPowerTable()` creates a grid of all combinations of control group response rates and treatment effects. All out of range values (i.e., where the experimental group response rate is not between 0 and 1) are filtered out. For each combination, it computes the power either analytically using `nBinomial()` or through simulation using `simBinomial()`. When using simulation, the `simPowerBinomial()` function (not exported) is called internally to perform the simulations. Assuming p is the true probability of a positive test, the simulation standard error is

$$SE = \sqrt{p(1-p)/nsim}.$$

For example, when approximating an underlying Type I error rate of 0.025, the simulation standard error is 0.000156 with 1000000 simulations and the approximated power 95 is $0.025 \pm 1.96 * SE = 0.025 \pm 0.000306$.

Value

A data frame containing:

`pC` Control group response or failure rate.

`delta` Treatment effect.

`pE` Experimental group response or failure rate.

`Power` Power for the test (asymptotic or simulated).

See Also

[nBinomial](#), [simBinomial](#)

Examples

```
# Create a power table with analytical power calculation
power_table <- binomialPowerTable(
  pC = c(0.8, 0.9),
  delta = seq(-0.05, 0.05, 0.025),
  n = 70
)

# Create a power table with simulation
power_table_sim <- binomialPowerTable(
  pC = c(0.8, 0.9),
  delta = seq(-0.05, 0.05, 0.025),
  n = 70,
  simulation = TRUE,
  nsim = 10000
)
```

checkLengths

*Utility functions to verify variable properties***Description**

Utility functions to verify an objects's properties including whether it is a scalar or vector, the class, the length, and (if numeric) whether the range of values is on a specified interval. Additionally, the checkLengths function can be used to ensure that all the supplied inputs have equal lengths.

isInteger is similar to `is.integer` except that `isInteger(1)` returns TRUE whereas `is.integer(1)` returns FALSE.

checkScalar is used to verify that the input object is a scalar as well as the other properties specified above.

checkVector is used to verify that the input object is an atomic vector as well as the other properties as defined above.

checkRange is used to check whether the numeric input object's values reside on the specified interval. If any of the values are outside the specified interval, a FALSE is returned.

checkLength is used to check whether all of the supplied inputs have equal lengths.

Usage

```
checkLengths(..., allowSingle = FALSE)

checkRange(
  x,
  interval = 0:1,
  inclusion = c(TRUE, TRUE),
  varname = deparse(substitute(x)),
  tol = 0
)

checkScalar(x, isType = "numeric", ...)

checkVector(x, isType = "numeric", ..., length = NULL)

isInteger(x)
```

Arguments

...	For the <code>checkScalar</code> and <code>checkVector</code> functions, this input represents additional arguments sent directly to the <code>checkRange</code> function. For the <code>checkLengths</code> function, this input represents the arguments to check for equal lengths.
allowSingle	logical flag. If TRUE, arguments that are vectors comprised of a single element are not included in the comparative length test in the <code>checkLengths</code> function. Partial matching on the name of this argument is not performed so you must specify 'allowSingle' in its entirety in the call.

x	any object.
interval	two-element numeric vector defining the interval over which the input object is expected to be contained. Use the inclusion argument to define the boundary behavior.
inclusion	two-element logical vector defining the boundary behavior of the specified interval. A TRUE value denotes inclusion of the corresponding boundary. For example, if interval=c(3,6) and inclusion=c(FALSE,TRUE), then all the values of the input object are verified to be on the interval (3,6].
varname	character string defining the name of the input variable as sent into the function by the caller. This is used primarily as a mechanism to specify the name of the variable being tested when checkRange is being called within a function.
tol	numeric scalar defining the tolerance to use in testing the intervals of the checkRange function.
isType	character string defining the class that the input object is expected to be.
length	integer specifying the expected length of the object in the case it is a vector. If length=NULL, the default, then no length check is performed.

Value

isInteger: Boolean value as checking result Other functions have no return value, called for side effects

Examples

```
# check whether input is an integer
isInteger(1)
isInteger(1:5)
try(isInteger("abc")) # expect error

# check whether input is an integer scalar
checkScalar(3, "integer")

# check whether input is an integer scalar that resides
# on the interval on [3, 6]. Then test for interval (3, 6].
checkScalar(3, "integer", c(3, 6))
try(checkScalar(3, "integer", c(3, 6), c(FALSE, TRUE))) # expect error

# check whether the input is an atomic vector of class numeric,
# of length 3, and whose value all reside on the interval [1, 10)
x <- c(3, pi, exp(1))
checkVector(x, "numeric", c(1, 10), c(TRUE, FALSE), length = 3)

# do the same but change the expected length; expect error
try(checkVector(x, "numeric", c(1, 10), c(TRUE, FALSE), length = 2))

# create faux function to check input variable
foo <- function(moo) checkVector(moo, "character")
foo(letters)
try(foo(1:5)) # expect error with function and argument name in message
```

```
# check for equal lengths of various inputs
checkLengths(1:2, 2:3, 3:4)
try(checkLengths(1, 2, 3, 4:5)) # expect error

# check for equal length inputs but ignore single element vectors
checkLengths(1, 2, 3, 4:5, 7:8, allowSingle = TRUE)
```

ciBinomial

Testing, Confidence Intervals, Sample Size and Power for Comparing Two Binomial Rates

Description

Support is provided for sample size estimation, power, testing, confidence intervals and simulation for fixed sample size trials (that is, not group sequential or adaptive) with two arms and binary outcomes. Both superiority and non-inferiority trials are considered. While all routines default to comparisons of risk-difference, options to base computations on risk-ratio and odds-ratio are also included.

nBinomial() computes sample size or power using the method of Farrington and Manning (1990) for a trial to test the difference between two binomial event rates. The routine can be used for a test of superiority or non-inferiority. For a design that tests for superiority nBinomial() is consistent with the method of Fleiss, Tytun, and Ury (but without the continuity correction) to test for differences between event rates. This routine is consistent with the Hmisc package routines bsamsize and bpower for superiority designs. Vector arguments allow computing sample sizes for multiple scenarios for comparative purposes.

testBinomial() computes a Z- or Chi-square-statistic that compares two binomial event rates using the method of Miettinen and Nurminen (1980). This can be used for superiority or non-inferiority testing. Vector arguments allow easy incorporation into simulation routines for fixed, group sequential and adaptive designs.

ciBinomial() computes confidence intervals for 1) the difference between two rates, 2) the risk-ratio for two rates or 3) the odds-ratio for two rates. This procedure provides inference that is consistent with testBinomial() in that the confidence intervals are produced by inverting the testing procedures in testBinomial(). The Type I error alpha input to ciBinomial is always interpreted as 2-sided.

simBinomial() performs simulations to estimate the power for a Miettinen and Nurminen (1985) test comparing two binomial rates for superiority or non-inferiority. As noted in documentation for bpower.sim() in the Hmisc package, by using testBinomial() you can see that the formulas without any continuity correction are quite accurate. In fact, Type I error for a continuity-corrected test is significantly lower (Gordon and Watson, 1996) than the nominal rate. Thus, as a default no continuity corrections are performed.

varBinomial computes blinded estimates of the variance of the estimate of 1) event rate differences, 2) logarithm of the risk ratio, or 3) logarithm of the odds ratio. This is intended for blinded sample size re-estimation for comparative trials with a binary outcome.

Testing is 2-sided when a Chi-square statistic is used and 1-sided when a Z-statistic is used. Thus, these 2 options will produce substantially different results, in general. For non-inferiority, 1-sided testing is appropriate.

You may wish to round sample sizes up using `ceiling()`.

Farrington and Manning (1990) begin with event rates p_1 and p_2 under the alternative hypothesis and a difference between these rates under the null hypothesis, δ_0 . From these values, actual rates under the null hypothesis are computed, which are labeled p_{10} and p_{20} when `outtype=3`. The rates p_1 and p_2 are used to compute a variance for a Z-test comparing rates under the alternative hypothesis, while p_{10} and p_{20} are used under the null hypothesis. This computational method is also used to estimate variances in `varBinomial()` based on the overall event rate observed and the input treatment difference specified in δ_0 .

Sample size with `scale="Difference"` produces an error if $p_1 - p_2 = \delta_0$. Normally, the alternative hypothesis under consideration would be $p_1 - p_2 - \delta_0 > 0$. However, the alternative can have $p_1 - p_2 - \delta_0 < 0$.

Usage

```
ciBinomial(x1, x2, n1, n2, alpha = 0.05, adj = 0, scale = "Difference")
```

```
nBinomial(
  p1,
  p2,
  alpha = 0.025,
  beta = 0.1,
  delta0 = 0,
  ratio = 1,
  sided = 1,
  outtype = 1,
  scale = "Difference",
  n = NULL
)
```

```
simBinomial(
  p1,
  p2,
  n1,
  n2,
  delta0 = 0,
  nsim = 10000,
  chisq = 0,
  adj = 0,
  scale = "Difference"
)
```

```
testBinomial(
  x1,
  x2,
  n1,
```

```

    n2,
    delta0 = 0,
    chisq = 0,
    adj = 0,
    scale = "Difference",
    tol = 1e-11
)

varBinomial(x, n, delta0 = 0, ratio = 1, scale = "Difference")

```

Arguments

x1	Number of “successes” in the control group
x2	Number of “successes” in the experimental group
n1	Number of observations in the control group
n2	Number of observations in the experimental group
alpha	type I error; see sided below to distinguish between 1- and 2-sided tests
adj	With adj=1, the standard variance with a continuity correction is used for a Miettinen and Nurminen test statistic This includes a factor of $n/(n - 1)$ where n is the total sample size. If adj is not 1, this factor is not applied. The default is adj=0 since nominal Type I error is generally conservative with adj=1 (Gordon and Watson, 1996).
scale	“Difference”, “RR”, “OR”; see the scale parameter documentation above and Details. This is a scalar argument.
p1	event rate in group 1 under the alternative hypothesis
p2	event rate in group 2 under the alternative hypothesis
beta	type II error
delta0	A value of 0 (the default) always represents no difference between treatment groups under the null hypothesis. delta0 is interpreted differently depending on the value of the parameter scale. If scale="Difference" (the default), delta0 is the difference in event rates under the null hypothesis ($p_{10} - p_{20}$). If scale="RR", delta0 is the logarithm of the relative risk of event rates (p_{10} / p_{20}) under the null hypothesis. If scale="LNOR", delta0 is the difference in natural logarithm of the odds-ratio under the null hypothesis $\log(p_{10} / (1 - p_{10})) - \log(p_{20} / (1 - p_{20}))$.
ratio	sample size ratio for group 2 divided by group 1
sided	2 for 2-sided test, 1 for 1-sided test
outtype	nBinomial only; 1 (default) returns total sample size; 2 returns a data frame with sample size for each group (n1, n2; if n is not input as NULL, power is returned in Power; 3 returns a data frame with total sample size (n), sample size in each group (n1, n2), Type I error (alpha), 1 or 2 (sided, as input), Type II error (beta), power (Power), null and alternate hypothesis standard deviations (sigma0, sigma1), input event rates (p1, p2), null hypothesis difference in treatment group means (delta0) and null hypothesis event rates (p10, p20).

n	If power is to be computed in <code>nBinomial()</code> , input total trial sample size in n; this may be a vector. This is also the sample size in <code>varBinomial</code> , in which case the argument must be a scalar.
nsim	The number of simulations to be performed in <code>simBinomial()</code>
chisq	An indicator of whether or not a chi-square (as opposed to Z) statistic is to be computed. If <code>delta0=0</code> (default), the difference in event rates divided by its standard error under the null hypothesis is used. Otherwise, a Miettinen and Nurminen chi-square statistic for a 2 x 2 table is used.
tol	Default should probably be used; this is used to deal with a rounding issue in interim calculations
x	Number of “successes” in the combined control and experimental groups.

Value

`testBinomial()` and `simBinomial()` each return a vector of either Chi-square or Z test statistics. These may be compared to an appropriate cutoff point (e.g., `qnorm(.975)` for normal or `qchisq(.95,1)` for chi-square).

`ciBinomial()` returns a data frame with 1 row with a confidence interval; variable names are lower and upper.

`varBinomial()` returns a vector of (blinded) variance estimates of the difference of event rates (`scale="Difference"`), logarithm of the odds-ratio (`scale="OR"`) or logarithm of the risk-ratio (`scale="RR"`).

With the default `outtype=1`, `nBinomial()` returns a vector of total sample sizes is returned. With `outtype=2`, `nBinomial()` returns a data frame containing two vectors `n1` and `n2` containing sample sizes for groups 1 and 2, respectively; if `n` is input, this option also returns the power in a third vector, `Power`. With `outtype=3`, `nBinomial()` returns a data frame with the following columns:

n	A vector with total samples size required for each event rate comparison specified
n1	A vector of sample sizes for group 1 for each event rate comparison specified
n2	A vector of sample sizes for group 2 for each event rate comparison specified
alpha	As input
sided	As input
beta	As input; if <code>n</code> is input, this is computed
Power	If <code>n=NULL</code> on input, this is 1-beta; otherwise, the power is computed for each sample size input
sigma0	A vector containing the standard deviation of the treatment effect difference under the null hypothesis times <code>sqrt(n)</code> when <code>scale="Difference"</code> or <code>scale="OR"</code> ; when <code>scale="RR"</code> , this is the standard deviation time <code>sqrt(n)</code> for the numerator of the Farrington-Manning test statistic <code>x1-exp(delta0)*x2</code> .
sigma1	A vector containing the values as <code>sigma0</code> , in this case estimated under the alternative hypothesis.
p1	As input
p2	As input
p10	group 1 event rate used for null hypothesis
p20	group 2 event rate used for null hypothesis

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References

Farrington, CP and Manning, G (1990), Test statistics and sample size formulae for comparative binomial trials with null hypothesis of non-zero risk difference or non-unity relative risk. *Statistics in Medicine*; 9: 1447-1454.

Fleiss, JL, Tytun, A and Ury (1980), A simple approximation for calculating sample sizes for comparing independent proportions. *Biometrics*;36:343-346.

Gordon, I and Watson R (1985), The myth of continuity-corrected sample size formulae. *Biometrics*; 52: 71-76.

Miettinen, O and Nurminen, M (1985), Comparative analysis of two rates. *Statistics in Medicine*; 4 : 213-226.

See Also

[Normal](#), [uniroot](#)

Examples

```
# Compute z-test test statistic comparing 39/500 to 13/500
# use continuity correction in variance
x <- testBinomial(x1 = 39, x2 = 13, n1 = 500, n2 = 500, adj = 1)
x
pnorm(x, lower.tail = FALSE)

# Compute with unadjusted variance
x0 <- testBinomial(x1 = 39, x2 = 23, n1 = 500, n2 = 500)
x0
pnorm(x0, lower.tail = FALSE)

# Perform 50k simulations to test validity of the above
# asymptotic p-values
# (you may want to perform more to reduce standard error of estimate)
sum(as.double(x0) <=
  simBinomial(p1 = .078, p2 = .078, n1 = 500, n2 = 500, nsim = 10000)) / 10000
sum(as.double(x0) <=
  simBinomial(p1 = .052, p2 = .052, n1 = 500, n2 = 500, nsim = 10000)) / 10000

# Perform a non-inferiority test to see if p2=400 / 500 is within 5% of
# p1=410 / 500 use a z-statistic with unadjusted variance
x <- testBinomial(x1 = 410, x2 = 400, n1 = 500, n2 = 500, delta0 = -.05)
x
pnorm(x, lower.tail = FALSE)

# since chi-square tests equivalence (a 2-sided test) rather than
# non-inferiority (a 1-sided test),
# the result is quite different
pchisq(testBinomial(
```

```

    x1 = 410, x2 = 400, n1 = 500, n2 = 500, delta0 = -.05,
    chisq = 1, adj = 1
  ), 1, lower.tail = FALSE)

# now simulate the z-statistic without continuity corrected variance
sum(qnorm(.975) <=
  simBinomial(p1 = .8, p2 = .8, n1 = 500, n2 = 500, nsim = 100000)) / 100000

# compute a sample size to show non-inferiority
# with 5% margin, 90% power
nBinomial(p1 = .2, p2 = .2, delta0 = .05, alpha = .025, sided = 1, beta = .1)

# assuming a slight advantage in the experimental group lowers
# sample size requirement
nBinomial(p1 = .2, p2 = .19, delta0 = .05, alpha = .025, sided = 1, beta = .1)

# compute a sample size for comparing 15% vs 10% event rates
# with 1 to 2 randomization
nBinomial(p1 = .15, p2 = .1, beta = .2, ratio = 2, alpha = .05)

# now look at total sample size using 1-1 randomization
n <- nBinomial(p1 = .15, p2 = .1, beta = .2, alpha = .05)
n
# check if inputting sample size returns the desired power
nBinomial(p1 = .15, p2 = .1, beta = .2, alpha = .05, n = n)

# re-do with alternate output types
nBinomial(p1 = .15, p2 = .1, beta = .2, alpha = .05, outtype = 2)
nBinomial(p1 = .15, p2 = .1, beta = .2, alpha = .05, outtype = 3)

# look at power plot under different control event rate and
# relative risk reductions
library(dplyr)
library(ggplot2)
p1 <- seq(.075, .2, .000625)
len <- length(p1)
p2 <- c(p1 * .75, p1 * 2/3, p1 * .6, p1 * .5)
Reduction <- c(rep("25 percent", len), rep("33 percent", len),
  rep("40 percent", len), rep("50 percent", len))
df <- tibble(p1 = rep(p1, 4), p2, Reduction) |>
  mutate(`Sample size` = nBinomial(p1, p2, beta = .2, alpha = .025, sided = 1))
ggplot(df, aes(x = p1, y = `Sample size`, col = Reduction)) +
  geom_line() +
  xlab("Control group event rate") +
  ylim(0,6000) +
  ggtitle("Binomial sample size computation for 80 pct power")

# compute blinded estimate of treatment effect difference
x1 <- rbinom(n = 1, size = 100, p = .2)
x2 <- rbinom(n = 1, size = 200, p = .1)
# blinded estimate of risk difference variance
varBinomial(x = x1 + x2, n = 300, ratio = 2, delta0 = 0)
# blinded estimate of log-risk-ratio variance

```

```
varBinomial(x = x1 + x2, n = 300, ratio = 2, delta0 = 0, scale = "RR")
# blinded estimate of log-odds-ratio variance
varBinomial(x = x1 + x2, n = 300, ratio = 2, delta0 = 0, scale = "OR")
```

condPower

Sample size re-estimation based on conditional power

Description

`ssrCP()` adapts 2-stage group sequential designs to 2-stage sample size re-estimation designs based on interim analysis conditional power. This is a simple case of designs developed by Lehman and Wassmer, *Biometrics* (1999). The conditional power designs of Bauer and Kohne (1994), Proschan and Hunsberger (1995), Cui, Hung and Wang (1999) and Liu and Chi (2001), Gao, Ware and Mehta (2008), and Mehta and Pocock (2011). Either the estimated treatment effect at the interim analysis or any chosen effect size can be used for sample size re-estimation. If not done carefully, these designs can be very inefficient. It is probably a good idea to compare any design to a simpler group sequential design; see, for example, Jennison and Turnbull (2003). The α assumes a small Type I error is included with the interim analysis and that the design is an adaptation from a 2-stage group sequential design. Related functions include 3 pre-defined combination test functions (`z2NC`, `z2Z`, `z2Fisher`) that represent the inverse normal combination test (Lehman and Wassmer, 1999), the sufficient statistic for the complete data, and Fisher's combination test. `Power.ssrCP` computes unconditional power for a conditional power design derived by `ssrCP`.

`condPower` is a supportive routine that also is interesting in its own right; it computes conditional power of a combination test given an interim test statistic, stage 2 sample size and combination test statistic. While the returned data frame should make general plotting easy, the function `plot.ssrCP()` prints a plot of study sample size by stage 1 outcome with multiple x-axis labels for stage 1 z-value, conditional power, and stage 1 effect size relative to the effect size for which the underlying group sequential design was powered.

Sample size re-estimation using conditional power and an interim estimate of treatment effect was proposed by several authors in the 1990's (see references below). Statistical testing for these original methods was based on combination tests since Type I error could be inflated by using a sufficient statistic for testing at the end of the trial. Since 2000, more efficient variations of these conditional power designs have been developed. Fully optimized designs have also been derived (Posch et al, 2003, Lokhnygina and Tsiatis, 2008). Some of the later conditional power methods allow use of sufficient statistics for testing (Chen, DeMets and Lan, 2004, Gao, Ware and Mehta, 2008, and Mehta and Pocock, 2011).

The methods considered here are extensions of 2-stage group sequential designs that include both an efficacy and a futility bound at the planned interim analysis. A maximum fold-increase in sample size (`maxinc`) from the supplied group sequential design (`x`) is specified by the user, as well as a range of conditional power (`cpadj`) where sample size should be re-estimated at the interim analysis, 1-the targeted conditional power to be used for sample size re-estimation (`beta`) and a combination test statistic (`z2`) to be used for testing at the end of the trial. The input value `overrun` represents incremental enrollment not included in the interim analysis that is not included in the analysis; this is used for calculating the required number of patients enrolled to complete the trial.

Whereas most of the methods proposed have been based on using the interim estimated treatment effect size (default for `ssrCP`), the variable `theta` allows the user to specify an alternative; Liu and

Chi (2001) suggest that using the parameter value for which the trial was originally powered is a good choice.

Usage

```
condPower(
  z1,
  n2,
  z2 = z2NC,
  theta = NULL,
  x = gsDesign(k = 2, timing = 0.5, beta = beta),
  ...
)

ssrCP(
  z1,
  theta = NULL,
  maxinc = 2,
  overrun = 0,
  beta = x$beta,
  cpadj = c(0.5, 1 - beta),
  x = gsDesign(k = 2, timing = 0.5),
  z2 = z2NC,
  ...
)

## S3 method for class 'ssrCP'
plot(
  x,
  z1ticks = NULL,
  mar = c(7, 4, 4, 4) + 0.1,
  ylab = "Adapted sample size",
  xlaboffset = -0.2,
  lty = 1,
  col = 1,
  ...
)

z2NC(z1, x, ...)

z2Z(z1, x, n2 = x$n.I[2] - x$n.I[1], ...)

z2Fisher(z1, x, ...)

Power.ssrCP(x, theta = NULL, delta = NULL, r = 18)
```

Arguments

z1	Scalar or vector with interim standardized Z-value(s). Input of multiple values makes it easy to plot the revised sample size as a function of the interim test statistic.
n2	stage 2 sample size to be used in computing sufficient statistic when combining stage 1 and 2 test statistics.
z2	a combination function that returns a test statistic cutoff for the stage 2 test based in the interim test statistic supplied in z1, the design x and the stage 2 sample size.
theta	If NULL (default), conditional power calculation will be based on estimated interim treatment effect. Otherwise, theta is the standardized effect size used for conditional power calculation. Using the alternate hypothesis treatment effect can be more efficient than the estimated effect size; see Liu and Chi, Biometrics (2001).
x	A group sequential design with 2 stages (k=2) generated by gsDesign . For <code>plot.ssrCP</code> , x is a design returned by <code>ssrCP()</code> .
...	Allows passing of arguments that may be needed by the user-supplied function, <code>codez2</code> . In the case of <code>plot.ssrCP()</code> , allows passing more graphical parameters.
maxinc	Maximum fold-increase from planned maximum sample size in underlying group sequential design provided in x.
overrun	The number of patients enrolled before the interim analysis is completed who are not included in the interim analysis.
beta	Targeted Type II error (1 - targeted conditional power); used for conditional power in sample size reestimation.
cpadj	Range of values strictly between 0 and 1 specifying the range of interim conditional power for which sample size re-estimation is to be performed. Outside of this range, the sample size supplied in x is used.
z1ticks	Test statistic values at which tick marks are to be made on x-axis; automatically calculated under default of NULL
mar	Plot margins; see help for <code>par</code>
ylab	y-axis label
xlaboffset	offset on x-axis for printing x-axis labels
lty	line type for stage 2 sample size
col	line color for stage 2 sample size
delta	Natural parameter values for power calculation; see gsDesign for a description of how this is related to theta.
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.

Value

ssrCP returns a list with the following items:

x	As input.
z2fn	As input in z2.
theta	standardize effect size used for conditional power; if NULL is input, this is computed as $z1/\sqrt{n1}$ where n1 is the stage 1 sample size.
maxinc	As input.
overrun	As input.
beta	As input.
cpadj	As input.
dat	A data frame containing the input z1 values, computed cutoffs for the standard normal test statistic based solely on stage 2 data (z2), stage 2 sample sizes (n2), stage 2 conditional power (CP), standardize effect size used for conditional power calculation (theta), and the natural parameter value corresponding to theta (delta). The relation between theta and delta is determined by the delta0 and delta1 values from x: $\text{delta} = \text{delta0} + \text{theta}(\text{delta1} - \text{delta0})$.

Author(s)

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References

- Bauer, Peter and Kohne, F., Evaluation of experiments with adaptive interim analyses, *Biometrics*, 50:1029-1041, 1994.
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- Gao, P, Ware, JH and Mehta, C, Sample size re-estimation for adaptive sequential design in clinical trials, *Journal of Biopharmaceutical Statistics*", 18:1184-1196, 2008.
- Jennison, C and Turnbull, BW. Mid-course sample size modification in clinical trials based on the observed treatment effect. *Statistics in Medicine*, 22:971-993", 2003.
- Lehmacher, W and Wassmer, G. Adaptive sample size calculations in group sequential trials, *Biometrics*, 55:1286-1290, 1999.
- Liu, Q and Chi, GY., On sample size and inference for two-stage adaptive designs, *Biometrics*, 57:172-177, 2001.
- Mehta, C and Pocock, S. Adaptive increase in sample size when interim results are promising: A practical guide with examples, *Statistics in Medicine*, 30:3267-3284, 2011.

See Also

[gsDesign](#)

Examples

```

library(ggplot2)
# quick trick for simple conditional power based on interim z-value, stage 1 and 2 sample size
# assumed treatment effect and final alpha level
# and observed treatment effect
alpha <- .01 # set final nominal significance level
timing <- .6 # set proportion of sample size, events or statistical information at IA
n2 <- 40 # set stage 2 sample size events or statistical information
hr <- .6 # for this example we will derive conditional power based on hazard ratios
n.fix <- nEvents(hr=hr,alpha=alpha) # you could otherwise make n.fix an arbitrary positive value
# this just derives a group sequential design that should not change sample size from n.fix
# due to stringent IA bound
x <- gsDesign(k=2,n.fix=n.fix,alpha=alpha,test.type=1,sfu=sfHSD,
sfupar=-20,timing=timing,delta1=log(hr))
# derive effect sizes for which you wish to compute conditional power
hrpostIA = seq(.4,1,.05)
# in the following, we convert HR into standardized effect size based on the design in x
powr <- condPower(x=x,z1=1,n2=x$n.I[2]-x$n.I[1],theta=log(hrpostIA)/x$delta1*x$theta[2])
ggplot(
  data.frame(
    x = hrpostIA,
    y = condPower(
      x = x,
      z1 = 1,
      n2 = x$n.I[2] - x$n.I[1],
      theta = log(hrpostIA) / x$delta1 * x$theta[2]
    )
  ),
  aes(x = x, y = y)
) +
  geom_line() +
  labs(
    x = "HR post IA",
    y = "Conditional power",
    title = "Conditional power as a function of assumed HR"
  )

# Following is a template for entering parameters for ssrCP
# Natural parameter value null and alternate hypothesis values
delta0 <- 0
delta1 <- 1
# timing of interim analysis for underlying group sequential design
timing <- .5
# upper spending function
sfu <- sfHSD
# upper spending function parameter
sfupar <- -12
# maximum sample size inflation
maxinflation <- 2
# assumed enrollment overrun at IA
overrun <- 25
# interim z-values for plotting

```

```

z <- seq(0, 4, .025)
# Type I error (1-sided)
alpha <- .025
# Type II error for design
beta <- .1
# Fixed design sample size
n.fix <- 100
# conditional power interval where sample
# size is to be adjusted
cpadj <- c(.3, .9)
# targeted Type II error when adapting sample size
betastar <- beta
# combination test (built-in options are: z2Z, z2NC, z2Fisher)
z2 <- z2NC

# use the above parameters to generate design
# generate a 2-stage group sequential design with
x <- gsDesign(
  k = 2, n.fix = n.fix, timing = timing, sfu = sfu, sfupar = sfupar,
  alpha = alpha, beta = beta, delta0 = delta0, delta1 = delta1
)
# extend this to a conditional power design
xx <- ssrCP(
  x = x, z1 = z, overrun = overrun, beta = betastar, cpadj = cpadj,
  maxinc = maxinflation, z2 = z2
)
# plot the stage 2 sample size
plot(xx)
# demonstrate overlays on this plot
# overlay with densities for z1 under different hypotheses
lines(z, 80 + 240 * dnorm(z, mean = 0), col = 2)
lines(z, 80 + 240 * dnorm(z, mean = sqrt(x$n.I[1]) * x$theta[2]), col = 3)
lines(z, 80 + 240 * dnorm(z, mean = sqrt(x$n.I[1]) * x$theta[2] / 2), col = 4)
lines(z, 80 + 240 * dnorm(z, mean = sqrt(x$n.I[1]) * x$theta[2] * .75), col = 5)
axis(side = 4, at = 80 + 240 * seq(0, .4, .1), labels = as.character(seq(0, .4, .1)))
mtext(side = 4, expression(paste("Density for ", z[1])), line = 2)
text(x = 1.5, y = 90, col = 2, labels = expression(paste("Density for ", theta, "=0")))
text(x = 3.00, y = 180, col = 3, labels = expression(paste("Density for ", theta, "=",
  theta[1])))
text(x = 1.00, y = 180, col = 4, labels = expression(paste("Density for ", theta, "=",
  theta[1], "/2")))
text(x = 2.5, y = 140, col = 5, labels = expression(paste("Density for ", theta, "=",
  theta[1], "*.75")))
# overall line for max sample size
nalt <- xx$maxinc * x$n.I[2]
lines(x = par("usr")[1:2], y = c(nalt, nalt), lty = 2)

# compare above design with different combination tests
# use sufficient statistic for final testing
xxZ <- ssrCP(
  x = x, z1 = z, overrun = overrun, beta = betastar, cpadj = cpadj,
  maxinc = maxinflation, z2 = z2Z
)

```

```

# use Fisher combination test for final testing
xxFisher <- ssrCP(
  x = x, z1 = z, overrun = overrun, beta = betastar, cpadj = cpadj,
  maxinc = maxinflation, z2 = z2Fisher
)
# combine data frames from these designs
y <- rbind(
  data.frame(xx$dat, Test = "Normal combination"),
  data.frame(xxZ$dat, Test = "Sufficient statistic"),
  data.frame(xxFisher$dat, Test = "Fisher combination")
)
# plot stage 2 statistic required for positive combination test
ggplot2::ggplot(data = y, ggplot2::aes(x = z1, y = z2, col = Test)) +
ggplot2::geom_line()
# plot total sample size versus stage 1 test statistic
ggplot2::ggplot(data = y, ggplot2::aes(x = z1, y = n2, col = Test)) +
ggplot2::geom_line()
# check achieved Type I error for sufficient statistic design
Power.ssrCP(x = xxZ, theta = 0)

# compare designs using observed vs planned theta for conditional power
xxtheta1 <- ssrCP(
  x = x, z1 = z, overrun = overrun, beta = betastar, cpadj = cpadj,
  maxinc = maxinflation, z2 = z2, theta = x$delta
)
# combine data frames for the 2 designs
y <- rbind(
  data.frame(xx$dat, "CP effect size" = "Obs. at IA"),
  data.frame(xxtheta1$dat, "CP effect size" = "Alt. hypothesis")
)
# plot stage 2 sample size by design
ggplot2::ggplot(data = y, ggplot2::aes(x = z1, y = n2, col = CP.effect.size)) +
ggplot2::geom_line()
# compare power and expected sample size
y1 <- Power.ssrCP(x = xx)
y2 <- Power.ssrCP(x = xxtheta1)
# combine data frames for the 2 designs
y3 <- rbind(
  data.frame(y1, "CP effect size" = "Obs. at IA"),
  data.frame(y2, "CP effect size" = "Alt. hypothesis")
)
# plot expected sample size by design and effect size
ggplot2::ggplot(data = y3, ggplot2::aes(x = delta, y = en, col = CP.effect.size)) +
ggplot2::geom_line() +
ggplot2::xlab(expression(delta)) +
ggplot2::ylab("Expected sample size")
# plot power by design and effect size
ggplot2::ggplot(data = y3, ggplot2::aes(x = delta, y = Power, col = CP.effect.size)) +
ggplot2::geom_line() +
ggplot2::xlab(expression(delta))

```

eEvents

*Expected number of events for a time-to-event study***Description**

eEvents() is used to calculate the expected number of events for a population with a time-to-event endpoint. It is based on calculations demonstrated in Lachin and Foulkes (1986) and is fundamental in computations for the sample size method they propose. Piecewise exponential survival and dropout rates are supported as well as piecewise uniform enrollment. A stratified population is allowed. Output is the expected number of events observed given a trial duration and the above rate parameters.

Usage

```
eEvents(
  lambda = 1,
  eta = 0,
  gamma = 1,
  R = 1,
  S = NULL,
  T = 2,
  Tfinal = NULL,
  minfup = 0,
  digits = 4
)

## S3 method for class 'eEvents'
print(x, digits = 4, ...)
```

Arguments

lambda	Scalar, vector or matrix of event hazard rates; rows represent time periods while columns represent strata; a vector implies a single stratum.
eta	Scalar, vector or matrix of dropout hazard rates; rows represent time periods while columns represent strata; if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
gamma	A scalar, vector or matrix of rates of entry by time period (rows) and strata (columns); if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
R	A scalar or vector of durations of time periods for recruitment rates specified in rows of gamma. Length is the same as number of rows in gamma. In eEvents(), recruitment after the final period is assumed to be 0 if T exceeds sum(R); the final period is only extended in designs where T is solved for (e.g., nSurv with T = NULL).

S	A scalar or vector of durations of piecewise constant event rates specified in rows of <code>lambda</code> , <code>eta</code> and <code>etaE</code> ; this is NULL if there is a single event rate per stratum (exponential failure) or length of the number of rows in <code>lambda</code> minus 1, otherwise.
T	Time of analysis; if <code>Tfinal</code> =NULL, this is also the study duration.
Tfinal	Study duration; if NULL, this will be replaced with T on output.
minfup	Time from end of planned enrollment (<code>sum(R)</code> from output value of R) until <code>Tfinal</code> .
digits	Which controls number of digits for printing.
x	An object of class <code>eEvents</code> returned from <code>eEvents()</code> .
...	Other arguments that may be passed to the generic print function.

Details

`eEvents()` produces an object of class `eEvents` with the number of subjects and events for a set of pre-specified trial parameters, such as accrual duration and follow-up period. The underlying power calculation is based on Lachin and Foulkes (1986) method for proportional hazards assuming a fixed underlying hazard ratio between 2 treatment groups. The method has been extended here to enable designs to test non-inferiority. Piecewise constant enrollment and failure rates are assumed and a stratified population is allowed. See also [nSurvival](#) for other Lachin and Foulkes (1986) methods assuming a constant hazard difference or exponential enrollment rate.

`print.eEvents()` formats the output for an object of class `eEvents` and returns the input value.

Value

`eEvents()` and `print.eEvents()` return an object of class `eEvents` which contains the following items:

<code>lambda</code>	As input; converted to a matrix on output.
<code>eta</code>	As input; converted to a matrix on output.
<code>gamma</code>	As input.
<code>R</code>	As input.
<code>S</code>	As input.
<code>T</code>	As input.
<code>Tfinal</code>	Planned duration of study.
<code>minfup</code>	As input.
<code>d</code>	Expected number of events.
<code>n</code>	Expected sample size.
<code>digits</code>	As input.

Author(s)

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References

Lachin JM and Foulkes MA (1986), Evaluation of Sample Size and Power for Analyses of Survival with Allowance for Nonuniform Patient Entry, Losses to Follow-Up, Noncompliance, and Stratification. *Biometrics*, 42, 507-519.

Bernstein D and Lagakos S (1978), Sample size and power determination for stratified clinical trials. *Journal of Statistical Computation and Simulation*, 8:65-73.

See Also

`vignette("gsDesignPackageOverview")`, [plot.gsDesign](#), [gsDesign](#), [gsHR](#), [nSurvival](#)

Examples

```
# 3 enrollment periods, 3 piecewise exponential failure rates
str(eEvents(
  lambda = c(.05, .02, .01), eta = .01, gamma = c(5, 10, 20),
  R = c(2, 1, 2), S = c(1, 1), T = 20
))

# Control group for example from Bernstein and Lagakos (1978)
lamC <- c(1, .8, .5)
n <- eEvents(
  lambda = matrix(c(lamC, lamC * 2 / 3), ncol = 6), eta = 0,
  gamma = matrix(.5, ncol = 6), R = 2, T = 4
)
```

gsBinomialExact

One-Sample Binomial Routines

Description

`gsBinomialExact` computes power/Type I error and expected sample size for a group sequential design in a single-arm trial with a binary outcome. This can also be used to compare event rates in two-arm studies. The print function has been extended using `print.gsBinomialExact` to print `gsBinomialExact` objects. Similarly, a plot function has been extended using `plot.gsBinomialExact` to plot `gsBinomialExact` objects.

`binomialSPRT` computes a truncated binomial sequential probability ratio test (SPRT) which is a specific instance of an exact binomial group sequential design for a single arm trial with a binary outcome.

`gsBinomialPP` computes a truncated binomial (group) sequential design based on predictive probability.

`nBinomial1Sample` uses exact binomial calculations to compute power and sample size for single arm binomial experiments.

`gsBinomialExact` is based on the book "Group Sequential Methods with Applications to Clinical Trials," Christopher Jennison and Bruce W. Turnbull, Chapter 12, Section 12.1.2 Exact Calculations for Binary Data. This computation is often used as an approximation for the distribution of the

number of events in one treatment group out of all events when the probability of an event is small and sample size is large.

An object of class `gsBinomialExact` is returned. On output, the values of `theta` input to `gsBinomialExact` will be the parameter values for which the boundary crossing probabilities and expected sample sizes are computed.

Note that `a[1]` equal to `-1` lower bound at `n.I[1]` means 0 successes continues at interim 1; `a[2]==0` at interim 2 means 0 successes stops trial for futility at 2nd analysis. For final analysis, set `a[k]` equal to `b[k]-1` to incorporate all possibilities into non-positive trial; see example.

The sequential probability ratio test (SPRT) is a sequential testing scheme allowing testing after each observation. This likelihood ratio is used to determine upper and lower cutoffs which are linear and parallel in the number of responses as a function of sample size. `binomialSPRT` produces a variation the the SPRT that tests only within a range of sample sizes. While the linear SPRT bounds are continuous, actual bounds are the integer number of response at or beyond each linear bound for each sample size where testing is performed. Because of the truncation and discretization of the bounds, power and Type I error achieve will be lower than the nominal levels specified by `alpha` and `beta` which can be altered to produce desired values that are achieved by the planned sample size. See also example that shows computation of Type I error when futility bound is considered non-binding.

Note that if the objective of a design is to demonstrate that a rate (e.g., failure rate) is lower than a certain level, two approaches can be taken. First, 1 minus the failure rate is the success rate and this can be used for planning. Second, the role of `beta` becomes to express Type I error and `alpha` is used to express Type II error.

Plots produced include boundary plots, expected sample size, response rate at the boundary and power.

`gsBinomial1Sample` uses exact binomial computations based on the base R functions `qbinom()` and `pbinom()`. The tabular output may be convenient for plotting. Note that input variables are largely not checked, so the user is largely responsible for results; it is a good idea to do a run with `outtype=3` to check that you have done things appropriately. If `n` is not ordered (a bad idea) or not sequential (maybe OK), be aware of possible consequences.

`nBinomial1Sample` is based on code from Marc Schwartz <marc_schwartz@me.com>. The possible sample size vector `n` needs to be selected in such a fashion that it covers the possible range of values that include the true minimum. NOTE: the one-sided evaluation of significance is more conservative than using the 2-sided exact test in `binom.test`.

Usage

```
gsBinomialExact(
  k = 2,
  theta = c(0.1, 0.2),
  n.I = c(50, 100),
  a = c(3, 7),
  b = c(20, 30)
)

binomialSPRT(
  p0 = 0.05,
  p1 = 0.25,
```

```

    alpha = 0.1,
    beta = 0.15,
    minn = 10,
    maxn = 35
  )

## S3 method for class 'gsBinomialExact'
plot(x, plottype = 1, ...)

## S3 method for class 'binomialSPRT'
plot(x, plottype = 1, ...)

nBinomial1Sample(
  p0 = 0.9,
  p1 = 0.95,
  alpha = 0.025,
  beta = NULL,
  n = 200:250,
  outtype = 1,
  conservative = FALSE
)
```

Arguments

k	Number of analyses planned, including interim and final.
theta	Vector of possible underlying binomial probabilities for a single binomial sample.
n.I	Sample size at analyses (increasing positive integers); vector of length k.
a	Number of "successes" required to cross lower bound cutoffs to reject p1 in favor of p0 at each analysis; vector of length k; -1 (minimum allowed) means no lower bound.
b	Number of "successes" required to cross upper bound cutoffs for rejecting p0 in favor of p1 at each analysis; vector of length k; value > n.I means no upper bound.
p0	Lower of the two response (event) rates hypothesized.
p1	Higher of the two response (event) rates hypothesized.
alpha	Nominal probability of rejecting response (event) rate p0 when it is true.
beta	Nominal probability of rejecting response (event) rate p1 when it is true. If NULL, Type II error will be computed for all input values of n and output will be in a data frame.
minn	Minimum sample size at which sequential testing begins.
maxn	Maximum sample size.
x	Item of class gsBinomialExact or binomialSPRT for print.gsBinomialExact. Item of class gsBinomialExact for plot.gsBinomialExact. Item of class binomialSPRT for item of class plot.binomialSPRT.

plottype	1 produces a plot with counts of response at bounds (for binomialSPRT, also produces linear SPRT bounds); 2 produces a plot with power to reject null and alternate response rates as well as the probability of not crossing a bound by the maximum sample size; 3 produces a plot with the response rate at the boundary as a function of sample size when the boundary is crossed; 6 produces a plot of the expected sample size by the underlying event rate (this assumes there is no enrollment beyond the sample size where the boundary is crossed).
...	arguments passed through to ggplot.
n	sample sizes to be considered for nBinomial1Sample. These should be ordered from smallest to largest and be > 0.
outtype	Operative when beta != NULL. 1 means routine will return a single integer sample size while for output=2 a data frame is returned (see Value); note that this not operative is beta is NULL in which case a data table is returned with Type II error and power for each input value of n.
conservative	operative when outtype=1 or 2 and beta != NULL. Default FALSE selects minimum sample size for which power is at least 1-beta. When conservative=TRUE, the minimum sample size for which power is at least 1-beta and there is no larger sample size in the input n where power is less than 1-beta.

Value

gsBinomialExact() returns a list of class gsBinomialExact and gsProbability (see example); when displaying one of these objects, the default function to print is print.gsProbability(). The object returned from gsBinomialExact() contains the following elements:

k	As input.
theta	As input.
n.I	As input.
lower	A list containing two elements: bound is as input in a and prob is a matrix of boundary crossing probabilities. Element i, j contains the boundary crossing probability at analysis i for the j-th element of theta input. All boundary crossing is assumed to be binding for this computation; that is, the trial must stop if a boundary is crossed.
upper	A list of the same form as lower containing the upper bound and upper boundary crossing probabilities.
en	A vector of the same length as theta containing expected sample sizes for the trial design corresponding to each value in the vector theta.

binomialSPRT produces an object of class binomialSPRT that is an extension of the gsBinomialExact class. The values returned in addition to those returned by gsBinomialExact are:

intercept	A vector of length 2 with the intercepts for the two SPRT bounds.
slope	A scalar with the common slope of the SPRT bounds.
alpha	As input. Note that this will exceed the actual Type I error achieved by the design returned.
beta	As input. Note that this will exceed the actual Type II error achieved by the design returned.

p0 As input.
 p1 As input.

nBinomial1Sample produces a data frame with power for each input value in n if beta=NULL. Otherwise, a sample size achieving the desired power is returned unless the minimum power for the values input in n is greater than or equal to the target or the maximum yields power less than the target, in which case an error message is shown. The input variable outtype has no effect if beta=NULL. Otherwise, outtype=1 results in return of an integer sample size and outtype=2 results in a data frame with one record which includes the desired sample size. When a data frame is returned, the variables include:

p0 Input null hypothesis event (or response) rate.
 p1 Input alternative hypothesis (or response) rate; must be > p0.
 alpha Input Type I error.
 beta Input Type II error except when input is NULL in which case realized Type II error is computed.
 n sample size.
 b cutoff given n to control Type I error; value is NULL if no such value exists.
 alphaR Type I error achieved for each output value of n; less than or equal to the input value alpha.
 Power Power achieved for each output value of n.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Jon Hartzel, Yevgen Tymofyeyev and Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Code for nBinomial1Sample was based on code developed by <marc_schwartz@me.com>.

See Also

[gsProbability](#)

Examples

```
library(ggplot2)

zz <- gsBinomialExact(
  k = 3, theta = seq(0.1, 0.9, 0.1), n.I = c(12, 24, 36),
  a = c(-1, 0, 11), b = c(5, 9, 12)
)
```

```

# let's see what class this is
class(zz)

# because of "gsProbability" class above, following is equivalent to
# \code{print.gsProbability(zz)}
zz

# also plot (see also plots below for \code{binomialSPRT})
# add lines using geom_line()
plot(zz) +
ggplot2::geom_line()

# now for SPRT examples
x <- binomialSPRT(p0 = .05, p1 = .25, alpha = .1, beta = .2)
# boundary plot
plot(x)
# power plot
plot(x, plotype = 2)
# Response (event) rate at boundary
plot(x, plotype = 3)
# Expected sample size at boundary crossing or end of trial
plot(x, plotype = 6)

# sample size for single arm exact binomial

# plot of table of power by sample size
# note that outtype need not be specified if beta is NULL
nb1 <- nBinomial1Sample(p0 = 0.05, p1=0.2,alpha = 0.025, beta=NULL, n = 25:40)
nb1
library(scales)
ggplot2::ggplot(nb1, ggplot2::aes(x = n, y = Power)) +
ggplot2::geom_line() +
ggplot2::geom_point() +
ggplot2::scale_y_continuous(labels = percent)

# simple call with same parameters to get minimum sample size yielding desired power
nBinomial1Sample(p0 = 0.05, p1 = 0.2, alpha = 0.025, beta = .2, n = 25:40)

# change to 'conservative' if you want all larger sample
# sizes to also provide adequate power
nBinomial1Sample(p0 = 0.05, p1 = 0.2, alpha = 0.025, beta = .2, n = 25:40, conservative = TRUE)

# print out more information for the selected derived sample size
nBinomial1Sample(p0 = 0.05, p1 = 0.2, alpha = 0.025, beta = .2, n = 25:40, conservative = TRUE,
  outtype = 2)
# Reproduce realized Type I error alphaR
stats::pbinom(q = 5, size = 39, prob = .05, lower.tail = FALSE)
# Reproduce realized power
stats::pbinom(q = 5, size = 39, prob = 0.2, lower.tail = FALSE)
# Reproduce critical value for positive finding
stats::qbinom(p = 1 - .025, size = 39, prob = .05) + 1
# Compute p-value for 7 successes

```

```

stats::pbinom(q = 6, size = 39, prob = .05, lower.tail = FALSE)
# what happens if input sample sizes not sufficient?
## Not run:
  nBinomial1Sample(p0 = 0.05, p1 = 0.2, alpha = 0.025, beta = .2, n = 25:30)

## End(Not run)

```

gsBound

Boundary derivation - low level

Description

gsBound() and gsBound1() are lower-level functions used to find boundaries for a group sequential design. They are not recommended (especially gsBound1()) for casual users. These functions do not adjust sample size as gsDesign() does to ensure appropriate power for a design.

gsBound() computes upper and lower bounds given boundary crossing probabilities assuming a mean of 0, the usual null hypothesis. gsBound1() computes the upper bound given a lower boundary, upper boundary crossing probabilities and an arbitrary mean (theta).

The function gsBound1() requires special attention to detail and knowledge of behavior when a design corresponding to the input parameters does not exist.

Usage

```
gsBound(I, trueneg, falsepos, tol = 1e-06, r = 18, printerr = 0)
```

```
gsBound1(theta, I, a, probhi, tol = 1e-06, r = 18, printerr = 0)
```

Arguments

I	Vector containing statistical information planned at each analysis.
trueneg	Vector of desired probabilities for crossing upper bound assuming mean of 0.
falsepos	Vector of desired probabilities for crossing lower bound assuming mean of 0.
tol	Tolerance for error (scalar; default is 0.000001). Normally this will not be changed by the user. This does not translate directly to number of digits of accuracy, so use extra decimal places.
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.
printerr	If this scalar argument set to 1, this will print messages from underlying C program. Mainly intended to notify user when an output solution does not match input specifications. This is not intended to stop execution as this often occurs when deriving a design in gsDesign that uses beta-spending.

theta	Scalar containing mean (drift) per unit of statistical information.
a	Vector containing lower bound that is fixed for use in gsBound1.
prophi	Vector of desired probabilities for crossing upper bound assuming mean of theta.

Value

Both routines return a list. Common items returned by the two routines are:

k	The length of vectors input; a scalar.
theta	As input in gsBound1(); 0 for gsBound().
I	As input.
a	For gsbound1, this is as input. For gsbound this is the derived lower boundary required to yield the input boundary crossing probabilities under the null hypothesis.
b	The derived upper boundary required to yield the input boundary crossing probabilities under the null hypothesis.
tol	As input.
r	As input.
error	Error code. 0 if no error; greater than 0 otherwise.

gsBound() also returns the following items:

rates	a list containing two items:
falsepos	vector of upper boundary crossing probabilities as input.
trueneg	vector of lower boundary crossing probabilities as input.

gsBound1() also returns the following items:

problo	vector of lower boundary crossing probabilities; computed using input lower bound and derived upper bound.
prophi	vector of upper boundary crossing probabilities as input.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("gsDesignPackageOverview")`, [gsDesign](#), [gsProbability](#)

Examples

```

# set boundaries so that probability is .01 of first crossing
# each upper boundary and .02 of crossing each lower boundary
# under the null hypothesis
x <- gsBound(
  I = c(1, 2, 3) / 3, trueneg = rep(.02, 3),
  falsepos = rep(.01, 3)
)
x

# use gsBound1 to set up boundary for a 1-sided test
x <- gsBound1(
  theta = 0, I = c(1, 2, 3) / 3, a = rep(-20, 3),
  probhi = c(.001, .009, .015)
)
x$b

# check boundary crossing probabilities with gsProbability
y <- gsProbability(k = 3, theta = 0, n.I = x$I, a = x$a, b = x$b)$upper$prob

# Note that gsBound1 only computes upper bound
# To get a lower bound under a parameter value theta:
#   use minus the upper bound as a lower bound
#   replace theta with -theta
#   set probhi as desired lower boundary crossing probabilities
# Here we let set lower boundary crossing at 0.05 at each analysis
# assuming theta=2.2
y <- gsBound1(
  theta = -2.2, I = c(1, 2, 3) / 3, a = -x$b,
  probhi = rep(.05, 3)
)
y$b

# Now use gsProbability to look at design
# Note that lower boundary crossing probabilities are as
# specified for theta=2.2, but for theta=0 the upper boundary
# crossing probabilities are smaller than originally specified
# above after first interim analysis
gsProbability(k = length(x$b), theta = c(0, 2.2), n.I = x$I, b = x$b, a = -y$b)

```

gsBoundCP

*Conditional Power at Interim Boundaries***Description**

gsBoundCP() computes the total probability of crossing future upper bounds given an interim test statistic at an interim bound. For each interim boundary, assumes an interim test statistic at the boundary and computes the probability of crossing any of the later upper boundaries.

See Conditional power section of manual for further clarification. See also Muller and Schaffer (2001) for background theory.

Usage

```
gsBoundCP(x, theta = "thetahat", r = 18)
```

Arguments

x	An object of type gsDesign or gsProbability
theta	if "thetahat" and class(x)!="gsDesign", conditional power computations for each boundary value are computed using estimated treatment effect assuming a test statistic at that boundary ($z_i/\sqrt{x\$n.I[i]}$) at analysis i, interim test statistic z_i and interim sample size/statistical information of $x\$n.I[i]$. Otherwise, conditional power is computed assuming the input scalar value theta.
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.

Value

A list containing two vectors, CPlo and CPhi.

CPlo	A vector of length $x\$k-1$ with conditional powers of crossing upper bounds given interim test statistics at each lower bound
CPhi	A vector of length $x\$k-1$ with conditional powers of crossing upper bounds given interim test statistics at each upper bound.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

- Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.
- Muller, Hans-Helge and Schaffer, Helmut (2001), Adaptive group sequential designs for clinical trials: combining the advantages of adaptive and classical group sequential approaches. *Biometrics*;57:886-891.

See Also

[gsDesign](#), [gsProbability](#), [gsCP](#)

Examples

```
# set up a group sequential design
x <- gsDesign(k = 5)
x

# compute conditional power based on interim treatment effects
gsBoundCP(x)

# compute conditional power based on original x$delta
gsBoundCP(x, theta = x$delta)
```

 gsCP

Conditional and Predictive Power, Overall and Conditional Probability of Success

Description

gsCP() computes conditional boundary crossing probabilities at future planned analyses for a given group sequential design assuming an interim z-statistic at a specified interim analysis. While gsCP() is designed toward computing conditional power for a variety of underlying parameter values, [condPower](#) is built to compute conditional power for a variety of interim test statistic values which is useful for sample size adaptation (see [ssrCP](#)). gsPP() averages conditional power across a posterior distribution to compute predictive power. gsPI() computes Bayesian prediction intervals for future analyses corresponding to results produced by gsPP(). gsPosterior() computes the posterior density for the group sequential design parameter of interest given a prior density and an interim outcome that is exact or in an interval. gsPOS() computes the probability of success for a trial using a prior distribution to average power over a set of theta values of interest. gsCPOS() assumes no boundary has been crossed before and including an interim analysis of interest, and computes the probability of success based on this event. Note that gsCP() and gsPP() take only the interim test statistic into account in computing conditional probabilities, while gsCPOS() conditions on not crossing any bound through a specified interim analysis.

See Conditional power section of manual for further clarification. See also Muller and Schaffer (2001) for background theory.

For gsPP(), gsPI(), gsPOS() and gsCPOS(), the prior distribution for the standardized parameter theta () for a group sequential design specified through a gsDesign object is specified through the arguments theta and wgts. This can be a discrete or a continuous probability density function. For a discrete function, generally all weights would be 1. For a continuous density, the wgts would contain integration grid weights, such as those provided by [normalGrid](#).

For gsPosterior, a prior distribution in prior must be composed of the vectors z density. The vector z contains points where the prior is evaluated and density the corresponding density or, for a discrete distribution, the probabilities of each point in z. Densities may be supplied as from normalGrid() where grid weights for numerical integration are supplied in gridwgts. If gridwgts are not supplied, they are defaulted to 1 (equal weighting). To ensure a proper prior distribution, you must have sum(gridwgts * density) equal to 1; this is NOT checked, however.

Usage

```

gsCP(x, theta = NULL, i = 1, zi = 0, r = 18)

gsPP(
  x,
  i = 1,
  zi = 0,
  theta = c(0, 3),
  wgts = c(0.5, 0.5),
  r = 18,
  total = TRUE
)

gsPI(
  x,
  i = 1,
  zi = 0,
  j = 2,
  level = 0.95,
  theta = c(0, 3),
  wgts = c(0.5, 0.5)
)

gsPosterior(x = gsDesign(), i = 1, zi = NULL, prior = normalGrid(), r = 18)

gsPOS(x, theta, wgts)

gsCPOS(i, x, theta, wgts)

```

Arguments

x	An object of type <code>gsDesign</code> or <code>gsProbability</code>
theta	a vector with θ value(s) at which conditional power is to be computed; for <code>gsCP()</code> if <code>NULL</code> , an estimated value of θ based on the interim test statistic ($z_i/\sqrt{x\$n.I[i]}$) as well as at <code>x\$theta</code> is computed. For <code>gsPosterior</code> , this may be a scalar or an interval; for <code>gsPP</code> and <code>gsCP</code> , this must be a scalar.
i	analysis at which interim z-value is given; must be from 1 to <code>x\$k-1</code>
zi	interim z-value at analysis <code>i</code> (scalar)
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with <code>r = 16</code> . This parameter is normally not changed by users.
wgts	Weights to be used with grid points in <code>theta</code> . Length can be one if weights are equal, otherwise should be the same length as <code>theta</code> . Values should be positive, but do not need to sum to 1.

total	The default of total=TRUE produces the combined probability for all planned analyses after the interim analysis specified in i. Otherwise, information on each analysis is provided separately.
j	specific analysis for which prediction is being made; must be >i and no more than x\$k
level	The level to be used for Bayes credible intervals (which approach confidence intervals for vague priors). The default level=.95 corresponds to a 95% credible interval. level=0 provides a point estimate rather than an interval.
prior	provides a prior distribution in the form produced by normalGrid

Value

gsCP() returns an object of the class gsProbability. Based on the input design and the interim test statistic, the output gsDesign object has bounds for test statistics computed based on solely on observations after interim i. Boundary crossing probabilities are computed for the input θ values. See manual and examples.

gsPP() if total==TRUE, returns a real value indicating the predictive power of the trial conditional on the interim test statistic z_i at analysis i; otherwise returns vector with predictive power for each future planned analysis.

gsPI() returns an interval (or point estimate if level=0) indicating 100level% credible interval for the z-statistic at analysis j conditional on the z-statistic at analysis $i < j$. The interval does not consider intervening interim analyses. The probability estimate is based on the predictive distribution used for gsPP() and requires a prior distribution for the group sequential parameter theta specified in theta and wgts.

gsPosterior() returns a posterior distribution containing the the vector z input in prior\$z, the posterior density in density, grid weights for integrating the posterior density as input in prior\$gridwgts or defaulted to a vector of ones, and the product of the output values in density and gridwgts in wgts.

gsPOS() returns a real value indicating the probability of a positive study weighted by the prior distribution input for theta.

gsCPOS() returns a real value indicating the probability of a positive study weighted by the posterior distribution derived from the interim test statistic and the prior distribution input for theta conditional on an interim test statistic.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Proschan, Michael A., Lan, KK Gordon and Wittes, Janet Turk (2006), *Statistical Monitoring of Clinical Trials*. NY: Springer.

Muller, Hans-Helge and Schaffer, Helmut (2001), Adaptive group sequential designs for clinical trials: combining the advantages of adaptive and classical group sequential approaches. *Biometrics*;57:886-891.

See Also

[normalGrid](#), [gsDesign](#), [gsProbability](#), [gsBoundCP](#), [ssrCP](#), [condPower](#)

Examples

```
library(ggplot2)
# set up a group sequential design
x <- gsDesign(k = 5)
x

# set up a prior distribution for the treatment effect
# that is normal with mean .75*x$delta and standard deviation x$delta/2
mu0 <- .75 * x$delta
sigma0 <- x$delta / 2
prior <- normalGrid(mu = mu0, sigma = sigma0)

# compute POS for the design given the above prior distribution for theta
gsPOS(x = x, theta = prior$z, wgts = prior$wgts)

# assume POS should only count cases in prior where theta >= x$delta/2
gsPOS(x = x, theta = prior$z, wgts = prior$wgts * (prior$z >= x$delta / 2))

# assuming a z-value at lower bound at analysis 2, what are conditional
# boundary crossing probabilities for future analyses
# assuming theta values from x as well as a value based on the interim
# observed z
CP <- gsCP(x, i = 2, zi = x$lower$bound[2])
CP

# summing values for crossing future upper bounds gives overall
# conditional power for each theta value
CP$theta
t(CP$upper$prob) %*% c(1, 1, 1)

# compute predictive probability based on above assumptions
gsPP(x, i = 2, zi = x$lower$bound[2], theta = prior$z, wgts = prior$wgts)

# if it is known that boundary not crossed at interim 2, use
# gsCPOS to compute conditional POS based on this
gsCPOS(x = x, i = 2, theta = prior$z, wgts = prior$wgts)

# 2-stage example to compare results to direct computation
x <- gsDesign(k = 2)
z1 <- 0.5
n1 <- x$n.I[1]
```

```

n2 <- x$n.I[2] - x$n.I[1]
thetahat <- z1 / sqrt(n1)
theta <- c(thetahat, 0, x$delta)

# conditional power direct computation - comparison w gsCP
pnorm((n2 * theta + z1 * sqrt(n1) - x$upper$bound[2] * sqrt(n1 + n2)) / sqrt(n2))

gsCP(x = x, zi = z1, i = 1)$upper$prob

# predictive power direct computation - comparison w gsPP
# use same prior as above
mu0 <- .75 * x$delta * sqrt(x$n.I[2])
sigma2 <- (.5 * x$delta)^2 * x$n.I[2]
prior <- normalGrid(mu = .75 * x$delta, sigma = x$delta / 2)
gsPP(x = x, zi = z1, i = 1, theta = prior$z, wgts = prior$wgts)
t <- .5
z1 <- .5
b <- z1 * sqrt(t)
# direct from Proschan, Lan and Wittes eqn 3.10
# adjusted drift at n.I[2]
pnorm(((b - x$upper$bound[2]) * (1 + t * sigma2) +
  (1 - t) * (mu0 + b * sigma2)) /
  sqrt((1 - t) * (1 + sigma2) * (1 + t * sigma2)))

# plot prior then posterior distribution for unblinded analysis with i=1, zi=1
xp <- gsPosterior(x = x, i = 1, zi = 1, prior = prior)
plot(x = xp$z, y = xp$density, type = "l", col = 2, xlab = expression(theta), ylab = "Density")
points(x = x$z, y = x$density, col = 1)

# add posterior plot assuming only knowlede that interim bound has
# not been crossed at interim 1
xpb <- gsPosterior(x = x, i = 1, zi = 1, prior = prior)
lines(x = xpb$z, y = xpb$density, col = 4)

# prediction interval based in interim 1 results
# start with point estimate, followed by 90% prediction interval
gsPI(x = x, i = 1, zi = z1, j = 2, theta = prior$z, wgts = prior$wgts, level = 0)
gsPI(x = x, i = 1, zi = z1, j = 2, theta = prior$z, wgts = prior$wgts, level = .9)

```

gsDensity

Group sequential design interim density function

Description

Given an interim analysis i of a group sequential design and a vector of real values z_i , `gsDensity()` computes an interim density function at analysis i at the values in z_i . For each value in z_i , this interim density is the derivative of the probability that the group sequential trial does not cross a boundary prior to the i -th analysis and at the i -th analysis the interim Z-statistic is less than that value. When integrated over the real line, this density computes the probability of not crossing

a bound at a previous analysis. It corresponds to the subdistribution function at analysis i that excludes the probability of crossing a bound at an earlier analysis.

The initial purpose of this routine was as a component needed to compute the predictive power for a trial given an interim result; see [gsPP](#).

See Jennison and Turnbull (2000) for details on how these computations are performed.

Usage

```
gsDensity(x, theta = 0, i = 1, zi = 0, r = 18)
```

Arguments

<code>x</code>	An object of type <code>gsDesign</code> or <code>gsProbability</code>
<code>theta</code>	a vector with θ value(s) at which the interim density function is to be computed.
<code>i</code>	analysis at which interim z-values are given; must be from 1 to <code>x\$k</code>
<code>zi</code>	interim z-value at analysis i (scalar)
<code>r</code>	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.

Value

<code>zi</code>	The input vector <code>zi</code> .
<code>theta</code>	The input vector <code>theta</code> .
<code>density</code>	A matrix with <code>length(zi)</code> rows and <code>length(theta)</code> columns. The subdensity function for <code>z[j]</code> , <code>theta[m]</code> at analysis i is returned in <code>density[j,m]</code> .

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

[gsDesign](#), [gsProbability](#), [gsBoundCP](#)

Examples

```

library(ggplot2)
# set up a group sequential design
x <- gsDesign()

# set theta values where density is to be evaluated
theta <- x$theta[2] * c(0, .5, 1, 1.5)

# set zi values from -1 to 7 where density is to be evaluated
zi <- seq(-3, 7, .05)

# compute subdensity values at analysis 2
y <- gsDensity(x, theta = theta, i = 2, zi = zi)

# plot sub-density function for each theta value
plot(y$zi, y$density[, 3],
     type = "l", xlab = "Z",
     ylab = "Interim 2 density", lty = 3, lwd = 2
)
lines(y$zi, y$density[, 2], lty = 2, lwd = 2)
lines(y$zi, y$density[, 1], lwd = 2)
lines(y$zi, y$density[, 4], lty = 4, lwd = 2)
title("Sub-density functions at interim analysis 2")
legend(
  x = c(3.85, 7.2), y = c(.27, .385), lty = 1:5, lwd = 2, cex = 1.5,
  legend = c(
    expression(paste(theta, "=0.0")),
    expression(paste(theta, "=0.5", delta)),
    expression(paste(theta, "=1.0", delta)),
    expression(paste(theta, "=1.5", delta))
  )
)

# add vertical lines with lower and upper bounds at analysis 2
# to demonstrate how likely it is to continue, stop for futility
# or stop for efficacy at analysis 2 by treatment effect
lines(rep(x$upper$bound[2], 2), c(0, .4), col = 2)
lines(rep(x$lower$bound[2], 2), c(0, .4), lty = 2, col = 2)

# Replicate part of figures 8.1 and 8.2 of Jennison and Turnbull text book
# O'Brien-Fleming design with four analyses

x <- gsDesign(k = 4, test.type = 2, sfu = "OF", alpha = .1, beta = .2)

z <- seq(-4.2, 4.2, .05)
d <- gsDensity(x = x, theta = x$theta, i = 4, zi = z)

plot(z, d$density[, 1], type = "l", lwd = 2, ylab = expression(paste(p[4], "(z,", theta, ")")))
lines(z, d$density[, 2], lty = 2, lwd = 2)
u <- x$upper$bound[4]
text(expression(paste(theta, "=", delta)), x = 2.2, y = .2, cex = 1.5)
text(expression(paste(theta, "=0")), x = .55, y = .4, cex = 1.5)

```

Description

`gsDesign()` is used to find boundaries and trial size required for a group sequential design.

Many parameters normally take on default values and thus do not require explicit specification. One- and two-sided designs are supported. Two-sided designs may be symmetric or asymmetric. Wang-Tsiatis designs, including O'Brien-Fleming and Pocock designs can be generated. Designs with common spending functions as well as other built-in and user-specified functions for Type I error and futility are supported. Type I error computations for asymmetric designs may assume binding or non-binding lower bounds. The print function has been extended using `print.gsDesign()` to print `gsDesign` objects; see examples.

The user may ignore the structure of the value returned by `gsDesign()` if the standard printing and plotting suffice; see examples.

`delta` and `n.fix` are used together to determine what sample size output options the user seeks. The default, `delta=0` and `n.fix=1`, results in a 'generic' design that may be used with any sampling situation. Sample size ratios are provided and the user multiplies these times the sample size for a fixed design to obtain the corresponding group sequential analysis times. If `delta>0`, `n.fix` is ignored, and `delta` is taken as the standardized effect size - the signal to noise ratio for a single observation; for example, the mean divided by the standard deviation for a one-sample normal problem. In this case, the sample size at each analysis is computed. When `delta=0` and `n.fix>1`, `n.fix` is assumed to be the sample size for a fixed design with no interim analyses. See examples below.

Following are further comments on the input argument `test.type` which is used to control what type of error measurements are used in trial design. The manual may also be worth some review in order to see actual formulas for boundary crossing probabilities for the various options. Options 3 and 5 assume the trial stops if the lower bound is crossed for Type I and Type II error computation (binding lower bound). For the purpose of computing Type I error, options 4 and 6 assume the trial continues if the lower bound is crossed (non-binding lower bound); that is a Type I error can be made by crossing an upper bound after crossing a previous lower bound. Beta-spending refers to error spending for the lower bound crossing probabilities under the alternative hypothesis (options 3 and 4). In this case, the final analysis lower and upper boundaries are assumed to be the same. The appropriate total beta spending (power) is determined by adjusting the maximum sample size through an iterative process for all options. Since options 3 and 4 must compute boundary crossing probabilities under both the null and alternative hypotheses, deriving these designs can take longer than other options. Options 5 and 6 compute lower bound spending under the null hypothesis.

Usage

```
gsDesign(  
  k = 3,  
  test.type = 4,  
  alpha = 0.025,  
  beta = 0.1,
```

```

    astar = 0,
    delta = 0,
    n.fix = 1,
    timing = 1,
    sfu = sfHSD,
    sfupar = -4,
    sfl = sfHSD,
    sflpar = -2,
    sfharm = sfHSD,
    sfharmparam = -2,
    tol = 1e-06,
    r = 18,
    n.I = 0,
    maxn.IPlan = 0,
    nFixSurv = 0,
    endpoint = NULL,
    delta1 = 1,
    delta0 = 0,
    overrun = 0,
    usTime = NULL,
    lsTime = NULL,
    testUpper = TRUE,
    testLower = TRUE,
    testHarm = TRUE
  )

## S3 method for class 'gsDesign'
xtable(
  x,
  caption = NULL,
  label = NULL,
  align = NULL,
  digits = NULL,
  display = NULL,
  ...
)

```

Arguments

k	Number of analyses planned, including interim and final.
test.type	1=one-sided 2=two-sided symmetric 3=two-sided, asymmetric, beta-spending with binding lower bound 4=two-sided, asymmetric, beta-spending with non-binding lower bound 5=two-sided, asymmetric, lower bound spending under the null hypothesis with binding lower bound 6=two-sided, asymmetric, lower bound spending under the null hypothesis with non-binding lower bound

	7=two-sided, asymmetric, with binding futility and binding harm bounds 8=two-sided, asymmetric, with non-binding futility and non-binding harm bounds. See details, examples and manual.
alpha	Type I error, always one-sided. Default value is 0.025.
beta	Type II error, default value is 0.1 (90% power).
astar	Total spending for the lower (test.type 5 or 6) or harm (test.type 7 or 8) bound under the null hypothesis. Default is 0. For test.type 5 or 6, astar specifies the total probability of crossing a lower bound at all analyses combined. For test.type 7 or 8, astar specifies the total probability of crossing the harm bound at all analyses combined under the null hypothesis. If $astar = 0$, it will be changed to $1 - \alpha$.
delta	Effect size for theta under alternative hypothesis. This can be set to the standardized effect size to generate a sample size if <code>n.fix=NULL</code> . See details and examples.
n.fix	Sample size for fixed design with no interim; used to find maximum group sequential sample size. For a time-to-event outcome, input number of events required for a fixed design rather than sample size and enter fixed design sample size (optional) in <code>nFixSurv</code> . See details and examples.
timing	Sets relative timing of interim analyses. Default of 1 produces equally spaced analyses. Otherwise, this is a vector of length k or $k-1$. The values should satisfy $0 < timing[1] < timing[2] < \dots < timing[k-1] < timing[k]=1$.
sfu	A spending function or a character string indicating a boundary type (that is, “WT” for Wang-Tsiatis bounds, “OF” for O’Brien-Fleming bounds and “Pocock” for Pocock bounds). For one-sided and symmetric two-sided testing is used to completely specify spending (test.type=1, 2), sfu. The default value is sfHSD which is a Hwang-Shih-DeCani spending function. See details, vignette(“SpendingFunction0v manual and examples.
sfupar	Real value, default is -4 which is an O’Brien-Fleming-like conservative bound when used with the default Hwang-Shih-DeCani spending function. This is a real-vector for many spending functions. The parameter sfupar specifies any parameters needed for the spending function specified by sfu; this is not needed for spending functions (sfLDOF, sfLDPocock) or bound types (“OF”, “Pocock”) that do not require parameters. Note that sfupar can be specified as a positive scalar for sfLDOF for a generalized O’Brien-Fleming spending function.
sf1	Specifies the spending function for lower boundary crossing probabilities when asymmetric, two-sided testing is performed (test.type = 3, 4, 5, or 6). Unlike the upper bound, only spending functions are used to specify the lower bound. The default value is sfHSD which is a Hwang-Shih-DeCani spending function. The parameter sf1 is ignored for one-sided testing (test.type=1) or symmetric 2-sided testing (test.type=2). See details, spending functions, manual and examples.
sf1par	Real value, default is -2 , which, with the default Hwang-Shih-DeCani spending function, specifies a less conservative spending rate than the default for the upper bound.
sfharm	A spending function for the harm bound, used with test.type = 7 or test.type = 8. Default is sfHSD. See spendingFunction for details.

sfharmparam	Real value, default is -2 . Parameter for the harm bound spending function sfharm.
tol	Tolerance for error (default is 0.000001). Normally this will not be changed by the user. This does not translate directly to number of digits of accuracy, so use extra decimal places.
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.
n.I	Used for re-setting bounds when timing of analyses changes from initial design; see examples.
maxn.IPlan	Used for re-setting bounds when timing of analyses changes from initial design; see examples.
nFixSurv	If a time-to-event variable is used, nFixSurv computed as the sample size from nSurvival may be entered to have gsDesign compute the total sample size required as well as the number of events at each analysis that will be returned in n.fix; this is rounded up to an even number.
endpoint	An optional character string that should represent the type of endpoint used for the study. This may be used by output functions. Types most likely to be recognized initially are "TTE" for time-to-event outcomes with fixed design sample size generated by nSurvival() and "Binomial" for 2-sample binomial outcomes with fixed design sample size generated by nBinomial().
delta1	delta1 and delta0 may be used to store information about the natural parameter scale compared to delta that is a standardized effect size. delta1 is the alternative hypothesis parameter value on the natural parameter scale (e.g., the difference in two binomial rates).
delta0	delta0 is the null hypothesis parameter value on the natural parameter scale.
overrun	Scalar or vector of length $k-1$ with patients enrolled that are not included in each interim analysis.
usTime	Default is NULL in which case upper bound spending time is determined by timing. Otherwise, this should be a vector of length k with the spending time at each analysis (see Details section of gsDesign).
lsTime	Default is NULL in which case lower bound spending time is determined by timing. Otherwise, this should be a vector of length k with the spending time at each analysis (see Details section of gsDesign).
testUpper	Indicator of which analyses should include an upper (efficacy) bound. A single value of TRUE (default) indicates all analyses have an efficacy bound. Otherwise, a logical vector of length k indicating which analyses will have an efficacy bound. Overridden to all TRUE for test.type 1 and 2. Must be TRUE at the final analysis to achieve targeted power. At each analysis, at least one of testUpper, testLower, or testHarm must be TRUE. Where testUpper is FALSE, the upper bound is set to $+2\theta$ (effectively Inf) and displayed as NA in output.

<code>testLower</code>	Indicator of which analyses should include a lower (futility) bound. A single value of TRUE (default) indicates all analyses have a lower bound; FALSE indicates none. Otherwise, a logical vector of length <code>k</code> . Ignored for <code>test.type</code> 1 (one-sided, no lower bound). Overridden to all TRUE for <code>test.type</code> 2 (symmetric). For <code>test.type</code> 3–8, at least one analysis must be TRUE. Where <code>testLower</code> is FALSE, the lower bound is set to -2θ (effectively $-\text{Inf}$) and displayed as NA in output.
<code>testHarm</code>	Indicator of which analyses should include a harm bound. A single value of TRUE (default) indicates all analyses have a harm bound; FALSE indicates none. Otherwise, a logical vector of length <code>k</code> . Only used for <code>test.type</code> 7 or 8; at least one analysis must be TRUE for those types. Where <code>testHarm</code> is FALSE, the harm bound is set to -2θ (effectively $-\text{Inf}$) and displayed as NA in output.
<code>x</code>	An R object of class found among <code>methods(xtable)</code> . See below on how to write additional method functions for <code>xtable</code> .
<code>caption</code>	Character vector of length 1 or 2 containing the table's caption or title. If length is 2, the second item is the "short caption" used when LaTeX generates a "List of Tables". Set to NULL to suppress the caption. Default value is NULL.
<code>label</code>	Character vector of length 1 containing the LaTeX label or HTML anchor. Set to NULL to suppress the label. Default value is NULL.
<code>align</code>	Character vector of length equal to the number of columns of the resulting table, indicating the alignment of the corresponding columns. Also, " " may be used to produce vertical lines between columns in LaTeX tables, but these are effectively ignored when considering the required length of the supplied vector. If a character vector of length one is supplied, it is split as <code>strsplit(align, "")[[1]]</code> before processing. Since the row names are printed in the first column, the length of <code>align</code> is one greater than <code>ncol(x)</code> if <code>x</code> is a <code>data.frame</code> . Use "l", "r", and "c" to denote left, right, and center alignment, respectively. Use "p{3cm}" etc. for a LaTeX column of the specified width. For HTML output the "p" alignment is interpreted as "l", ignoring the width request. Default depends on the class of <code>x</code> .
<code>digits</code>	Numeric vector of length equal to one (in which case it will be replicated as necessary) or to the number of columns of the resulting table or matrix of the same size as the resulting table, indicating the number of digits to display in the corresponding columns. Since the row names are printed in the first column, the length of the vector <code>digits</code> or the number of columns of the matrix <code>digits</code> is one greater than <code>ncol(x)</code> if <code>x</code> is a <code>data.frame</code> . Default depends on the class of <code>x</code> . If values of <code>digits</code> are negative, the corresponding values of <code>x</code> are displayed in scientific format with <code>abs(digits)</code> digits.
<code>display</code>	Character vector of length equal to the number of columns of the resulting table, indicating the format for the corresponding columns. Since the row names are printed in the first column, the length of <code>display</code> is one greater than <code>ncol(x)</code> if <code>x</code> is a <code>data.frame</code> . These values are passed to the <code>formatC</code> function. Use "d" (for integers), "f", "e", "E", "g", "G", "fg" (for reals), or "s" (for strings). "f" gives numbers in the usual xxx.xxx format; "e" and "E" give n.ddde+nn or n.dddE+nn (scientific format); "g" and "G" put <code>x[i]</code> into scientific format only if it saves space to do so. "fg" uses fixed format as "f", but <code>digits</code> as number

of *significant* digits. Note that this can lead to quite long result strings. Default depends on the class of *x*.

... Additional arguments. (Currently ignored.)

Value

An object of the class `gsDesign`. This class has the following elements and upon return from `gsDesign()` contains:

<code>k</code>	As input.
<code>test.type</code>	As input.
<code>alpha</code>	As input.
<code>beta</code>	As input.
<code>astar</code>	As input, except when <code>test.type=5</code> or <code>6</code> and <code>astar</code> is input as <code>0</code> ; in this case <code>astar</code> is changed to <code>1-alpha</code> .
<code>delta</code>	The standardized effect size for which the design is powered. Will be as input to <code>gsDesign()</code> unless it was input as <code>0</code> ; in that case, value will be computed to give desired power for fixed design with input sample size <code>n.fix</code> .
<code>n.fix</code>	Sample size required to obtain desired power when effect size is <code>delta</code> .
<code>timing</code>	A vector of length <code>k</code> containing the portion of the total planned information or sample size at each analysis.
<code>tol</code>	As input.
<code>r</code>	As input.
<code>n.I</code>	Vector of length <code>k</code> . If values are input, same values are output. Otherwise, <code>n.I</code> will contain the sample size required at each analysis to achieve desired <code>timing</code> and <code>beta</code> for the output value of <code>delta</code> . If <code>delta=0</code> was input, then this is the sample size required for the specified group sequential design when a fixed design requires a sample size of <code>n.fix</code> . If <code>delta=0</code> and <code>n.fix=1</code> then this is the relative sample size compared to a fixed design; see details and examples.
<code>maxn.IPlan</code>	As input.
<code>nFixSurv</code>	As input.
<code>nSurv</code>	Sample size for Lachin and Foulkes method when <code>nSurvival</code> is used for fixed design input. If <code>nSurvival</code> is used to compute <code>n.fix</code> , then <code>nFixSurv</code> is inflated by the same amount as <code>n.fix</code> and stored in <code>nSurv</code> . Note that if you use <code>gsSurv</code> for time-to-event sample size, this is not needed and a more complete output summary is given.
<code>endpoint</code>	As input.
<code>delta1</code>	As input.
<code>delta0</code>	As input.
<code>overrun</code>	As input.
<code>usTime</code>	As input.
<code>lsTime</code>	As input.

testUpper	Logical vector of length k indicating which analyses have an efficacy (upper) bound.
testLower	Logical vector of length k indicating which analyses have a futility (lower) bound.
testHarm	Logical vector of length k indicating which analyses have a harm bound (only for test.type 7 or 8).
upper	Upper bound spending function, boundary and boundary crossing probabilities under the NULL and alternate hypotheses. See vignette("SpendingFunctionOverview") and manual for further details.
lower	Lower bound spending function, boundary and boundary crossing probabilities at each analysis. Lower spending is under alternative hypothesis (beta spending) for test.type=3 or 4. For test.type=2, 5 or 6, lower spending is under the null hypothesis. For test.type=1, output value is NULL. See vignette("SpendingFunctionOverview") and manual.
theta	Standardized effect size under null (0) and alternate hypothesis. If delta is input, theta[1]=delta. If n.fix is input, theta[1] is computed using a standard sample size formula (pseudocode): $((Z_{\alpha} + Z_{\beta}) / \theta[1])^2 = n_{\text{fix}}$.
falseprobnb	For test.type=4 or 6, this contains false positive probabilities under the null hypothesis assuming that crossing a futility bound does not stop the trial.
en	Expected sample size accounting for early stopping. For time-to-event outcomes, this would be the expected number of events (although gsSurv will give expected sample size). For information-based-design, this would give the expected information when the trial stops. If overrun is specified, the expected sample size includes the overrun at each interim.

An object of class "xtable" with attributes specifying formatting options for a table

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall. Lan KK, DeMets DL (1989). Group sequential procedures: calendar versus information time. *Statistics in medicine* 8(10):1191-8. Liu, Q, Lim, P, Nuamah, I, and Li, Y (2012), On adaptive error spending approach for group sequential trials with random information levels. *Journal of biopharmaceutical statistics*; 22(4), 687-699.

See Also

vignette("gsDesignPackageOverview"), [gsBoundSummary](#), [plot.gsDesign](#), [gsProbability](#), vignette("SpendingFunctionOverview"), [Normal xtable](#)

Examples

```

library(ggplot2)
# symmetric, 2-sided design with O'Brien-Fleming-like boundaries
# lower bound is non-binding (ignored in Type I error computation)
# sample size is computed based on a fixed design requiring n=800
x <- gsDesign(k = 5, test.type = 2, n.fix = 800)

# note that "x" below is equivalent to print(x) and print.gsDesign(x)
x
plot(x)
plot(x, plotype = 2)

# Assuming after trial was designed actual analyses occurred after
# 300, 600, and 860 patients, reset bounds
y <- gsDesign(
  k = 3, test.type = 2, n.fix = 800, n.I = c(300, 600, 860),
  maxn.IPlan = x$n.I[x$k]
)
y

# asymmetric design with user-specified spending that is non-binding
# sample size is computed relative to a fixed design with n=1000
sfup <- c(.033333, .063367, .1)
sflp <- c(.25, .5, .75)
timing <- c(.1, .4, .7)
x <- gsDesign(
  k = 4, timing = timing, sfu = sfPoints, sfupar = sfup, sfl = sfPoints,
  sflpar = sflp, n.fix = 1000
)
x
plot(x)
plot(x, plotype = 2)

# same design, but with relative sample sizes
gsDesign(
  k = 4, timing = timing, sfu = sfPoints, sfupar = sfup, sfl = sfPoints,
  sflpar = sflp
)

```

Description

Computes power/Type I error and expected sample size for a group sequential design across a selected set of parameter values for a given set of analyses and boundaries. The print function has been extended using `print.gsProbability` to print `gsProbability` objects; see examples.

Depending on the calling sequence, an object of class `gsProbability` or class `gsDesign` is returned. If it is of class `gsDesign` then the members of the object will be the same as described in

gsDesign. If *d* is input as NULL (the default), all other arguments (other than *r*) must be specified and an object of class *gsProbability* is returned. If *d* is passed as an object of class *gsProbability* or *gsDesign* the only other argument required is *theta*; the object returned has the same class as the input *d*. On output, the values of *theta* input to *gsProbability* will be the parameter values for which the design is characterized.

Usage

```
gsProbability(k = 0, theta, n.I, a, b, r = 18, d = NULL, overrun = 0)
```

```
## S3 method for class 'gsProbability'
print(x, ...)
```

Arguments

<i>k</i>	Number of analyses planned, including interim and final.
<i>theta</i>	Vector of standardized effect sizes for which boundary crossing probabilities are to be computed.
<i>n.I</i>	Sample size or relative sample size at analyses; vector of length <i>k</i> . See gsDesign and manual.
<i>a</i>	Lower bound cutoffs (z-values) for futility or harm at each analysis, vector of length <i>k</i> .
<i>b</i>	Upper bound cutoffs (z-values) for futility at each analysis; vector of length <i>k</i> .
<i>r</i>	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.
<i>d</i>	If not NULL, this should be an object of type <i>gsDesign</i> returned by a call to <i>gsDesign()</i> . When this is specified, the values of <i>k</i> , <i>n.I</i> , <i>a</i> , <i>b</i> , and <i>r</i> will be obtained from <i>d</i> and only <i>theta</i> needs to be specified by the user.
<i>overrun</i>	Scalar or vector of length <i>k</i> -1 with patients enrolled that are not included in each interim analysis.
<i>x</i>	An item of class <i>gsProbability</i> .
...	Not implemented (here for compatibility with generic print input).

Value

<i>k</i>	As input.
<i>theta</i>	As input.
<i>n.I</i>	As input.
<i>lower</i>	A list containing two elements: <i>bound</i> is as input in <i>a</i> and <i>prob</i> is a matrix of boundary crossing probabilities. Element <i>i, j</i> contains the boundary crossing probability at analysis <i>i</i> for the <i>j</i> -th element of <i>theta</i> input. All boundary crossing is assumed to be binding for this computation; that is, the trial must stop if a boundary is crossed.

upper	A list of the same form as lower containing the upper bound and upper boundary crossing probabilities.
en	A vector of the same length as theta containing expected sample sizes for the trial design corresponding to each value in the vector theta.
r	As input.

Note: `print.gsProbability()` returns the input `x`.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

[plot.gsDesign](#), [gsDesign](#), [vignette\("gsDesignPackageOverview"\)](#)

Examples

```
library(ggplot2)
# making a gsDesign object first may be easiest...
x <- gsDesign()

# take a look at it
x

# default plot for gsDesign object shows boundaries
plot(x)

# \code{plottype=2} shows boundary crossing probabilities
plot(x, plottype = 2)

# now add boundary crossing probabilities and
# expected sample size for more theta values
y <- gsProbability(d = x, theta = x$delta * seq(0, 2, .25))
class(y)

# note that "y" below is equivalent to \code{print(y)} and
# \code{print.gsProbability(y)}
y

# the plot does not change from before since this is a
# gsDesign object; note that theta/delta is on x axis
plot(y, plottype = 2)
```

```
# now let's see what happens with a gsProbability object
z <- gsProbability(
  k = 3, a = x$lower$bound, b = x$upper$bound,
  n.I = x$n.I, theta = x$delta * seq(0, 2, .25)
)

# with the above form, the results is a gsProbability object
class(z)
z

# default plottype is now 2
# this is the same range for theta, but plot now has theta on x axis
plot(z)
```

gsSurvCalendar

Group sequential design with calendar-based timing of analyses

Description

This is like `gsSurv()`, but the timing of analyses is specified in calendar time units. Information fraction is computed from the input rates and the calendar times. Spending can be based on information fraction as in Lan and DeMets (1983) or calendar time units as in Lan and DeMets (1989).

Usage

```
gsSurvCalendar(
  test.type = 4,
  alpha = 0.025,
  sided = 1,
  beta = 0.1,
  astar = 0,
  sfu = gsDesign::sfHSD,
  sfupar = -4,
  sfl = gsDesign::sfHSD,
  sflpar = -2,
  sfharm = gsDesign::sfHSD,
  sfharmparam = -2,
  calendarTime = c(12, 24, 36),
  spending = c("information", "calendar"),
  lambdaC = log(2)/6,
  hr = 0.6,
  hr0 = 1,
  eta = 0,
  etaE = NULL,
  gamma = 1,
  R = 12,
  S = NULL,
```

```

minfup = 18,
ratio = 1,
r = 18,
tol = .Machine$double.eps^0.25,
testUpper = TRUE,
testLower = TRUE,
testHarm = TRUE,
method = c("LachinFoulkes", "Schoenfeld", "Freedman", "BernsteinLagakos")
)

```

Arguments

test.type	1=one-sided 2=two-sided symmetric 3=two-sided, asymmetric, beta-spending with binding lower bound 4=two-sided, asymmetric, beta-spending with non-binding lower bound 5=two-sided, asymmetric, lower bound spending under the null hypothesis with binding lower bound 6=two-sided, asymmetric, lower bound spending under the null hypothesis with non-binding lower bound 7=two-sided, asymmetric, with binding futility and binding harm bounds 8=two-sided, asymmetric, with non-binding futility and non-binding harm bounds. See details, examples and manual.
alpha	Type I error rate. Default is 0.025 since 1-sided testing is default.
sided	1 for 1-sided testing, 2 for 2-sided testing.
beta	Type II error rate. Default is 0.10 (90% power); NULL if power is to be computed based on other input values.
astar	Total spending for the lower (test.type 5 or 6) or harm (test.type 7 or 8) bound under the null hypothesis. Default is 0. For test.type 5 or 6, astar specifies the total probability of crossing a lower bound at all analyses combined. For test.type 7 or 8, astar specifies the total probability of crossing the harm bound at all analyses combined under the null hypothesis. If astar = 0, it will be changed to 1−alpha.
sfu	A spending function or a character string indicating a boundary type (that is, “WT” for Wang-Tsiatis bounds, “OF” for O’Brien-Fleming bounds and “Pocock” for Pocock bounds). For one-sided and symmetric two-sided testing is used to completely specify spending (test.type=1, 2), sfu. The default value is sfHSD which is a Hwang-Shih-DeCani spending function. See details, vignette("SpendingFunctionOverview") manual and examples.
sfupar	Real value, default is −4 which is an O’Brien-Fleming-like conservative bound when used with the default Hwang-Shih-DeCani spending function. This is a real-vector for many spending functions. The parameter sfupar specifies any parameters needed for the spending function specified by sfu; this is not needed for spending functions (sfLDOF, sfLDPocock) or bound types (“OF”, “Pocock”) that do not require parameters. Note that sfupar can be specified as a positive scalar for sfLDOF for a generalized O’Brien-Fleming spending function.

sf1	Specifies the spending function for lower boundary crossing probabilities when asymmetric, two-sided testing is performed (<code>test.type = 3, 4, 5, or 6</code>). Unlike the upper bound, only spending functions are used to specify the lower bound. The default value is <code>sfHSD</code> which is a Hwang-Shih-DeCani spending function. The parameter <code>sf1</code> is ignored for one-sided testing (<code>test.type=1</code>) or symmetric 2-sided testing (<code>test.type=2</code>). See details, spending functions, manual and examples.
sf1par	Real value, default is -2 , which, with the default Hwang-Shih-DeCani spending function, specifies a less conservative spending rate than the default for the upper bound.
sfharm	A spending function for the harm bound, used with <code>test.type = 7</code> or <code>test.type = 8</code> . Default is <code>sfHSD</code> . See spendingFunction for details.
sfharmparam	Real value, default is -2 . Parameter for the harm bound spending function <code>sfharm</code> .
calendarTime	Vector of increasing positive numbers with calendar times of analyses. Time 0 is start of randomization.
spending	Select between calendar-based spending and information-based spending.
lambdaC	Scalar, vector or matrix of event hazard rates for the control group; rows represent time periods while columns represent strata; a vector implies a single stratum. Note that rates corresponding the final time period are extended indefinitely.
hr	Hazard ratio (experimental/control) under the alternate hypothesis (scalar, > 0 , must differ from <code>hr0</code>). Both $hr < hr0$ (experimental is beneficial when lower hazard is better) and $hr > hr0$ (e.g., time-to-response or safety designs) are supported.
hr0	Hazard ratio (experimental/control) under the null hypothesis (scalar, > 0 , must differ from <code>hr</code>).
eta	Scalar, vector or matrix of dropout hazard rates for the control group; rows represent time periods while columns represent strata; if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
etaE	Matrix dropout hazard rates for the experimental group specified in like form as <code>eta</code> ; if NULL, this is set equal to <code>eta</code> .
gamma	A scalar, vector or matrix of rates of entry by time period (rows) and strata (columns); if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
R	A scalar or vector of durations of time periods for recruitment rates specified in rows of <code>gamma</code> . Length is the same as number of rows in <code>gamma</code> . Note that when variable enrollment duration is specified (input <code>T = NULL</code>), the final enrollment period is extended as long as needed.
S	A scalar or vector of durations of piecewise constant event rates specified in rows of <code>lambda</code> , <code>eta</code> and <code>etaE</code> ; this is NULL if there is a single event rate per stratum (exponential failure) or length of the number of rows in <code>lambda</code> minus 1, otherwise. The final time period is extended indefinitely for each stratum.

minfup	A non-negative scalar less than the maximum value in calendarTime. Enrollment will be cut off at the difference between the maximum value in calendarTime and minfup.
ratio	Randomization ratio of experimental treatment divided by control; normally a scalar, but may be a vector with length equal to number of strata.
r	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.
tol	Tolerance for error passed to the gsDesign function.
testUpper	Indicator of which analyses should include an upper (efficacy) bound. A single value of TRUE (default) indicates all analyses have an efficacy bound. Otherwise, a logical vector of length k indicating which analyses will have an efficacy bound. Overridden to all TRUE for test.type 1 and 2. Must be TRUE at the final analysis to achieve targeted power. At each analysis, at least one of testUpper, testLower, or testHarm must be TRUE. Where testUpper is FALSE, the upper bound is set to $+2\theta$ (effectively Inf) and displayed as NA in output.
testLower	Indicator of which analyses should include a lower (futility) bound. A single value of TRUE (default) indicates all analyses have a lower bound; FALSE indicates none. Otherwise, a logical vector of length k. Ignored for test.type 1 (one-sided, no lower bound). Overridden to all TRUE for test.type 2 (symmetric). For test.type 3–8, at least one analysis must be TRUE. Where testLower is FALSE, the lower bound is set to -2θ (effectively $-\text{Inf}$) and displayed as NA in output.
testHarm	Indicator of which analyses should include a harm bound. A single value of TRUE (default) indicates all analyses have a harm bound; FALSE indicates none. Otherwise, a logical vector of length k. Only used for test.type 7 or 8; at least one analysis must be TRUE for those types. Where testHarm is FALSE, the harm bound is set to -2θ (effectively $-\text{Inf}$) and displayed as NA in output.
method	One of "LachinFoulkes" (default), "Schoenfeld", "Freedman", or "BernsteinLagakos". Note: "Schoenfeld" and "Freedman" methods only support superiority testing ($hr_0 = 1$). "Freedman" does not support stratified populations.

Value

An object of class `c("gsSurv", "gsDesign")` with group sequential boundaries and expected counts at each calendar analysis time. See [gsSurv](#) for full component details.

References

- Lan KKG and DeMets DL (1983), Discrete Sequential Boundaries for Clinical Trials. *Biometrika*, 70, 659-663.
- Lan KKG and DeMets DL (1989), Group Sequential Procedures: Calendar vs. Information Time. *Statistics in Medicine*, 8, 1191-1198.

Schoenfeld D (1981), The Asymptotic Properties of Nonparametric Tests for Comparing Survival Distributions. *Biometrika*, 68, 316-319.

Freedman LS (1982), Tables of the Number of Patients Required in Clinical Trials Using the Logrank Test. *Statistics in Medicine*, 1, 121-129.

See Also

`vignette("SeqDesignSurvival", package = "gsDesign")` for a SAS PROC SEQDESIGN sample size translation example and `vignette("gsSurvPower", package = "gsDesign")` for power calculations with fixed calendar analysis assumptions.

[gsSurv](#), [gsSurvPower](#), [gsDesign](#), [gsBoundSummary](#)

Examples

```
# First example: while timing is calendar-based, spending is event-based
x <- gsSurvCalendar() |> toInteger()
gsBoundSummary(x)

# Second example: both timing and spending are calendar-based
# This results in less spending at interims and leaves more for final analysis
y <- gsSurvCalendar(spending = "calendar") |> toInteger()
gsBoundSummary(y)

# Note that calendar timing for spending relates to planned timing for y
# rather than timing in y after toInteger() conversion

# Values plugged into spending function for calendar time
y$usTime
# Actual calendar fraction from design after toInteger() conversion
y$T / max(y$T)
```

gsSurvPower

Compute power for a group sequential survival design

Description

`gsSurvPower()` computes power for a group sequential survival design with specified enrollment, dropout, treatment effect, and analysis timing. Unlike `gsSurv()` and `gsSurvCalendar()` which solve for sample size to achieve target power, `gsSurvPower()` takes fixed design assumptions and computes the resulting power. It is meant to compute for a single set of assumptions at a time; different scenarios are evaluated with separate calls.

Usage

```
gsSurvPower(
  x = NULL,
  k = NULL,
  test.type = NULL,
```

```

alpha = NULL,
sided = NULL,
astar = NULL,
sfu = NULL,
sfupar = NULL,
sfl = NULL,
sflpar = NULL,
sfharm = NULL,
sfharmparam = NULL,
r = NULL,
usTime = NULL,
lsTime = NULL,
testUpper = NULL,
testLower = NULL,
testHarm = NULL,
lambdaC = NULL,
hr = NULL,
hr0 = NULL,
hr1 = NULL,
eta = NULL,
etaE = NULL,
gamma = NULL,
R = NULL,
targetN = NULL,
S = NULL,
ratio = NULL,
minfup = NULL,
method = NULL,
spending = c("information", "calendar"),
plannedCalendarTime = NULL,
targetEvents = NULL,
maxExtension = NULL,
minTimeFromPreviousAnalysis = NULL,
minN = NULL,
minFollowUp = NULL,
informationRates = NULL,
fullSpendingAtFinal = FALSE,
tol = .Machine$double.eps^0.25
)

```

Arguments

x	Optional gsSurv or gsSurvCalendar object providing defaults for all parameters. When provided, any user-specified parameter overrides the corresponding value from x.
k	Number of analyses planned, including interim and final.
test.type	1 = one-sided, 2 = two-sided symmetric, 3 = two-sided, asymmetric, beta-spending with binding lower bound, 4 = two-sided, asymmetric, beta-spending

	with non-binding lower bound, 5 = two-sided, asymmetric, lower bound spending under the null hypothesis with binding lower bound, 6 = two-sided, asymmetric, lower bound spending under the null hypothesis with non-binding lower bound, 7 = two-sided, asymmetric, with binding futility and binding harm bounds, 8 = two-sided, asymmetric, with non-binding futility and non-binding harm bounds.
alpha	Type I error rate. Default is 0.025 since 1-sided testing is default. Internally divided by sided before passing to gsDesign(), matching the convention used by gsSurv() and gsSurvCalendar().
sided	1 for 1-sided, 2 for 2-sided testing. Used to convert alpha to one-sided via α / sided for internal calculations, matching the convention of gsSurv() and nSurv(). When x is provided and sided is omitted, gsSurvPower() reuses the stored sided value from the design call when available.
astar	Lower bound total crossing probability for test.type 5 or 6. Default 0.
sfu	Upper bound spending function (default sfHSD).
sfupar	Parameter for sfu (default -4).
sfl	Lower bound spending function (default sfHSD).
sflpar	Parameter for sfl (default -2).
sfharm	Spending function for the harm bound, used with test.type = 7 or test.type = 8. Default sfHSD.
sfharmparam	Real value, default -2. Parameter for the harm bound spending function sfharm.
r	Integer grid parameter for numerical integration (default 18).
usTime	Upper spending time override; vector of length k or NULL (default) to use information fractions. Ignored when spending = "calendar", because realized analysis times determine the spending fractions.
lsTime	Lower spending time override; vector of length k or NULL (default) to use information fractions. Ignored when spending = "calendar".
testUpper	Indicator of which analyses include an efficacy test. TRUE (default) for all analyses. A logical vector of length k may be specified.
testLower	Indicator of which analyses include a futility test. TRUE (default) for all analyses. A logical vector of length k may be specified.
testHarm	Indicator of which analyses include a harm bound. TRUE (default) for all analyses. A logical vector of length k may be specified. Only used for test.type 7 or 8.
lambdaC	Scalar, vector, or matrix of control event hazard rates. Rows = time periods, columns = strata.
hr	Assumed hazard ratio (experimental/control) for power computation. This is the "what-if" treatment effect.
hr0	Null hazard ratio. Set $\text{hr0} > 1$ for non-inferiority.
hr1	Design alternative hazard ratio used to calibrate futility bounds (test.type 3, 4, 7, 8 only; not used for 5, 6 or harm bounds). Defaults to x\$hr when x is provided, otherwise hr.

eta	Scalar, vector, or matrix of control dropout hazard rates.
etaE	Experimental dropout hazard rates; if NULL, set to eta.
gamma	Scalar, vector, or matrix of enrollment rates by period (rows) and strata (columns).
R	Scalar or vector of enrollment period durations.
targetN	Target total sample size. When specified, R is uniformly rescaled so that $\text{sum}(\text{gamma} * R) == \text{targetN}$, preserving the relative duration of each enrollment period. This is a convenience for "what-if" analyses where the enrollment rate changes but the target sample size stays the same (or vice versa). Cannot be used together with an explicit R.
S	Scalar or vector of piecewise failure period durations; NULL for exponential failure.
ratio	Randomization ratio (experimental/control). Default 1.
minfup	Minimum follow-up time.
method	Sample-size variance formulation. One of "LachinFoulkes" (default), "Schoenfeld", "Freedman", or "BernsteinLagakos". Affects n.fix computation when x is not provided.
spending	One of "information" (default) or "calendar". Controls whether alpha/beta spending tracks information fractions or calendar time fractions ($T / \max(T)$). With calendar spending, usTime and lsTime are derived from the realized analysis times and any user-supplied overrides are ignored.
plannedCalendarTime	Calendar times for analyses (time 0 = start of randomization). Scalar (recycled) or vector of length k. Use NA for analyses not determined by calendar time.
targetEvents	Target number of events at each analysis. Scalar (recycled), vector of length k (overall targets), or matrix with k rows and nstrata columns (per-stratum targets). Use NA for analyses not determined by events. When a matrix is supplied, row sums give the total event target used to solve each analysis time.
maxExtension	Maximum time extension beyond the floor time to wait for targetEvents. Scalar or vector of length k.
minTimeFromPreviousAnalysis	Minimum elapsed time since the previous analysis. Scalar or vector of length k. Ignored for the first analysis.
minN	Minimum total sample size enrolled before analysis can proceed. Scalar or vector of length k.
minFollowUp	Minimum follow-up time after minN is reached. Scalar or vector of length k. Must be ≥ 0 .
informationRates	Numeric vector of length k specifying planned information fractions. When provided, spending fractions are $\text{pmin}(\text{informationRates}, \text{actual_timing})$ at each analysis, where actual_timing is expected events divided by maximum expected events. This prevents over-spending when events are ahead of schedule and under-spends when behind. When supplied, these planned-vs-actual information fractions take precedence over spending, usTime, and lsTime; both upper and lower spending times use the same capped vector. Default NULL uses actual information fractions (or calendar fractions when spending = "calendar").

fullSpendingAtFinal	Logical. When TRUE, the spending fraction at the final analysis is forced to 1 after applying informationRates, calendar spending, or user-supplied usTime/lstime. This ensures full alpha spending whenever the selected spending-time vector would otherwise end below 1. Default FALSE.
tol	Tolerance for <code>uniroot</code> when solving for analysis times.

Details

Accepting a gsSurv object: An optional `gsSurv`-class object `x` provides defaults for all parameters. This includes output from `gsSurv()` and `gsSurvCalendar()`. User-specified parameters override these defaults, enabling "what-if" analyses: e.g., `gsSurvPower(x = design, hr = 0.8)` evaluates power under HR = 0.8 using all other parameters from the design. When `x` is not provided, all design parameters must be specified directly.

Hazard ratio roles: Two distinct hazard ratios serve different purposes. `hr` is the assumed treatment effect under which power is evaluated. `hr1` is the design alternative used to calibrate futility bounds (for `test.type` 3, 4, 7, 8). It is not used for `test.type` 5 or 6 (which use H0 spending for the lower bound) or for harm bounds. When `x` is provided, `hr1` defaults to `x$hr`, so futility bounds remain calibrated to the original design even when power is evaluated under a different `hr`.

Analysis timing: Analysis times are determined by per-analysis criteria. Each timing parameter can be a scalar (recycled to all `k` analyses), a vector of length `k`, or NA at position `i` to indicate the criterion does not apply to analysis `i`.

The choice between `plannedCalendarTime` and `targetEvents` has an important consequence for sensitivity analyses:

- `plannedCalendarTime` fixes calendar times; expected events are recomputed under the assumed HR. A worse HR produces more events at the same calendar time (the experimental arm fails faster). This gives an "unconditional" power.
- `targetEvents` fixes event counts; calendar times adjust. Since events are held constant, information fractions do not change with HR, and results match the `gsDesign` power plot (`plot(x, plottype = 2)`) to numerical precision.

How criteria combine within a single analysis: For analysis `i`, the analysis time `T[i]` is determined as:

1. Compute floor times from applicable criteria: `plannedCalendarTime[i]`, `T[i-1] + minTimeFromPreviousAnalysis` and time when `minN[i]` enrolled + `minFollowUp[i]`.
2. `floor_time = max(all applicable floor times)`.
3. If `targetEvents[i]` is specified: find `t_events` when expected events reach target. If `t_events <= floor_time`, analysis at `floor_time`. If `t_events > floor_time` and `maxExtension[i]` is set, analysis at `min(t_events, floor_time + maxExtension[i])`. Otherwise, analysis at `t_events`.
4. If no `targetEvents`: analysis at `floor_time`.
5. `maxExtension` is a hard cap: the analysis time is never pushed beyond `plannedCalendarTime[i] + maxExtension[i]` (or `T[i-1] + maxExtension[i]` when no calendar time is specified), even if other criteria such as `minTimeFromPreviousAnalysis` or `minN + minFollowUp` would require a later time.

Normalization and consistency: When x is provided, $x\$n.fix$ is used for the `gsDesign::gsDesign()` call to ensure the internal drift parameter θ and bounds match the original design exactly. The assumed HR's drift is obtained by scaling: $\theta_{\text{assumed}} = \theta_{\text{design}} \times |\log(hr/hr_0)|/|\log(hr_1/hr_0)|$. Power is computed via `gsDesign::gsProbability()` with actual expected events as $n.I$. At the design HR, this reproduces the design power exactly.

Stratified targetEvents: `targetEvents` accepts a scalar (recycled), a vector of length k (overall targets per analysis), or a matrix with k rows and `nstrata` columns (per-stratum targets). A vector of length k is always interpreted as overall targets; use a matrix for per-stratum specification.

Bound recalculation when parameters change: When x is provided, the handling of bounds depends on which parameters change relative to the original design:

- **No bound parameters changed** (same α , s_{fu} , s_{fupar}) and timing matches: both bounds are reused from x exactly.
- **Upper-bound parameters changed** (α , s_{fu} , or s_{fupar}) but timing matches: new efficacy bounds are computed via `gsDesign(test.type = 1)` at the new α , while the original futility bounds from x are preserved. Any futility bound that exceeds the new efficacy bound is clipped. This follows the same convention as `gsBoundSummary()`. Lower-bound spending settings from x are intentionally kept in this branch, which avoids complications with `astar` validation for binding types.
- **Timing changed** (different target events or calendar times): both bounds are recomputed from scratch using the full `test.type` and all spending parameters.

Value

An object of class `c("gsSurv", "gsDesign")` containing:

<code>k</code>	Number of analyses.
<code>n.I</code>	Total expected events at each analysis.
<code>timing</code>	Information fractions at each analysis.
<code>T</code>	Calendar times of analyses.
<code>eDC, eDE</code>	Expected events by stratum (control, experimental).
<code>eNC, eNE</code>	Expected sample sizes by stratum (control, experimental).
<code>upper, lower</code>	Bounds and crossing probabilities.
<code>harm</code>	Harm-bound information when <code>test.type</code> is 7 or 8.
<code>en, theta</code>	Expected sample size summary and drift values returned by <code>gsDesign::gsProbability()</code> .
<code>hr, hr0, hr1</code>	Assumed, null, and design hazard ratios.
<code>power</code>	Overall power (sum of upper-bound crossing probabilities under the assumed HR).
<code>beta</code>	Type II error ($1 - \text{power}$).
<code>variable</code>	Always "Power".
<code>test.type, alpha, sided, method, spending, call</code>	Design settings used for the power calculation.
<code>testUpper, testLower, testHarm</code>	Logical indicators of which analyses include each bound type, when relevant.
<code>lambdaC, etaC, etaE, gamma, R, S, ratio, minfup</code>	Rate and timing inputs used in the calculation.

See Also

`vignette("gsSurvPower", package = "gsDesign")` for worked examples including calendar spending, stratified event targets, and biomarker subgroup analyses.

`vignette("gsSurvBasicExamples", package = "gsDesign")` for deriving survival sample size designs and `vignette("SeqDesignSurvival", package = "gsDesign")` for reproducing SAS PROC SEQDESIGN survival output.

[gsSurv](#), [gsSurvCalendar](#), [gsDesign](#), [gsProbability](#)

Examples

```
# Create a design, then evaluate power at the design HR
design <- gsSurv(
  k = 3, test.type = 4, alpha = 0.025, sided = 1, beta = 0.1,
  lambdaC = log(2) / 12, hr = 0.7, eta = 0.01,
  gamma = 10, R = 16, minfup = 12, T = 28
)
pwr <- gsSurvPower(x = design, plannedCalendarTime = design$T)
pwr$power # should be 0.9

# Power under a worse HR
gsSurvPower(x = design, hr = 0.8, plannedCalendarTime = design$T)$power

# Event-driven timing (matches gsDesign power plot)
design_events <- design$n.I
gsSurvPower(x = design, hr = 0.8, targetEvents = design_events)$power

# Without a reference design
gsSurvPower(
  k = 2, test.type = 4, alpha = 0.025, sided = 1,
  lambdaC = log(2) / 6, hr = 0.65, eta = 0.01,
  gamma = 8, R = 18, ratio = 1,
  plannedCalendarTime = c(24, 36)
)$power
```

nNormal

Normal distribution sample size (2-sample)

Description

`nNormal()` computes a fixed design sample size for comparing 2 means where variance is known. The function allows computation of sample size for a non-inferiority hypothesis. Note that you may wish to investigate other R packages such as the `pwr` package which uses the t-distribution. In the examples below we show how to set up a 2-arm group sequential design with a normal outcome.

`nNormal()` computes sample size for comparing two normal means when the variance for observations in

Usage

```
nNormal(
  delta1 = 1,
  sd = 1.7,
  sd2 = NULL,
  alpha = 0.025,
  beta = 0.1,
  ratio = 1,
  sided = 1,
  n = NULL,
  delta0 = 0,
  outtype = 1
)
```

Arguments

delta1	difference between sample means under the alternate hypothesis.
sd	Standard deviation for the control arm.
sd2	Standard deviation of experimental arm; this will be set to be the same as the control arm with the default of NULL.
alpha	type I error rate. Default is 0.025 since 1-sided testing is default.
beta	type II error rate. Default is 0.10 (90% power). Not needed if n is provided.
ratio	randomization ratio of experimental group compared to control.
sided	1 for 1-sided test (default), 2 for 2-sided test.
n	Sample size; may be input to compute power rather than sample size. If NULL (default) then sample size is computed.
delta0	difference between sample means under the null hypothesis; normally this will be left as the default of 0.
outtype	controls output; see value section below.

Details

This is more of a convenience routine than one recommended for broad use without careful considerations such as those outlined in Jennison and Turnbull (2000). For larger studies where a conservative estimate of within group standard deviations is available, it can be useful. A more detailed formulation is available in the vignette on two-sample normal sample size.

Value

If n is NULL (default), total sample size (2 arms combined) is computed. Otherwise, power is computed. If outtype=1 (default), the computed value (sample size or power) is returned in a scalar or vector. If outtype=2, a data frame with sample sizes for each arm (n1, n2) is returned; if n is not input as NULL, a third variable, Power, is added to the output data frame. If outtype=3, a data frame with is returned with the following columns:

n	A vector with total samples size required for each event rate comparison specified
---	--

n1	A vector of sample sizes for group 1 for each event rate comparison specified
n2	A vector of sample sizes for group 2 for each event rate comparison specified
alpha	As input
sided	As input
beta	As input; if n is input, this is computed
Power	If n=NULL on input, this is 1-beta; otherwise, the power is computed for each sample size input
sd	As input
sd2	As input
delta1	As input
delta0	As input
se	standard error for estimate of difference in treatment group means

Author(s)

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Lachin JM (1981), Introduction to sample size determination and power analysis for clinical trials. *Controlled Clinical Trials* 2:93-113.

Snedecor GW and Cochran WG (1989), *Statistical Methods*. 8th ed. Ames, IA: Iowa State University Press.

See Also

See vignette("nNormal") for the full story, including the derivation of the sample size formula, power checks via simulation, and a group sequential design example.

Examples

```
# EXAMPLES
# equal variances
n=nNormal(delta1=.5,sd=1.1,alpha=.025,beta=.2)
n
x <- gsDesign(k = 3, n.fix = n, test.type = 4, alpha = 0.025, beta = 0.1, timing = c(.5,.75),
sfu = sfLDOF, sfl = sfHSD, sflpar = -1, delta1 = 0.5, endpoint = 'normal')
gsBoundSummary(x)
summary(x)
# unequal variances, fixed design
nNormal(delta1 = .5, sd = 1.1, sd2 = 2, alpha = .025, beta = .2)
# unequal sample sizes
nNormal(delta1 = .5, sd = 1.1, alpha = .025, beta = .2, ratio = 2)
# non-inferiority assuming a better effect than null
nNormal(delta1 = .5, delta0 = -.1, sd = 1.2)
```

normalGrid	<i>Normal Density Grid</i>
------------	----------------------------

Description

normalGrid() is intended to be used for computation of the expected value of a function of a normal random variable. The function produces grid points and weights to be used for numerical integration.

This is a utility function to provide a normal density function and a grid to integrate over as described by Jennison and Turnbull (2000), Chapter 19. While integration can be performed over the real line or over any portion of it, the numerical integration does not extend beyond 6 standard deviations from the mean. The grid used for integration uses equally spaced points over the middle of the distribution function, and spreads points further apart in the tails. The values returned in gridwghts may be used to integrate any function over the given grid, although the user should take care that the function integrated is not large in the tails of the grid where points are spread further apart.

Usage

```
normalGrid(r = 18, bounds = c(0, 0), mu = 0, sigma = 1)
```

Arguments

r	Control for grid points as in Jennison and Turnbull (2000), Chapter 19; default is 18. Range: 1 to 80. This might be changed by the user (e.g., r=6 which produces 65 gridpoints compare to 185 points when r=18) when speed is more important than precision.
bounds	Range of integration. Real-valued vector of length 2. Default value of 0, 0 produces a range of + or - 6 standard deviations (6*sigma) from the mean (=mu).
mu	Mean of the desired normal distribution.
sigma	Standard deviation of the desired normal distribution.

Value

z	Grid points for numerical integration.
density	The standard normal density function evaluated at the values in z; see examples.
gridwghts	Simpson's rule weights for numerical integration on the grid in z; see examples.
wghts	Weights to be used with the grid in z for integrating the normal density function; see examples. This is equal to density * gridwghts.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Examples

```
library(ggplot2)
# standard normal distribution
x <- normalGrid(r = 3)
plot(x$z, x$wgts)

# verify that numerical integration replicates sigma
# get grid points and weights
x <- normalGrid(mu = 2, sigma = 3)

# compute squared deviation from mean for grid points
dev <- (x$z - 2)^2

# multiply squared deviations by integration weights and sum
sigma2 <- sum(dev * x$wgts)

# square root of sigma2 should be sigma (3)
sqrt(sigma2)

# do it again with larger r to increase accuracy
x <- normalGrid(r = 22, mu = 2, sigma = 3)
sqrt(sum((x$z - 2)^2 * x$wgts))

# this can also be done by combining gridwgts and density
sqrt(sum((x$z - 2)^2 * x$gridwgts * x$density))

# integrate normal density and compare to built-in function
# to compute probability of being within 1 standard deviation
# of the mean
pnorm(1) - pnorm(-1)
x <- normalGrid(bounds = c(-1, 1))
sum(x$wgts)
sum(x$gridwgts * x$density)

# find expected sample size for default design with
# n.fix=1000
x <- gsDesign(n.fix = 1000)
x

# set a prior distribution for theta
y <- normalGrid(r = 3, mu = x$theta[2], sigma = x$theta[2] / 1.5)
z <- gsProbability(
  k = 3, theta = y$z, n.I = x$n.I, a = x$lower$bound,
  b = x$upper$bound
)
z <- gsProbability(d = x, theta = y$z)
cat(
```

```

      "Expected sample size averaged over normal\n prior distribution for theta with \n mu=",
      x$theta[2], "sigma=", x$theta[2] / 1.5, ":",
      round(sum(z$en * y$wgt), 1), "\n"
    )
  plot(y$z, z$en,
       xlab = "theta", ylab = "E{N}",
       main = "Expected sample size for different theta values"
  )
  lines(y$z, z$en)

```

plot.gsDesign

Plots for group sequential designs

Description

The `plot()` function has been extended to work with objects returned by `gsDesign()` and `gsProbability()`. For objects of type `gsDesign`, seven types of plots are provided: z-values at boundaries (default), power, approximate treatment effects at boundaries, conditional power at boundaries, spending functions, expected sample size, and B-values at boundaries. For objects of type `gsProbability` plots are available for z-values at boundaries, power (default), approximate treatment effects at boundaries, conditional power, expected sample size and B-values at boundaries.

The intent is that many standard `plot()` parameters will function as expected; exceptions to this rule exist. In particular, `main`, `xlab`, `ylab`, `lty`, `col`, `lwd`, `type`, `pch`, `cex` have been tested and work for most values of `plottype`; one exception is that `type="l"` cannot be overridden when `plottype=2`. Default values for labels depend on `plottype` and the class of `x`.

Note that there is some special behavior for values plotted and returned for power and expected sample size (ASN) plots for a `gsDesign` object. A call to `x<-gsDesign()` produces power and expected sample size for only two theta values: 0 and `x$delta`. The call `plot(x,plottype="Power")` (or `plot(x,plottype="ASN")` for a `gsDesign` object produces power (expected sample size) curves and returns a `gsDesign` object with theta values determined as follows. If theta is non-null on input, the input value(s) are used. Otherwise, for a `gsProbability` object, the theta values from that object are used. For a `gsDesign` object where theta is input as `NULL` (the default), `theta=seq(0,2,.05)*x$delta` is used. For a `gsDesign` object, the x-axis values are rescaled to `theta/x$delta` and the label for the x-axis θ/δ . For a `gsProbability` object, the values of theta are plotted and are labeled as θ . See examples below.

Approximate treatment effects at boundaries are computed dividing the Z-values at the boundaries by the square root of `n.I` at that analysis.

Spending functions are plotted for a continuous set of values from 0 to 1. This option should not be used if a boundary is used or a pointwise spending function is used (`sfu` or `sf1="WT"`, `"OF"`, `"Pocock"` or `sfPoints`).

Conditional power is computed using the function `gsBoundCP()`. The default input for this routine is `theta="thetahat"` which will compute the conditional power at each bound using the approximate treatment effect at that bound. Otherwise, if the input is `gsDesign` object conditional power is computed assuming `theta=x$delta`, the original effect size for which the trial was planned.

Average sample number/expected sample size is computed using $n.I$ at each analysis times the probability of crossing a boundary at that analysis. If no boundary is crossed at any analysis, this is counted as stopping at the final analysis.

B-values are Z-values multiplied by $\sqrt{t} = \sqrt{x\$n.I / x\$n.I[x\$k]}$. Thus, the expected value of a B-value at an analysis is the true value of θ multiplied by the proportion of total planned observations at that time. See Proschan, Lan and Wittes (2006).

Usage

```
## S3 method for class 'gsDesign'
plot(x, plottype = 1, base = FALSE, ...)

## S3 method for class 'gsProbability'
plot(x, plottype = 2, base = FALSE, ...)
```

Arguments

x	Object of class <code>gsDesign</code> for <code>plot.gsDesign()</code> or <code>gsProbability</code> for <code>plot.gsProbability()</code> .
plottype	1=boundary plot (default for <code>gsDesign</code>), 2=power plot (default for <code>gsProbability</code>), 3=approximate treatment effect at boundaries, 4=conditional power at boundaries, 5=spending function plot (only available if <code>class(x)=="gsDesign"</code>), 6=expected sample size plot, and 7=B-values at boundaries. Character values for <code>plottype</code> may also be entered: "Z" for plot type 1, "power" for plot type 2, "thetahat" for plot type 3, "CP" for plot type 4, "sf" for plot type 5, "ASN", "N" or "n" for plot type 6, and "B", "B-val" or "B-value" for plot type 7.
base	Default is <code>FALSE</code> , which means <code>ggplot2</code> graphics are used. If true, base graphics are used for plotting.
...	This allows many optional arguments that are standard when calling <code>plot</code> . Other arguments include: <code>theta</code> which is used for <code>plottype=2, 4, 6</code> ; normally defaults will be adequate; see details. <code>ses=TRUE</code> which applies only when <code>plottype=3</code> and <code>class(x)=="gsDesign"</code> ; indicates that approximate standardized effect size at the boundary is to be plotted rather than the approximate natural parameter. <code>xval="Default"</code> which is only effective when <code>plottype=2</code> or <code>6</code> . Appropriately scaled (reparameterized) values for x-axis for power and expected sample size graphs; see details.

Value

An object of `class(x)`; in many cases this is the input value of `x`, while in others `x$theta` is replaced and corresponding characteristics computed; see details.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Proschan, MA, Lan, KKG, Wittes, JT (2006), *Statistical Monitoring of Clinical Trials. A Unified Approach*. New York: Springer.

See Also

[gsDesign](#), [gsProbability](#)

Examples

```
library(ggplot2)
# symmetric, 2-sided design with O'Brien-Fleming-like boundaries
# lower bound is non-binding (ignored in Type I error computation)
# sample size is computed based on a fixed design requiring n=100
x <- gsDesign(k = 5, test.type = 2, n.fix = 100)
x

# the following translate to calls to plot.gsDesign since x was
# returned by gsDesign; run these commands one at a time
plot(x)
plot(x, plottype = 2)
plot(x, plottype = 3)
plot(x, plottype = 4)
plot(x, plottype = 5)
plot(x, plottype = 6)
plot(x, plottype = 7)

# choose different parameter values for power plot
# start with design in x from above
y <- gsProbability(
  k = 5, theta = seq(0, .5, .025), x$n.I,
  x$lower$bound, x$upper$bound
)

# the following translates to a call to plot.gsProbability since
# y has that type
plot(y)
```

print.nSurvival

*Time-to-event sample size calculation (Lachin-Foulkes)***Description**

nSurvival() is used to calculate the sample size for a clinical trial with a time-to-event endpoint. The Lachin and Foulkes (1986) method is used. nEvents uses the Schoenfeld (1981) approximation to provide sample size and power in terms of the underlying hazard ratio and the number of events observed in a survival analysis. The functions hrz2n(), hrn2z() and zn2hr() also use the Schoenfeld approximation to provide simple translations between hazard ratios, z-values and the number of events in an analysis; input variables can be given as vectors.

nSurvival() produces an object of class "nSurvival" with the number of subjects and events for a set of pre-specified trial parameters, such as accrual duration and follow-up period. The calculation is based on Lachin and Foulkes (1986) method and can be used for risk ratio or risk difference. The function also consider non-uniform (exponential) entry as well as uniform entry.

If the logical approx is TRUE, the variance under alternative hypothesis is used to replace the variance under null hypothesis. For non-uniform entry, a non-zero value of gamma for exponential entry must be supplied. For positive gamma, the entry distribution is convex, whereas for negative gamma, the entry distribution is concave.

nEvents() uses the Schoenfeld (1981) method to approximate the number of events n (given beta) or the power (given n). Arguments may be vectors or scalars, but any vectors must have the same length.

The functions hrz2n, hrn2z and zn2hr also all apply the Schoenfeld approximation for proportional hazards modeling. This approximation is based on the asymptotic normal distribution of the logrank statistic as well as related statistics are asymptotically normal. Let λ denote the underlying hazard ratio (λ_1/λ_2 in terms of the arguments to nSurvival). Further, let n denote the number of events observed when computing the statistic of interest and r the ratio of the sample size in an experimental group relative to a control. The estimated natural logarithm of the hazard ratio from a proportional hazards ratio is approximately normal with a mean of $\log \lambda$ and variance $(1 + r)^2/nr$. Let z denote a logrank statistic (or a Wald statistic or score statistic from a proportional hazards regression model). The same asymptotic theory implies z is asymptotically equivalent to a normalized estimate of the hazard ratio λ and thus z is asymptotically normal with variance 1 and mean

$$\frac{\log \lambda r}{(1 + r)^2}.$$

Plugging the estimated hazard ratio into the above equation allows approximating any one of the following based on the other two: the estimate hazard ratio, the number of events and the z-statistic. That is,

$$\begin{aligned}\hat{\lambda} &= \exp(z(1 + r)/\sqrt{nr}) \\ z &= \log(\hat{\lambda})\sqrt{nr}/(1 + r) \\ n &= (z(1 + r)/\log(\hat{\lambda}))^2/r.\end{aligned}$$

hrz2n() translates an observed interim hazard ratio and interim z-value into the number of events required for the Z-value and hazard ratio to correspond to each other. hrn2z() translates a hazard ratio and number of events into an approximate corresponding Z-value. zn2hr() translates a

Z-value and number of events into an approximate corresponding hazard ratio. Each of these functions has a default assumption of an underlying hazard ratio of 1 which can be changed using the argument `hr0`. `hrn2z()` and `zn2hr()` also have an argument `hr1` which is only used to compute the sign of the computed Z-value in the case of `hrn2z()` and whether or not a z-value > 0 corresponds to a hazard ratio $>$ or $<$ the null hazard ratio `hr0`.

Usage

```
## S3 method for class 'nSurvival'
print(x, ...)

nSurvival(
  lambda1 = 1/12,
  lambda2 = 1/24,
  Ts = 24,
  Tr = 12,
  eta = 0,
  ratio = 1,
  alpha = 0.025,
  beta = 0.1,
  sided = 1,
  approx = FALSE,
  type = c("rr", "rd"),
  entry = c("unif", "expo"),
  gamma = NA
)

nEvents(
  hr = 0.6,
  alpha = 0.025,
  beta = 0.1,
  ratio = 1,
  sided = 1,
  hr0 = 1,
  n = 0,
  tbl = FALSE
)

zn2hr(z, n, ratio = 1, hr0 = 1, hr1 = 0.7)

hrn2z(hr, n, ratio = 1, hr0 = 1, hr1 = 0.7)

hrz2n(hr, z, ratio = 1, hr0 = 1)
```

Arguments

<code>x</code>	An object of class "nSurvival" returned by <code>nSurvival()</code> (optional: used for output; "months" or "years" would be the 'usual' choices).
<code>...</code>	Allows additional arguments for <code>print.nSurvival()</code> .

<code>lambda1, lambda2</code>	event hazard rate for placebo and treatment group respectively.
<code>Ts</code>	maximum study duration.
<code>Tr</code>	accrual (recruitment) duration.
<code>eta</code>	equal dropout hazard rate for both groups.
<code>ratio</code>	randomization ratio between placebo and treatment group. Default is balanced design, i.e., randomization ratio is 1.
<code>alpha</code>	type I error rate. Default is 0.025 since 1-sided testing is default.
<code>beta</code>	type II error rate. Default is 0.10 (90% power). Not needed for <code>nEvents()</code> if <code>n</code> is provided.
<code>sided</code>	one or two-sided test? Default is one-sided test.
<code>approx</code>	logical. If TRUE, the approximation sample size formula for risk difference is used.
<code>type</code>	type of sample size calculation: risk ratio ("rr") or risk difference ("rd").
<code>entry</code>	patient entry type: uniform entry ("unif") or exponential entry ("expo").
<code>gamma</code>	rate parameter for exponential entry. NA if entry type is "unif" (uniform). A non-zero value is supplied if entry type is "expo" (exponential).
<code>hr</code>	Hazard ratio. For <code>nEvents</code> , this is the hazard ratio under the alternative hypothesis (> 0 , $\neq hr_0$). Both $hr < hr_0$ (experimental is beneficial when lower hazard is better) and $hr > hr_0$ (e.g., time-to-response or safety designs) are supported.
<code>hr0</code>	Hazard ratio under the null hypothesis (> 0 , $\neq hr$).
<code>n</code>	Number of events. For <code>nEvents</code> may be input to compute power rather than sample size.
<code>tbl</code>	Indicator of whether or not scalar (vector) or tabular output is desired for <code>nEvents()</code> .
<code>z</code>	A z-statistic.
<code>hr1</code>	Hazard ratio under the alternate hypothesis for <code>hrn2z, zn2hr</code> (> 0 , $\neq hr_0$). Used to set the sign convention for converting between hazard ratios and Z-values.

Value

`nSurvival` produces a list with the following component returned:

<code>type</code>	As input.
<code>entry</code>	As input.
<code>n</code>	Sample size required (computed).
<code>nEvents</code>	Number of events required (computed).
<code>lambda1</code>	As input.
<code>lambda2</code>	As input.
<code>eta</code>	As input.
<code>ratio</code>	As input.
<code>gamma</code>	As input.

alpha	As input.
beta	As input.
sided	As input.
Ts	As input.
Tr	As input.

nEvents produces a scalar or vector of sample sizes (or powers) when tbl=FALSE or, when tbl=TRUE a data frame of values with the following columns:

hr	As input.
n	If n[1]=0 on input (default), output contains the number of events need to obtain the input Type I and II error. If n[1]>0 on input, the input value is returned.
alpha	As input.
beta	If n[1]=0 on input (default), beta is output as input. Otherwise, this is the computed Type II error based on the input n.
Power	One minus the output beta. When tbl=FALSE, n[1]>0, this is the value or vector of values returned.
delta	Standardized effect size represented by input difference between null and alternative hypothesis hazard ratios.
ratio	Ratio of experimental to control sample size where 'experimental' is the same as the group with hazard represented in the numerator of the hazard ratio.
se	Estimated standard error for the observed log(hazard ratio) with the given sample size.

hrz2n outputs a number of events required to approximately have the input hazard ratio, z-statistic and sample size correspond. hrn2z outputs an approximate z-statistic corresponding to an input hazard ratio and number of events. zn2hr outputs an approximate hazard ratio corresponding to an input z-statistic and number of events.

Author(s)

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References

Lachin JM and Foulkes MA (1986), Evaluation of Sample Size and Power for Analyses of Survival with Allowance for Nonuniform Patient Entry, Losses to Follow-Up, Noncompliance, and Stratification. *Biometrics*, 42, 507-519.

Schoenfeld D (1981), The Asymptotic Properties of Nonparametric Tests for Comparing Survival Distributions. *Biometrika*, 68, 316-319.

See Also

vignette("gsDesignPackageOverview"), [plot.gsDesign](#), [gsDesign](#), [gsHR](#)

Examples

```

library(ggplot2)

# consider a trial with
# 2 year maximum follow-up
# 6 month uniform enrollment
# Treatment/placebo hazards = 0.1/0.2 per 1 person-year
# drop out hazard 0.1 per 1 person-year
# alpha = 0.025 (1-sided)
# power = 0.9 (default beta=.1)

ss <- nSurvival(
  lambda1 = .2, lambda2 = .1, eta = .1, Ts = 2, Tr = .5,
  sided = 1, alpha = .025
)

# group sequential translation with default bounds
# note that delta1 is log hazard ratio; used later in gsBoundSummary summary
x <- gsDesign(
  k = 5, test.type = 2, n.fix = ss$nEvents, nFixSurv = ss$n,
  delta1 = log(ss$lambda2 / ss$lambda1)
)

# boundary plot
plot(x)
# effect size plot
plot(x, plottype = "hr")
# total sample size
x$nSurv
# number of events at analyses
x$n.I
# print the design
x
# overall design summary
cat(summary(x))
# tabular summary of bounds
gsBoundSummary(x, deltaname = "HR", Nname = "Events", logdelta = TRUE)

# approximate number of events required using Schoenfeld's method
# for 2 different hazard ratios
nEvents(hr = c(.5, .6), tbl = TRUE)
# vector output
nEvents(hr = c(.5, .6))

# approximate power using Schoenfeld's method
# given 2 sample sizes and hr=.6
nEvents(hr = .6, n = c(50, 100), tbl = TRUE)
# vector output
nEvents(hr = .6, n = c(50, 100))

# approximate hazard ratio corresponding to 100 events and z-statistic of 2

```

```

zn2hr(n = 100, z = 2)
# same when hr0 is 1.1
zn2hr(n = 100, z = 2, hr0 = 1.1)
# same when hr0 is .9 and hr1 is greater than hr0
zn2hr(n = 100, z = 2, hr0 = .9, hr1 = 1)

# approximate number of events corresponding to z-statistic of 2 and
# estimated hazard ratio of .5 (or 2)
hrz2n(hr = .5, z = 2)
hrz2n(hr = 2, z = 2)

# approximate z statistic corresponding to 75 events
# and estimated hazard ratio of .6 (or 1/.6)
# assuming 2-to-1 randomization of experimental to control
hrn2z(hr = .6, n = 75, ratio = 2)
hrn2z(hr = 1 / .6, n = 75, ratio = 2)

```

repeatedPValueBinomialExact

Exact binomial repeated p-values for a group sequential design

Description

Computes repeated p-values for the exact binomial design implied by a [gsSurv()] object. The p-value at analysis 'j' is the smallest local one-sided alpha level for which the observed experimental-arm event count crosses the exact lower efficacy bound at that analysis. Non-binding futility bounds are ignored for the Type I error calculation.

Usage

```

repeatedPValueBinomialExact(
  gsD,
  n.I = NULL,
  x = NULL,
  interval = c(1e-20, 0.9999),
  tol = 1e-08,
  maxiter = 100,
  check = FALSE
)

```

Arguments

gsD	A 'gsSurv' object with 'test.type' 1 or 4.
n.I	Increasing integer total event counts at completed analyses. If 'NULL', the planned exact binomial event counts from 'toBinomialExact(gsD)' are used. This must have at most 1 value greater than or equal to planned final events ('gsD\$maxn.Iplan' if available, otherwise 'max(gsD\$n.I)').
x	Integer experimental-arm event counts at the analyses in 'n.I'.

interval	Search interval for the p-values. As in [sequentialPValue()], values outside this interval are truncated to the nearest endpoint.
tol	Relative tolerance for the monotone bisection search on the alpha scale.
maxiter	Maximum number of bisection iterations for each analysis.
check	Logical. If 'TRUE', checks the monotonicity of the alpha-indexed integer efficacy bounds on a coarse grid and warns if it is violated.

Value

A data frame with one row per completed analysis containing:

'Analysis' Analysis index.

'n.I' Total events at analysis.

'x' Observed experimental-arm events.

'repeated_p_value' Repeated p-value for the analysis.

'bound_at_repeated_p_value' Integer efficacy bound at the repeated p-value.

See Also

[sequentialPValueBinomialExact()], [sequentialPValue()], [toBinomialExact()], [gsBinomialExact()]

Examples

```
x <- gsSurv(
  k = 3, test.type = 4, alpha = 0.025, beta = 0.1, timing = c(0.45, 0.7),
  sfu = sfHSD, sfupar = -4, sfl = sfLDOF, sflpar = 0,
  lambdaC = 0.001, hr = 0.3, hr0 = 0.7, eta = 5e-04,
  gamma = 10, R = 16, T = 24, minfup = 8, ratio = 3
)
counts <- toBinomialExact(x)$n.I

repeatedPValueBinomialExact(gsd = x, n.I = counts, x = c(12, 23, 38))
```

sequentialPValue

Sequential p-value computation

Description

sequentialPValue computes a sequential p-value for a group sequential design using a spending function as described in Maurer and Bretz (2013) and previously defined by Liu and Anderson (2008). It is the minimum of repeated p-values computed at each analysis (Jennison and Turnbull, 2000). This is particularly useful for multiplicity methods such as the graphical method for group sequential designs where sequential p-values for multiple hypotheses can be used as nominal p-values to plug into a multiplicity graph. A sequential p-value is described as the minimum alpha level at which a one-sided group sequential bound would be rejected given interim and final observed results. It is meaningful for both one-sided designs and designs with non-binding futility

bounds (test.type 1, 4, 6), but not for 2-sided designs with binding futility bounds (test.type 2, 3 or 5). Mild restrictions are required on spending functions used, but these are satisfied for commonly used spending functions such as the Lan-DeMets spending function approximating an O'Brien-Fleming bound or a Hwang-Shih-DeCani spending function; see Maurer and Bretz (2013).

Usage

```
sequentialPValue(
  gsD = gsDesign(),
  n.I = NULL,
  Z = NULL,
  usTime = NULL,
  interval = c(1e-05, 0.9999)
)
```

Arguments

gsD	Group sequential design generated by gsDesign or gsSurv.
n.I	Event counts (for time-to-event outcomes) or sample size (for most other designs); numeric vector with increasing, positive values with at most one value greater than or equal to largest value in gsD\$n.I; NOTE: if NULL, planned n.I will be used (gsD\$n.I).
Z	Z-value tests corresponding to analyses in n.I; positive values indicate a positive finding; must have the same length as n.I.
usTime	Spending time for upper bound at specified analyses; specify default: NULL if this is to be based on information fraction; if not NULL, must have the same length as n.I; increasing positive values with at most 1 greater than or equal to 1.
interval	Interval for search to derive p-value; Default: c(1e-05, 0.9999). Lower end of interval must be >0 and upper end must be < 1. The primary reason to not use the defaults would likely be if a test were vs a Type I error <0.0001.

Details

Solution is found with a search using uniroot. This finds the maximum alpha-level for which an efficacy bound is crossed, completely ignoring any futility bound.

Value

Sequential p-value (single numeric one-sided p-value between 0 and 1). Note that if the sequential p-value is less than the lower end of the input interval, the lower of interval will be returned. Similarly, if the sequential p-value is greater than the upper end of the input interval, then the upper end of interval is returned.

Author(s)

Keaven Anderson

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Liu, Qing, and Keaven M. Anderson. "On adaptive extensions of group sequential trials for clinical investigations." *Journal of the American Statistical Association* 103.484 (2008): 1621-1630.

Maurer, Willi, and Frank Bretz. "Multiple testing in group sequential trials using graphical approaches." *Statistics in Biopharmaceutical Research* 5.4 (2013): 311-320.;

Examples

```
# Derive Group Sequential Design
x <- gsSurv(k = 4, alpha = 0.025, beta = 0.1, timing = c(.5,.65,.8), sfu = sfLDOF,
           sfl = sfHSD, sflpar = 2, lambdaC = log(2)/6, hr = 0.6,
           eta = 0.01, gamma = c(2.5,5,7.5,10), R = c( 2,2,2,6 ),
           T = 30, minfup = 18)

x$n.I
# Analysis at IA2
sequentialPValue(gSD=x,n.I=c(100,160),Z=c(1.5,2))
# Use planned spending instead of information fraction; do final analysis
seqp <- sequentialPValue(gSD=x,n.I=c(100,160,190,230),Z=c(1.5,2,2.5,3),usTime=x$timing)
seqp
# Check bounds for updated design to verify at least one was crossed
xupdate <- gsDesign(maxn.IPlan=max(x$n.I),n.I=c(100,160,190,230),usTime=x$timing,
                   delta=x$delta,delta1=x$delta1,k=4,alpha=x$alpha,test.type=1,
                   sfu=x$upper$sf,sfupar=x$upper$param)
gsBoundSummary(xupdate,logdelta=TRUE,Nname="Events",deltaname="HR")
# Now update bound using sequential p-value as alpha level
xupdate <- gsDesign(maxn.IPlan=max(x$n.I),n.I=c(100,160,190,230),usTime=x$timing,
                   delta=x$delta,delta1=x$delta1,k=4,alpha=seqp,test.type=1,
                   sfu=x$upper$sf,sfupar=x$upper$param)
# Check that we are at bound for 1 (or more) analysis and did not exceed bound for others
gsBoundSummary(xupdate,logdelta=TRUE,Nname="Events",deltaname="HR")
```

sequentialPValueBinomialExact

Exact binomial sequential p-value for a group sequential design

Description

Computes the sequential p-value as the minimum repeated p-value over completed analyses, using [repeatedPValueBinomialExact()].

Usage

```
sequentialPValueBinomialExact(
  gSD,
  n.I = NULL,
  x = NULL,
```

```

interval = c(1e-20, 0.9999),
tol = 1e-08,
maxiter = 100,
check = FALSE
)

```

Arguments

<code>gsD</code>	A 'gsSurv' object with 'test.type' 1 or 4.
<code>n.I</code>	Increasing integer total event counts at completed analyses. If 'NULL', the planned exact binomial event counts from 'toBinomialExact(gsD)' are used. This must have at most 1 value greater than or equal to planned final events ('gsD\$maxn.Iplan' if available, otherwise 'max(gsD\$n.I)').
<code>x</code>	Integer experimental-arm event counts at the analyses in 'n.I'.
<code>interval</code>	Search interval for the p-values. As in [sequentialPValue()], values outside this interval are truncated to the nearest endpoint.
<code>tol</code>	Relative tolerance for the monotone bisection search on the alpha scale.
<code>maxiter</code>	Maximum number of bisection iterations for each analysis.
<code>check</code>	Logical. If 'TRUE', checks the monotonicity of the alpha-indexed integer efficacy bounds on a coarse grid and warns if it is violated.

Value

A single numeric one-sided sequential p-value.

See Also

[repeatedPValueBinomialExact()], [sequentialPValue()]

Examples

```

x <- gsSurv(
  k = 3, test.type = 4, alpha = 0.025, beta = 0.1, timing = c(0.45, 0.7),
  sfu = sfHSD, sfupar = -4, sfl = sfLDOF, sflpar = 0,
  lambdaC = 0.001, hr = 0.3, hr0 = 0.7, eta = 5e-04,
  gamma = 10, R = 16, T = 24, minfup = 8, ratio = 3
)
counts <- toBinomialExact(x)$n.I

sequentialPValueBinomialExact(gsD = x, n.I = counts, x = c(12, 23, 38))

```

sfExponential

*Exponential Spending Function***Description**

The function `sfExponential` implements the exponential spending function (Anderson and Clark, 2009). Normally `sfExponential` will be passed to `gsDesign` in the parameter `sfu` for the upper bound or `sf1` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence. The calling sequence is useful, however, when the user wishes to plot a spending function as demonstrated below in examples.

An exponential spending function is defined for any positive ν and $0 \leq t \leq 1$ as

$$f(t; \alpha, \nu) = \alpha(t) = \alpha^{t^{-\nu}}.$$

A value of $\nu=0.8$ approximates an O'Brien-Fleming spending function well.

The general class of spending functions this family is derived from requires a continuously increasing cumulative distribution function defined for $x > 0$ and is defined as

$$f(t; \alpha, \nu) = 1 - F(F^{-1}(1 - \alpha)/t^\nu).$$

The exponential spending function can be derived by letting $F(x) = 1 - \exp(-x)$, the exponential cumulative distribution function. This function was derived as a generalization of the Lan-DeMets (1983) spending function used to approximate an O'Brien-Fleming spending function (`sfLDOF()`),

$$f(t; \alpha) = 2 - 2\Phi\left(\Phi^{-1}(1 - \alpha/2)/t^{1/2}\right).$$

Usage

```
sfExponential(alpha, t, param)
```

Arguments

<code>alpha</code>	Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information.
<code>t</code>	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed.
<code>param</code>	A single positive value specifying the ν parameter for which the exponential spending is to be computed; allowable range is (0, 1.5].

Value

An object of type `spendfn`.

Note

The gsDesign technical manual shows how to use sfExponential() to closely approximate an O'Brien-Fleming design. An example is given below. The manual is available at <<https://keaven.github.io/gsd-tech-manual/>>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Anderson KM and Clark JB (2009), Fitting spending functions. *Statistics in Medicine*; 29:321-327.
 Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.
 Lan, KKG and DeMets, DL (1983), Discrete sequential boundaries for clinical trials. *Biometrika*; 70:659-663.

See Also

vignette("SpendingFunctionOverview"), [gsDesign](#), vignette("gsDesignPackageOverview")

Examples

```
library(ggplot2)
# use 'best' exponential approximation for k=6 to O'Brien-Fleming design
# (see manual for details)
gsDesign(
  k = 6, sfu = sfExponential, sfupar = 0.7849295,
  test.type = 2
)$upper$bound

# show actual O'Brien-Fleming bound
gsDesign(k = 6, sfu = "OF", test.type = 2)$upper$bound

# show Lan-DeMets approximation
# (not as close as sfExponential approximation)
gsDesign(k = 6, sfu = sfLD0F, test.type = 2)$upper$bound

# plot exponential spending function across a range of values of interest
t <- 0:100 / 100
plot(t, sfExponential(0.025, t, 0.8)$spend,
     xlab = "Proportion of final sample size",
     ylab = "Cumulative Type I error spending",
     main = "Exponential Spending Function Example", type = "l"
)
lines(t, sfExponential(0.025, t, 0.5)$spend, lty = 2)
lines(t, sfExponential(0.025, t, 0.3)$spend, lty = 3)
lines(t, sfExponential(0.025, t, 0.2)$spend, lty = 4)
lines(t, sfExponential(0.025, t, 0.15)$spend, lty = 5)
legend(
```

```
x = c(.0, .3), y = .025 * c(.7, 1), lty = 1:5,
legend = c(
  "nu = 0.8", "nu = 0.5", "nu = 0.3", "nu = 0.2",
  "nu = 0.15"
)
)
text(x = .59, y = .95 * .025, labels = "<--approximates O'Brien-Fleming")
```

sfHSD

*Hwang-Shih-DeCani Spending Function***Description**

The function `sfHSD` implements a Hwang-Shih-DeCani spending function. This is the default spending function for `gsDesign()`. Normally it will be passed to `gsDesign` in the parameter `sfu` for the upper bound or `sf1` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence. The calling sequence is useful, however, when the user wishes to plot a spending function as demonstrated below in examples.

A Hwang-Shih-DeCani spending function takes the form

$$f(t; \alpha, \gamma) = \alpha(1 - e^{-\gamma t}) / (1 - e^{-\gamma})$$

where γ is the value passed in `param`. A value of $\gamma = -4$ is used to approximate an O'Brien-Fleming design (see [sfExponential](#) for a better fit), while a value of $\gamma = 1$ approximates a Pocock design well.

Usage

```
sfHSD(alpha, t, param)
```

Arguments

<code>alpha</code>	Real value > 0 and no more than 1. Normally, <code>alpha=0.025</code> for one-sided Type I error specification or <code>alpha=0.1</code> for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information.
<code>t</code>	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed.
<code>param</code>	A single real value specifying the gamma parameter for which Hwang-Shih-DeCani spending is to be computed; allowable range is $[-40, 40]$

Value

An object of type `spendfn`. See `vignette("SpendingFunctionOverview")` for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
library(ggplot2)
# design a 4-analysis trial using a Hwang-Shih-DeCani spending function
# for both lower and upper bounds
x <- gsDesign(k = 4, sfu = sfHSD, sfupar = -2, sfl = sfHSD, sflpar = 1)

# print the design
x

# since sfHSD is the default for both sfu and sfl,
# this could have been written as
x <- gsDesign(k = 4, sfupar = -2, sflpar = 1)

# print again
x

# plot the spending function using many points to obtain a smooth curve
# show default values of gamma to see how the spending function changes
# also show gamma=1 which is supposed to approximate a Pocock design
t <- 0:100 / 100
plot(t, sfHSD(0.025, t, -4)$spend,
     xlab = "Proportion of final sample size",
     ylab = "Cumulative Type I error spending",
     main = "Hwang-Shih-DeCani Spending Function Example", type = "l"
)
lines(t, sfHSD(0.025, t, -2)$spend, lty = 2)
lines(t, sfHSD(0.025, t, 1)$spend, lty = 3)
legend(
  x = c(.0, .375), y = .025 * c(.8, 1), lty = 1:3,
  legend = c("gamma= -4", "gamma= -2", "gamma= 1")
)
```

Description

Lan and DeMets (1983) first published the method of using spending functions to set boundaries for group sequential trials. In this publication they proposed two specific spending functions: one to approximate an O'Brien-Fleming design and the other to approximate a Pocock design. The spending function to approximate O'Brien-Fleming has been generalized as proposed by Liu, et al (2012)

With `param=1=rho`, the Lan-DeMets (1983) spending function to approximate an O'Brien-Fleming bound is implemented in the function `(sfLDOF())`:

$$f(t; \alpha) = 2 - 2\Phi \left(\Phi^{-1}(1 - \alpha/2)/t^{\rho/2} \right).$$

For `rho` otherwise in `[.005, 2]`, this is the generalized version of Liu et al (2012). For `param` outside of `[.005, 2]`, `rho` is set to 1. The Lan-DeMets (1983) spending function to approximate a Pocock design is implemented in the function `sfLDPocock()`:

$$f(t; \alpha) = \alpha \ln(1 + (e - 1)t).$$

As shown in examples below, other spending functions can be used to get as good or better approximations to Pocock and O'Brien-Fleming bounds. In particular, O'Brien-Fleming bounds can be closely approximated using [sfExponential](#).

Usage

```
sfLDOF(alpha, t, param = NULL)
```

```
sfLDPocock(alpha, t, param)
```

Arguments

<code>alpha</code>	Real value > 0 and no more than 1. Normally, <code>alpha=0.025</code> for one-sided Type I error specification or <code>alpha=0.1</code> for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information.
<code>t</code>	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed.
<code>param</code>	This parameter is not used for <code>sfLDPocock</code> , not required for <code>sfLDOF</code> and need not be specified. For <code>sfLDPocock</code> it is here so that the calling sequence conforms to the standard for spending functions used with <code>gsDesign()</code> . For <code>sfLDOF</code> it will default to 1 (Lan-DeMets function to approximate O'Brien-Fleming) if <code>NULL</code> or if outside of the range <code>[.005, 2]</code> . otherwise, it will be use to set <code>rho</code> from Liu et al (2012).

Value

An object of type `spendfn`. See spending functions for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Lan, KKG and DeMets, DL (1983), Discrete sequential boundaries for clinical trials. *Biometrika*;70: 659-663.

Liu, Q, Lim, P, Nuamah, I, and Li, Y (2012), On adaptive error spending approach for group sequential trials with random information levels. *Journal of biopharmaceutical statistics*; 22(4), 687-699.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
library(ggplot2)
# 2-sided, symmetric 6-analysis trial Pocock
# spending function approximation
gsDesign(k = 6, sfu = sfLDPocock, test.type = 2)$upper$bound

# show actual Pocock design
gsDesign(k = 6, sfu = "Pocock", test.type = 2)$upper$bound

# approximate Pocock again using a standard
# Hwang-Shih-DeCani approximation
gsDesign(k = 6, sfu = sfHSD, sfupar = 1, test.type = 2)$upper$bound

# use 'best' Hwang-Shih-DeCani approximation for Pocock, k=6;
# see manual for details
gsDesign(k = 6, sfu = sfHSD, sfupar = 1.3354376, test.type = 2)$upper$bound

# 2-sided, symmetric 6-analysis trial
# O'Brien-Fleming spending function approximation
gsDesign(k = 6, sfu = sfLDof, test.type = 2)$upper$bound

# show actual O'Brien-Fleming bound
gsDesign(k = 6, sfu = "OF", test.type = 2)$upper$bound

# approximate again using a standard Hwang-Shih-DeCani
# approximation to O'Brien-Fleming
x <- gsDesign(k = 6, test.type = 2)
x$upper$bound
x$upper$param

# use 'best' exponential approximation for k=6; see manual for details
gsDesign(
  k = 6, sfu = sfExponential, sfupar = 0.7849295,
```

```

    test.type = 2
  )$upper$bound

# plot spending functions for generalized Lan-DeMets approximation of
ti <- (0:100)/100
rho <- c(.05, .5, 1, 1.5, 2, 2.5, 3:6, 8, 10, 12.5, 15, 20, 30, 200)/10
df <- NULL
for(r in rho){
  df <- rbind(df, data.frame(t=ti, rho=r, alpha=.025, spend=sfLDOF(alpha=.025, t=ti, param=r)$spend))
}
ggplot(df, aes(x=t, y=spend, col=as.factor(rho)))+
  geom_line()+
  guides(col=guide_legend(expression(rho)))+
  ggtitle("Generalized Lan-DeMets O'Brien-Fleming Spending Function")

```

sfLinear

Piecewise Linear and Step Function Spending Functions

Description

The function `sfLinear()` allows specification of a piecewise linear spending function. The function `sfStep()` specifies a step function spending function. Both functions provide complete flexibility in setting spending at desired timepoints in a group sequential design. Normally these function will be passed to `gsDesign()` in the parameter `sfu` for the upper bound or `sf1` for the lower bound to specify a spending function family for a design. When passed to `gsDesign()`, the value of `param` would be passed to `sfLinear()` or `sfStep()` through the `gsDesign()` arguments `sfupar` for the upper bound and `sf1par` for the lower bound.

Note that `sfStep()` allows setting a particular level of spending when the timing is not strictly known; an example shows how this can inflate Type I error when timing of analyses are changed based on knowing the treatment effect at an interim.

Usage

```
sfLinear(alpha, t, param)
```

```
sfStep(alpha, t, param)
```

Arguments

- | | |
|-------|---|
| alpha | Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size or information. |
| t | A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size or information for which the spending function will be computed. |

param A vector with a positive, even length. Values must range from 0 to 1, inclusive. Letting $m \leftarrow \text{length}(\text{param}/2)$, the first m points in `param` specify increasing values strictly between 0 and 1 corresponding to interim timing (proportion of final total statistical information). The last m points in `param` specify non-decreasing values from 0 to 1, inclusive, with the cumulative proportion of spending at the specified timepoints.

Value

An object of type `spendfn`. The cumulative spending returned in `sfLinear$spend` is 0 for $t \leq 0$ and `alpha` for $t \geq 1$. For t between specified points, linear interpolation is used to determine `sfLinear$spend`.

The cumulative spending returned in `sfStep$spend` is 0 for $t < \text{param}[1]$ and `alpha` for $t \geq 1$. Letting $m \leftarrow \text{length}(\text{param}/2)$, for $i = 1, 2, \dots, m-1$ and $\text{param}[i] \leq t < \text{param}[i+1]$, the cumulative spending is set at $\text{alpha} * \text{param}[i+m]$ (also for $\text{param}[m] \leq t < 1$).

Note that if `param[2m]` is 1, then the first time an analysis is performed after the last proportion of final planned information (`param[m]`) will be the final analysis, using any remaining error that was not previously spent.

See `vignette("SpendingFunctionOverview")` for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
library(ggplot2)
# set up alpha spending and beta spending to be piecewise linear
sfupar <- c(.2, .4, .05, .2)
sflpar <- c(.3, .5, .65, .5, .75, .9)
x <- gsDesign(sfu = sfLinear, sfl = sfLinear, sfupar = sfupar, sflpar = sflpar)
plot(x, plottype = "sf")
x

# now do an example where there is no lower-spending at interim 1
# and no upper spending at interim 2
sflpar <- c(1 / 3, 2 / 3, 0, .25)
```

```

sfupar <- c(1 / 3, 2 / 3, .1, .1)
x <- gsDesign(sfu = sfLinear, sfl = sfLinear, sfupar = sfupar, sflpar = sflpar)
plot(x, plottype = "sf")
x

# now do an example where timing of interims changes slightly, but error spending does not
# also, spend all alpha when at least >=90 percent of final information is in the analysis
sfupar <- c(.2, .4, .9, ((1:3) / 3)^3)
x <- gsDesign(k = 3, n.fix = 100, sfu = sfStep, sfupar = sfupar, test.type = 1)
plot(x, pl = "sf")
# original planned sample sizes
ceiling(x$n.I)
# cumulative spending planned at original interims
cumsum(x$upper$spend)
# change timing of analyses;
# note that cumulative spending "P(Cross) if delta=0" does not change from cumsum(x$upper$spend)
# while full alpha is spent, power is reduced by reduced sample size
y <- gsDesign(
  k = 3, sfu = sfStep, sfupar = sfupar, test.type = 1,
  maxn.IPlan = x$n.I[x$k], n.I = c(30, 70, 95),
  n.fix = x$n.fix
)
# note that full alpha is used, but power is reduced due to lowered sample size
gsBoundSummary(y)

# now show how step function can be abused by 'adapting' stage 2 sample size based on interim result
x <- gsDesign(k = 2, delta = .05, sfu = sfStep, sfupar = c(.02, .001), timing = .02, test.type = 1)
# spending jumps from miniscule to full alpha at first analysis after interim 1
plot(x, pl = "sf")
# sample sizes at analyses:
ceiling(x$n.I)
# simulate 1 million stage 1 sum of 178 Normal(0,1) random variables
# Normal(0,Variance=178) under null hypothesis
s1 <- rnorm(1000000, 0, sqrt(178))
# compute corresponding z-values
z1 <- s1 / sqrt(178)
# set stage 2 sample size to 1 if z1 is over final bound, otherwise full sample size
n2 <- rep(1, 1000000)
n2[z1 < 1.96] <- ceiling(x$n.I[2]) - ceiling(178)
# now sample n2 observations for second stage
s2 <- rnorm(1000000, 0, sqrt(n2))
# add sum and divide by standard deviation
z2 <- (s1 + s2) / (sqrt(178 + n2))
# By allowing full spending when final analysis is either
# early or late depending on observed interim z1,
# Type I error is now almost twice the planned .025
sum(z1 >= x$upper$bound[1] | z2 >= x$upper$bound[2]) / 1000000
# if stage 2 sample size is random and independent of z1 with same frequency,
# this is not a problem
s1alt <- rnorm(1000000, 0, sqrt(178))
z1alt <- s1alt / sqrt(178)
z2alt <- (s1alt + s2) / sqrt(178 + n2)
sum(z1alt >= x$upper$bound[1] | z2alt >= x$upper$bound[2]) / 1000000

```

sfLogistic

*Two-parameter Spending Function Families***Description**

The functions `sfLogistic()`, `sfNormal()`, `sfExtremeValue()`, `sfExtremeValue2()`, `sfCauchy()`, and `sfBetaDist()` are all 2-parameter spending function families. These provide increased flexibility in some situations where the flexibility of a one-parameter spending function family is not sufficient. These functions all allow fitting of two points on a cumulative spending function curve; in this case, four parameters are specified indicating an x and a y coordinate for each of 2 points. Normally each of these functions will be passed to `gsDesign()` in the parameter `sfu` for the upper bound or `sf1` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence. The calling sequence is useful, however, when the user wishes to plot a spending function as demonstrated in the examples; note, however, that an automatic α - and β -spending function plot is also available.

`sfBetaDist(alpha, t, param)` is simply α times the incomplete beta cumulative distribution function with parameters a and b passed in `param` evaluated at values passed in `t`.

The other spending functions take the form

$$f(t; \alpha, a, b) = \alpha F(a + bF^{-1}(t))$$

where $F()$ is a cumulative distribution function with values > 0 on the real line (logistic for `sfLogistic()`, normal for `sfNormal()`, extreme value for `sfExtremeValue()` and Cauchy for `sfCauchy()`) and $F^{-1}()$ is its inverse.

For the logistic spending function this simplifies to

$$f(t; \alpha, a, b) = \alpha(1 - (1 + e^{a(t/(1-t))^b})^{-1}).$$

For the extreme value distribution with

$$F(x) = \exp(-\exp(-x))$$

this simplifies to

$$f(t; \alpha, a, b) = \alpha \exp(-e^a(-\ln t)^b).$$

Since the extreme value distribution is not symmetric, there is also a version where the standard distribution is flipped about 0. This is reflected in `sfExtremeValue2()` where

$$F(x) = 1 - \exp(-\exp(x)).$$

Usage

```
sfLogistic(alpha, t, param)

sfBetaDist(alpha, t, param)

sfCauchy(alpha, t, param)

sfExtremeValue(alpha, t, param)

sfExtremeValue2(alpha, t, param)

sfNormal(alpha, t, param)
```

Arguments

alpha	Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size or information.
t	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size or information for which the spending function will be computed.
param	In the two-parameter specification, <code>sfBetaDist()</code> requires 2 positive values, while <code>sfLogistic()</code> , <code>sfNormal()</code> , <code>sfExtremeValue()</code> , <code>sfExtremeValue2()</code> and <code>sfCauchy()</code> require the first parameter to be any real value and the second to be a positive value. The four parameter specification is <code>c(t1, t2, u1, u2)</code> where the objective is that $\text{sf}(t1)=\alpha*u1$ and $\text{sf}(t2)=\alpha*u2$. In this parameterization, all four values must be between 0 and 1 and $t1 < t2$, $u1 < u2$.

Value

An object of type `spendfn`. See `vignette("SpendingFunctionOverview")` for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also[gsDesign](#)**Examples**

```

library(ggplot2)
# design a 4-analysis trial using a Kim-DeMets spending function
# for both lower and upper bounds
x <- gsDesign(k = 4, sfu = sfPower, sfupar = 3, sfl = sfPower, sflpar = 1.5)

# print the design
x

# plot the alpha- and beta-spending functions
plot(x, plottype = 5)

# start by showing how to fit two points with sfLogistic
# plot the spending function using many points to obtain a smooth curve
# note that curve fits the points x=.1, y=.01 and x=.4, y=.1
# specified in the 3rd parameter of sfLogistic
t <- 0:100 / 100
plot(t, sfLogistic(1, t, c(.1, .4, .01, .1))$spend,
     xlab = "Proportion of final sample size",
     ylab = "Cumulative Type I error spending",
     main = "Logistic Spending Function Examples",
     type = "l", cex.main = .9
)
lines(t, sfLogistic(1, t, c(.01, .1, .1, .4))$spend, lty = 2)

# now just give a=0 and b=1 as 3rd parameters for sfLogistic
lines(t, sfLogistic(1, t, c(0, 1))$spend, lty = 3)

# try a couple with unconventional shapes again using
# the xy form in the 3rd parameter
lines(t, sfLogistic(1, t, c(.4, .6, .1, .7))$spend, lty = 4)
lines(t, sfLogistic(1, t, c(.1, .7, .4, .6))$spend, lty = 5)
legend(
  x = c(.0, .475), y = c(.76, 1.03), lty = 1:5,
  legend = c(
    "Fit (.1, .01) and (.4, .1)", "Fit (.01, .1) and (.1, .4)",
    "a=0, b=1", "Fit (.4, .1) and (.6, .7)",
    "Fit (.1, .4) and (.7, .6)"
  )
)

# set up a function to plot comparisons of all
# 2-parameter spending functions
plotsf <- function(alpha, t, param) {
  plot(t, sfCauchy(alpha, t, param)$spend,
       xlab = "Proportion of enrollment",
       ylab = "Cumulative spending", type = "l", lty = 2
  )
}

```

```

lines(t, sfExtremeValue(alpha, t, param)$spend, lty = 5)
lines(t, sfLogistic(alpha, t, param)$spend, lty = 1)
lines(t, sfNormal(alpha, t, param)$spend, lty = 3)
lines(t, sfExtremeValue2(alpha, t, param)$spend, lty = 6, col = 2)
lines(t, sfBetaDist(alpha, t, param)$spend, lty = 7, col = 3)
legend(
  x = c(.05, .475), y = .025 * c(.55, .9),
  lty = c(1, 2, 3, 5, 6, 7),
  col = c(1, 1, 1, 1, 2, 3),
  legend = c(
    "Logistic", "Cauchy", "Normal", "Extreme value",
    "Extreme value 2", "Beta distribution"
  )
)
}
# do comparison for a design with conservative early spending
# note that Cauchy spending function is quite different
# from the others
param <- c(.25, .5, .05, .1)
plotsf(.025, t, param)

```

sfPoints

*Pointwise Spending Function***Description**

The function `sfPoints` implements a spending function with values specified for an arbitrary set of specified points. It is now recommended to use `sfLinear` rather than `sfPoints`. Normally `sfPoints` will be passed to `gsDesign` in the parameter `sfu` for the upper bound or `sfl` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence, just the points they wish to specify. If using `sfPoints()` in a design, it is recommended to specify how to interpolate between the specified points (e.g., linear interpolation); also consider fitting smooth spending functions; see `vignette("SpendingFunctionOverview")`.

Usage

```
sfPoints(alpha, t, param)
```

Arguments

<code>alpha</code>	Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information.
<code>t</code>	A vector of points with increasing values from >0 and ≤ 1 . Values of the proportion of sample size/information for which the spending function will be computed.
<code>param</code>	A vector of the same length as <code>t</code> specifying the cumulative proportion of spending to corresponding to each point in <code>t</code> ; must be ≥ 0 and ≤ 1 .

Value

An object of type `spendfn`. See spending functions for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`, [sfLogistic](#)

Examples

```
library(ggplot2)
# example to specify spending on a pointwise basis
x <- gsDesign(
  k = 6, sfu = sfPoints, sfupar = c(.01, .05, .1, .25, .5, 1),
  test.type = 2
)
x

# get proportion of upper spending under null hypothesis
# at each analysis
y <- x$upper$prob[, 1] / .025

# change to cumulative proportion of spending
for (i in 2:length(y))
  y[i] <- y[i - 1] + y[i]

# this should correspond to input sfupar
round(y, 6)

# plot these cumulative spending points
plot(1:6 / 6, y,
  main = "Pointwise spending function example",
  xlab = "Proportion of final sample size",
  ylab = "Cumulative proportion of spending",
  type = "p"
)

# approximate this with a t-distribution spending function
```

```
# by fitting 3 points
tx <- 0:100 / 100
lines(tx, sfTDist(1, tx, c(c(1, 3, 5) / 6, .01, .1, .5))$spend)
text(x = .6, y = .9, labels = "Pointwise Spending Approximated by")
text(x = .6, y = .83, "t-Distribution Spending with 3-point interpolation")

# example without lower spending at initial interim or
# upper spending at last interim
x <- gsDesign(
  k = 3, sfu = sfPoints, sfupar = c(.25, .25),
  sfl = sfPoints, sflpar = c(0, .25)
)
x
```

sfPower

Kim-DeMets (power) Spending Function

Description

The function `sfPower()` implements a Kim-DeMets (power) spending function. This is a flexible, one-parameter spending function recommended by Jennison and Turnbull (2000). Normally it will be passed to `gsDesign()` in the parameter `sfu` for the upper bound or `sfl` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence. The calling sequence is useful, however, when the user wishes to plot a spending function as demonstrated below in examples.

A Kim-DeMets spending function takes the form

$$f(t; \alpha, \rho) = \alpha t^\rho$$

where ρ is the value passed in `param`. See examples below for a range of values of ρ that may be of interest (`param=0.75` to `3` are documented there).

Usage

```
sfPower(alpha, t, param)
```

Arguments

- | | |
|--------------------|--|
| <code>alpha</code> | Real value > 0 and no more than 1. Normally, <code>alpha=0.025</code> for one-sided Type I error specification or <code>alpha=0.1</code> for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information. |
| <code>t</code> | A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed. |
| <code>param</code> | A single, positive value specifying the ρ parameter for which Kim-DeMets spending is to be computed; allowable range is (0,50] |

Value

An object of type `spendfn`. See `vignette("SpendingFunctionOverview")` for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
library(ggplot2)
# design a 4-analysis trial using a Kim-DeMets spending function
# for both lower and upper bounds
x <- gsDesign(k = 4, sfu = sfPower, sfupar = 3, sfl = sfPower, sflpar = 1.5)

# print the design
x

# plot the spending function using many points to obtain a smooth curve
# show rho=3 for approximation to O'Brien-Fleming and rho=.75 for
# approximation to Pocock design.
# Also show rho=2 for an intermediate spending.
# Compare these to Hwang-Shih-DeCani spending with gamma=-4, -2, 1
t <- 0:100 / 100
plot(t, sfPower(0.025, t, 3)$spend,
     xlab = "Proportion of sample size",
     ylab = "Cumulative Type I error spending",
     main = "Kim-DeMets (rho) versus Hwang-Shih-DeCani (gamma) Spending",
     type = "l", cex.main = .9
)
lines(t, sfPower(0.025, t, 2)$spend, lty = 2)
lines(t, sfPower(0.025, t, 0.75)$spend, lty = 3)
lines(t, sfHSD(0.025, t, 1)$spend, lty = 3, col = 2)
lines(t, sfHSD(0.025, t, -2)$spend, lty = 2, col = 2)
lines(t, sfHSD(0.025, t, -4)$spend, lty = 1, col = 2)
legend(
  x = c(.0, .375), y = .025 * c(.65, 1), lty = 1:3,
  legend = c("rho= 3", "rho= 2", "rho= 0.75")
)
```

```

legend(
  x = c(.0, .357), y = .025 * c(.65, .85), lty = 1:3, bty = "n", col = 2,
  legend = c("gamma= -4", "gamma= -2", "gamma=1")
)

```

sfTDist

t-distribution Spending Function

Description

The function `sfTDist()` provides perhaps the maximum flexibility among spending functions provided in the `gsDesign` package. This function allows fitting of three points on a cumulative spending function curve; in this case, six parameters are specified indicating an x and a y coordinate for each of 3 points. Normally this function will be passed to `gsDesign()` in the parameter `sfu` for the upper bound or `sfl` for the lower bound to specify a spending function family for a design. In this case, the user does not need to know the calling sequence. The calling sequence is useful, however, when the user wishes to plot a spending function as demonstrated below in examples.

The t -distribution spending function takes the form

$$f(t; \alpha) = \alpha F(a + bF^{-1}(t))$$

where $F()$ is a cumulative t -distribution function with df degrees of freedom and $F^{-1}()$ is its inverse.

Usage

```
sfTDist(alpha, t, param)
```

Arguments

- | | |
|-------|---|
| alpha | Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information. |
| t | A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed. |
| param | In the three-parameter specification, the first parameter (a) may be any real value, the second (b) any positive value, and the third parameter (df =degrees of freedom) any real value 1 or greater. When <code>gsDesign()</code> is called with a t -distribution spending function, this is the parameterization printed. The five parameter specification is <code>c(t1, t2, u1, u2, df)</code> where the objective is that the resulting cumulative proportion of spending at t represented by <code>sf(t)</code> satisfies <code>sf(t1)=alpha*u1</code> , <code>sf(t2)=alpha*u2</code> . The t -distribution used has df degrees of freedom. In this parameterization, all the first four values must be between 0 and 1 and $t1 < t2$, $u1 < u2$. The final parameter is any real value of 1 or |

more. This parameterization can fit any two points satisfying these requirements. The six parameter specification attempts to fit 3 points, but does not have flexibility to fit any three points. In this case, the specification for param is $c(t1, t2, t3, u1, u2, u3)$ where the objective is that $sf(t1) = \alpha * u1$, $sf(t2) = \alpha * u2$, and $sf(t3) = \alpha * u3$. See examples to see what happens when points are specified that cannot be fit.

Value

An object of type `spendfn`. See spending functions for further details.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
library(ggplot2)
# 3-parameter specification: a, b, df
sfTDist(1, 1:5 / 6, c(-1, 1.5, 4))$spend

# 5-parameter specification fits 2 points, in this case
# the 1st 2 interims are at 25% and 50% of observations with
# cumulative error spending of 10% and 20%, respectively
# final parameter is df
sfTDist(1, 1:3 / 4, c(.25, .5, .1, .2, 4))$spend

# 6-parameter specification fits 3 points
# Interims are at 25%, 50% and 75% of observations
# with cumulative spending of 10%, 20% and 50%, respectively
# Note: not all 3 point combinations can be fit
sfTDist(1, 1:3 / 4, c(.25, .5, .75, .1, .2, .5))$spend

# Example of error message when the 3-points specified
# in the 6-parameter version cannot be fit
try(sfTDist(1, 1:3 / 4, c(.25, .5, .75, .1, .2, .3))$errmsg)

# sfCauchy (sfTDist with 1 df) and sfNormal (sfTDist with infinite df)
# show the limits of what sfTDist can fit
```

```

# for the third point are u3 from 0.344 to 0.6 when t3=0.75
sfNormal(1, 1:3 / 4, c(.25, .5, .1, .2))$spend[3]
sfCauchy(1, 1:3 / 4, c(.25, .5, .1, .2))$spend[3]

# plot a few t-distribution spending functions fitting
# t=0.25, .5 and u=0.1, 0.2
# to demonstrate the range of flexibility
t <- 0:100 / 100
plot(t, sfTDist(0.025, t, c(.25, .5, .1, .2, 1))$spend,
     xlab = "Proportion of final sample size",
     ylab = "Cumulative Type I error spending",
     main = "t-Distribution Spending Function Examples", type = "l"
)
lines(t, sfTDist(0.025, t, c(.25, .5, .1, .2, 1.5))$spend, lty = 2)
lines(t, sfTDist(0.025, t, c(.25, .5, .1, .2, 3))$spend, lty = 3)
lines(t, sfTDist(0.025, t, c(.25, .5, .1, .2, 10))$spend, lty = 4)
lines(t, sfTDist(0.025, t, c(.25, .5, .1, .2, 100))$spend, lty = 5)
legend(
  x = c(.0, .3), y = .025 * c(.7, 1), lty = 1:5,
  legend = c("df = 1", "df = 1.5", "df = 3", "df = 10", "df = 100")
)

```

sfTruncated

Truncated, trimmed and gapped spending functions

Description

The functions `sfTruncated()` and `sfTrimmed` apply any other spending function over a restricted range. This allows eliminating spending for early interim analyses when you desire not to stop for the bound being specified; this is usually applied to eliminate early tests for a positive efficacy finding. The truncation can come late in the trial if you desire to stop a trial any time after, say, 90 percent of information is available and an analysis is performed. This allows full Type I error spending if the final analysis occurs early. Both functions set cumulative spending to 0 below a 'spending interval' in the interval $[0,1]$, and set cumulative spending to 1 above this range. `sfTrimmed()` otherwise does not change an input spending function that is specified; probably the preferred and more intuitive method in most cases. `sfTruncated()` resets the time scale on which the input spending function is computed to the 'spending interval.'

`sfGapped()` allows elimination of analyses after some time point in the trial; see details and examples.

`sfTrimmed` simply computes the value of the input spending function and parameters in the sub-range of $[0,1]$, sets spending to 0 below this range and sets spending to 1 above this range.

`sfGapped` spends outside of the range provided in `trange`. Below `trange`, the input spending function is used. Above `trange`, full spending is used; i.e., the first analysis performed above the interval in `trange` is the final analysis. As long as the input spending function is strictly increasing, this means that the first interim in the interval `trange` is the final interim analysis for the bound being specified.

`sfTruncated` compresses spending into a sub-range of $[0,1]$. The parameter `param$strange` specifies the range over which spending is to occur. Within this range, spending is spent according

to the spending function specified in `param$sf` along with the corresponding spending function parameter(s) in `param$param`. See example using `sfLinear` that spends uniformly over specified range.

Usage

```
sfTruncated(alpha, t, param)
```

```
sfTrimmed(alpha, t, param)
```

```
sfGapped(alpha, t, param)
```

Arguments

<code>alpha</code>	Real value > 0 and no more than 1. Normally, $\alpha=0.025$ for one-sided Type I error specification or $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size or information.
<code>t</code>	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size or information for which the spending function will be computed.
<code>param</code>	a list containing the elements <code>sf</code> (a <code>spendfn</code> object such as <code>sfHSD</code>), <code>trange</code> (the range over which the spending function increases from 0 to 1; $0 \leq \text{trange}[1] < \text{trange}[2] \leq 1$; for <code>sfGapped</code> , <code>trange[1]</code> must be > 0), and <code>param</code> (null for a spending function with no parameters or a scalar or vector of parameters needed to fully specify the spending function in <code>sf</code>).

Value

An object of type `spendfn`. See `vignette("SpendingFunctionOverview")` for further details.

Note

The `gsDesign` technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

`vignette("SpendingFunctionOverview")`, [gsDesign](#), `vignette("gsDesignPackageOverview")`

Examples

```
# Eliminate efficacy spending for any interim at or before 20 percent of information.
# Complete spending at first interim at or after 80 percent of information.
tx <- (0:100) / 100
s <- sfHSD(alpha = .05, t = tx, param = 1)$spend
x <- data.frame(t = tx, Spending = s, sf = "Original spending")
param <- list(trange = c(.2, .8), sf = sfHSD, param = 1)
s <- sfTruncated(alpha = .05, t = tx, param = param)$spend
x <- rbind(x, data.frame(t = tx, Spending = s, sf = "Truncated"))
s <- sfTrimmed(alpha = .05, t = tx, param = param)$spend
x <- rbind(x, data.frame(t = tx, Spending = s, sf = "Trimmed"))
s <- sfGapped(alpha = .05, t = tx, param = param)$spend
x <- rbind(x, data.frame(t = tx, Spending = s, sf = "Gapped"))
ggplot2::ggplot(x, ggplot2::aes(x = t, y = Spending, col = sf)) +
  ggplot2::geom_line()

# now apply the sfTrimmed version in gsDesign
# initially, eliminate the early efficacy analysis
# note: final spend must occur at > next to last interim
x <- gsDesign(
  k = 4, n.fix = 100, sfu = sfTrimmed,
  sfupar = list(sf = sfHSD, param = 1, trange = c(.3, .9))
)

# first upper bound=20 means no testing there
gsBoundSummary(x)

# now, do not eliminate early efficacy analysis
param <- list(sf = sfHSD, param = 1, trange = c(0, .9))
x <- gsDesign(k = 4, n.fix = 100, sfu = sfTrimmed, sfupar = param)

# The above means if final analysis is done a little early, all spending can occur
# Suppose we set calendar date for final analysis based on
# estimated full information, but come up with only 97 pct of plan
xA <- gsDesign(
  k = x$k, n.fix = 100, n.I = c(x$n.I[1:3], .97 * x$n.I[4]),
  test.type = x$test.type,
  maxn.IPlan = x$n.I[x$k],
  sfu = sfTrimmed, sfupar = param
)
# now accelerate without the trimmed spending function
xNT <- gsDesign(
  k = x$k, n.fix = 100, n.I = c(x$n.I[1:3], .97 * x$n.I[4]),
  test.type = x$test.type,
  maxn.IPlan = x$n.I[x$k],
  sfu = sfHSD, sfupar = 1
)
# Check last bound if analysis done at early time
x$upper$bound[4]
# Now look at last bound if done at early time with trimmed spending function
```

```

# that allows capture of full alpha
xA$upper$bound[4]
# With original spending function, we don't get full alpha and therefore have
# unnecessarily stringent bound at final analysis
xNT$upper$bound[4]

# note that if the last analysis is LATE, all 3 approaches should give the same
# final bound that has a little larger z-value
xlate <- gsDesign(
  k = x$k, n.fix = 100, n.I = c(x$n.I[1:3], 1.25 * x$n.I[4]),
  test.type = x$test.type,
  maxn.Iplan = x$n.I[x$k],
  sfu = sfHSD, sfupar = 1
)
xlate$upper$bound[4]

# eliminate futility after the first interim analysis
# note that by setting trange[1] to .2, the spend at t=.2 is used for the first
# interim at or after 20 percent of information
x <- gsDesign(n.fix = 100, sfl = sfGapped, sflpar = list(trange = c(.2, .9), sf = sfHSD, param = 1))

```

sfXG1

Xi and Gallo conditional error spending functions

Description

Error spending functions based on Xi and Gallo (2019). The intention of these spending functions is to provide bounds where the conditional error at an efficacy bound is approximately equal to the conditional error rate for crossing the final analysis bound. This is explained in greater detail in `vignette("ConditionalErrorSpending")`.

Usage

```
sfXG1(alpha, t, param)
```

```
sfXG2(alpha, t, param)
```

```
sfXG3(alpha, t, param)
```

Arguments

alpha	Real value > 0 and no more than 1. Normally, $\alpha = 0.025$ for one-sided Type I error specification or $\alpha = 0.1$ for Type II error specification. However, this could be set to 1 if for descriptive purposes you wish to see the proportion of spending as a function of the proportion of sample size/information.
t	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed.

param This is the gamma parameter in the Xi and Gallo spending function paper, distinct for each function. See the details section for functional forms and range of param acceptable for each spending function.

Details

Xi and Gallo use an additive boundary for group sequential designs with connection to conditional error. Three spending functions are defined: sfXG1(), sfXG2(), and sfXG3().

Method 1 is defined for $\gamma \in [0.5, 1)$ as

$$f(Z_K \geq u_K | Z_k = u_k) = 2 - 2 \times \Phi \left(\frac{z_{\alpha/2} - z_\gamma \sqrt{1-t}}{\sqrt{t}} \right).$$

Method 2 is defined for $\gamma \in [1 - \Phi(z_{\alpha/2}/2), 1)$ as

$$f_\gamma(t; \alpha) = 2 - 2\Phi \left(\Phi^{-1}(1 - \alpha/2)/t^{1/2} \right).$$

Method 3 is defined as for $\gamma \in (\alpha/2, 1)$ as

$$f(t; \alpha) = 2 - 2 \times \Phi \left(\frac{z_{\alpha/2} - z_\gamma(1 - \sqrt{t})}{\sqrt{t}} \right).$$

Value

An object of type spendfn. See spending functions for further details.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

vignette("SpendingFunctionOverview"), [gsDesign](#), vignette("gsDesignPackageOverview")

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

Xi D and Gallo P (2019), An additive boundary for group sequential designs with connection to conditional error. *Statistics in Medicine*; 38 (23), 4656–4669.

Examples

```
# Plot conditional error spending functions across
# a range of values of interest
pts <- seq(0, 1.2, 0.01)
pal <- palette()

plot(
  pts,
  sfXG1(0.025, pts, 0.5)$spend,
```

```

    type = "l", col = pal[1],
    xlab = "t", ylab = "Spending", main = "Xi-Gallo, Method 1"
  )
  lines(pts, sfXG1(0.025, pts, 0.6)$spend, col = pal[2])
  lines(pts, sfXG1(0.025, pts, 0.75)$spend, col = pal[3])
  lines(pts, sfXG1(0.025, pts, 0.9)$spend, col = pal[4])
  legend(
    "topleft",
    legend = c("gamma=0.5", "gamma=0.6", "gamma=0.75", "gamma=0.9"),
    col = pal[1:4],
    lty = 1
  )
)

plot(
  pts,
  sfXG2(0.025, pts, 0.14)$spend,
  type = "l", col = pal[1],
  xlab = "t", ylab = "Spending", main = "Xi-Gallo, Method 2"
)
  lines(pts, sfXG2(0.025, pts, 0.25)$spend, col = pal[2])
  lines(pts, sfXG2(0.025, pts, 0.5)$spend, col = pal[3])
  lines(pts, sfXG2(0.025, pts, 0.75)$spend, col = pal[4])
  lines(pts, sfXG2(0.025, pts, 0.9)$spend, col = pal[5])
  legend(
    "topleft",
    legend = c("gamma=0.14", "gamma=0.25", "gamma=0.5", "gamma=0.75", "gamma=0.9"),
    col = pal[1:5],
    lty = 1
  )
)

plot(
  pts,
  sfXG3(0.025, pts, 0.013)$spend,
  type = "l", col = pal[1],
  xlab = "t", ylab = "Spending", main = "Xi-Gallo, Method 3"
)
  lines(pts, sfXG3(0.025, pts, 0.02)$spend, col = pal[2])
  lines(pts, sfXG3(0.025, pts, 0.05)$spend, col = pal[3])
  lines(pts, sfXG3(0.025, pts, 0.1)$spend, col = pal[4])
  lines(pts, sfXG3(0.025, pts, 0.25)$spend, col = pal[5])
  lines(pts, sfXG3(0.025, pts, 0.5)$spend, col = pal[6])
  lines(pts, sfXG3(0.025, pts, 0.75)$spend, col = pal[7])
  lines(pts, sfXG3(0.025, pts, 0.9)$spend, col = pal[8])
  legend(
    "bottomright",
    legend = c(
      "gamma=0.013", "gamma=0.02", "gamma=0.05", "gamma=0.1",
      "gamma=0.25", "gamma=0.5", "gamma=0.75", "gamma=0.9"
    ),
    col = pal[1:8],
    lty = 1
  )
)

```

simBinomialSeasonalExact

Simulate exact-binomial seasonal monitoring scenarios

Description

Simulate seasonal rare-event trials monitored with exact-binomial efficacy bounds derived from a ‘gsSurv’ design. This helper supports fixed enrollment and a simple blinded information-adaptive enrollment rule while keeping the spending framework fixed through the original ‘gsSurv’ design object.

The function summarizes empirical rejection rates (Type I error or power), futility stopping rates (binding interpretation), Monte Carlo standard errors, average final events, average total enrollment, and average number of informative looks.

Usage

```
simBinomialSeasonalExact(
  gsD,
  ve = c(0.3, 0.8),
  nsim = c(600, 600),
  control_event_rate = c(0.003, 0.003),
  season_length = 0.5,
  dropout_rate = 0.1,
  planned_counts = NULL,
  timing = NULL,
  enroll_control_per_look = NULL,
  enroll_experimental_per_look = NULL,
  adaptive = c(FALSE, TRUE),
  adapt_looks = NULL,
  max_multiplier = 2,
  usTime = NULL,
  lsTime = NULL,
  final_full_spending = FALSE,
  seed = NULL,
  return_trials = FALSE
)
```

Arguments

gsD	A ‘gsSurv’ object with ‘test.type’ 1 or 4.
ve	Numeric vector of vaccine efficacy (or prevention efficacy) scenarios to simulate. Each value must be finite and less than 1. ‘ve = 0’ corresponds to equal event rates (superiority null); ‘ve < 0’ corresponds to experimental-arm event rates above control (non-inferiority margin or harmful scenarios).
nsim	Integer scalar or vector giving the number of simulations per element of ‘ve’.

control_event_rate	Numeric scalar or vector with control seasonal event probabilities corresponding to 've'.
season_length	Numeric scalar > 0 giving season duration in years.
dropout_rate	Seasonal dropout probability in '[0, 1)'.
planned_counts	Optional increasing integer vector of planned cumulative events at analyses. If 'NULL', these are derived from 'timing * toInteger(gsD)\$n.I[k]'.
timing	Optional increasing cumulative spending-time vector ending at 1 used to derive 'planned_counts' when 'planned_counts = NULL'.
enroll_control_per_look	Optional control-arm enrollment by look (scalar or length 'k' integer vector). If both enrollment vectors are 'NULL', defaults are derived from the seasonal accrual pattern in 'gsD'.
enroll_experimental_per_look	Optional experimental-arm enrollment by look (scalar or length 'k' integer vector). If 'NULL' and 'enroll_control_per_look' is supplied, this is set using 'gsD\$ratio'.
adaptive	Logical vector specifying whether to simulate fixed and/or adaptive enrollment scenarios.
adapt_looks	Integer vector of look indices after which adaptation can be applied (default: all interim looks).
max_multiplier	Maximum multiplicative enrollment increase at a look when adaptation is enabled.
usTime	Optional upper spending-time override passed to [toBinomialExact()]. If 'NULL', spending time defaults to '1 / k, 2 / k, ..., 1'.
lsTime	Optional lower spending-time override for 'test.type = 4'. If 'NULL', this defaults to 'usTime'.
final_full_spending	Logical scalar. If 'TRUE', force full alpha spending at the final analysis even when the final observed total event count is below planned final events.
seed	Optional integer seed for reproducibility.
return_trials	Logical. If 'TRUE', return trial-level simulation outcomes.

Value

A list with:

'summary' Data frame with scenario-level summaries.

'planned' List with planned counts, exact design object, and planned/calibrated enrollment by look.

'inputs' List of simulation inputs used.

'trials' Optional trial-level data frame ('NULL' unless 'return_trials = TRUE').

See Also

[toBinomialExact()], [repeatedPValueBinomialExact()], [sequentialPValueBinomialExact()]

Examples

```
x <- gsSurv(
  k = 3, test.type = 4, alpha = 0.025, beta = 0.1, timing = c(1 / 3, 2 / 3),
  sfu = sfHSD, sfupar = 1, sfl = sfHSD, sflpar = -2,
  lambdaC = -log(1 - 0.003) / 0.5,
  hr = 0.2, hr0 = 0.7, eta = -log(1 - 0.1) / 0.5,
  gamma = c(1, 0, 1, 0, 1, 0), R = c(2, 10, 2, 10, 2, 10),
  T = 42, minfup = 6, ratio = 3
) |> toInteger()

simBinomialSeasonalExact(
  gsD = x,
  ve = c(0.3, 0.8),
  nsim = c(50, 50),
  control_event_rate = c(0.003, 0.003),
  seed = 123
)$summary
```

summary.gsDesign

Bound Summary and Z-transformations

Description

A tabular summary of a group sequential design's bounds and their properties are often useful. The 'vintage' `print.gsDesign()` function provides a complete but minimally formatted summary of a group sequential design derived by `gsDesign()`. A brief description of the overall design can also be useful (`summary.gsDesign()`). A tabular summary of boundary characteristics oriented only towards LaTeX output is produced by `xtable.gsSurv`. More flexibility is provided by `gsBoundSummary()` which produces a tabular summary of a user-specifiable set of package-provided boundary properties in a data frame. This can also be used to along with functions such as `print.data.frame()`, `write.table()`, `write.csv()`, `write.csv2()` or, from the RTF package, `addTable.RTF()` (from the rtf package) to produce console or R Markdown output or output to a variety of file types. `xprint()` is provided for LaTeX output by setting default options for `print.xtable` when producing tables summarizing design bounds.

Individual transformation of z-value test statistics for interim and final analyses are obtained from `gsBValue()`, `gsDelta()`, `gsHR()` and `gsCPz()` for B-values, approximate treatment effect (see details), approximate hazard ratio and conditional power, respectively.

The `print.gsDesign` function is intended to provide an easier output to review than is available from a simple list of all the output components. The `gsBoundSummary` function is intended to provide a summary of boundary characteristics that is often useful for evaluating boundary selection; this outputs an extension of the `data.frame` class that sets up default printing without row names using `print.gsBoundSummary`. `summary.gsDesign`, on the other hand, provides a summary of the overall design at a higher level; this provides characteristics not included in the `gsBoundSummary` summary and no detail concerning interim analysis bounds.

In brief, the computed descriptions of group sequential design bounds are as follows: Z: Standardized normal test statistic at design bound.

p (1-sided): 1-sided p-value for Z. This will be computed as the probability of a greater EXCEPT for lower bound when a 2-sided design is being summarized.

delta at bound: Approximate value of the natural parameter at the bound. The approximate standardized effect size at the bound is generally computed as $Z/\sqrt{n}$. Calling this theta, this is translated to the delta using the values delta0 and delta1 from the input x by the formula $\text{delta0} + (\text{delta1} - \text{delta0}) / \text{theta1} * \text{theta}$ where theta1 is the alternate hypothesis value of the standardized parameter. Note that this value will be exponentiated in the case of relative risks, hazard ratios or when the user specifies logdelta=TRUE. In the case of hazard ratios, the value is computed instead by gsHR() to be consistent with plot.gsDesign(). Similarly, the value is computed by gsRR() when the relative risk is the natural parameter.

Spending: Incremental error spending at each given analysis. For asymmetric designs, futility bound will have beta-spending summarized. Efficacy bound always has alpha-spending summarized.

B-value: $\sqrt{t} * Z$ where t is the proportion of information at the analysis divided by the final analysis planned information. The expected value for B-values is directly proportional to t.

CP: Conditional power under the estimated treatment difference assuming the interim Z-statistic is at the study bound

CP H1: Conditional power under the alternate hypothesis treatment effect assuming the interim test statistic is at the study bound.

PP: Predictive power assuming the interim test statistic is at the study bound and the input prior distribution for the standardized effect size. This is the conditional power averaged across the posterior distribution for the treatment effect given the interim test statistic value. $P\{\text{Cross if } \text{delta} = x\}$: For each of the parameter values in x, the probability of crossing either bound given that treatment effect is computed. This value is cumulative for each bound. For example, the probability of crossing the efficacy bound at or before the analysis of interest.

Usage

```
## S3 method for class 'gsDesign'
summary(object, information = FALSE, timeunit = "months", ...)

## S3 method for class 'gsDesign'
print(x, ...)

gsBoundSummary(
  x,
  deltaname = NULL,
  logdelta = FALSE,
  Nname = NULL,
  digits = 4,
  ddigits = 2,
  tdigits = 0,
  timename = "Month",
  prior = normalGrid(mu = x$delta/2, sigma = 10/sqrt(x$n.fix)),
  POS = FALSE,
  ratio = NULL,
  exclude = c("B-value", "Spending", "CP", "CP H1", "PP"),
```

```

    r = 18,
    alpha = NULL,
    ...
)

xprint(
  x,
  include.rownames = FALSE,
  hline.after = c(-1, which(x$Value == x[1, ]$Value) - 1, nrow(x)),
  ...
)

## S3 method for class 'gsBoundSummary'
print(x, row.names = FALSE, digits = 4, ...)

gsBValue(z, i, x, ylab = "B-value", ...)

gsDelta(z, i, x, ylab = NULL, ...)

gsRR(z, i, x, ratio = 1, ylab = "Approximate risk ratio", ...)

gsHR(z, i, x, ratio = 1, ylab = "Approximate hazard ratio", ...)

gsCPz(z, i, x, theta = NULL, ylab = NULL, ...)

```

Arguments

object	An item of class gsDesign or gsSurv
information	indicator of whether n.I in object represents statistical information rather than sample size or event counts.
timeunit	Text string with time units used for time-to-event designs created with gsSurv()
...	This allows many optional arguments that are standard when calling plot for gsBValue, gsDelta, gsHR, gsRR and gsCPz
x	An item of class gsDesign or gsSurv, except for print.gsBoundSummary() where x is an object created by gsBoundSummary() and xprint() which is used with xtable (see examples)
deltaname	Natural parameter name. If default NULL is used, routine will default to "HR" when class is gsSurv or if nFixSurv was input when creating x with gsDesign().
logdelta	Indicates whether natural parameter is the natural logarithm of the actual parameter. For example, the relative risk or odds-ratio would be put on the logarithmic scale since the asymptotic behavior is 'more normal' than a non-transformed value. As with deltaname, the default will be changed to true if x has class gsDesign or if nFixSurv>0 was input when x was created by gsDesign(); that is, the natural parameter for a time-to-event endpoint will be on the logarithmic scale.
Nname	This will normally be changed to "N" or, if a time-to-event endpoint is used, "Events". Other immediate possibility are "Deaths" or "Information".

digits	Number of digits past the decimal to be printed in the body of the table.
ddigits	Number of digits past the decimal to be printed for the natural parameter delta.
tdigits	Number of digits past the decimal point to be shown for estimated timing of each analysis.
timename	Text string indicating time unit.
prior	A prior distribution for the standardized effect size. Must be of the format produced by normalGrid(), but can reflect an arbitrary prior distribution. The default reflects a normal prior centered half-way between the null and alternate hypothesis with the variance being equivalent to the treatment effect estimate if 1 percent of the sample size for a fixed design were sampled. The prior is intended to be relatively uninformative. This input will only be applied if POS=TRUE is input.
POS	This is an indicator of whether or not probability of success (POS) should be estimated at baseline or at each interim based on the prior distribution input in prior. The prior probability of success before the trial starts is the power of the study averaged over the prior distribution for the standardized effect size. The POS after an interim analysis assumes the interim test statistic is an unknown value between the futility and efficacy bounds. Based on this, a posterior distribution for the standardized parameter is computed and the conditional power of the trial is averaged over this posterior distribution.
ratio	Sample size ratio assumed for experimental to control treatment group sample sizes. This only matters when x for a binomial or time-to-event endpoint where gsRR or gsHR are used for approximating the treatment effect if a test statistic falls on a study bound.
exclude	A list of test statistics to be excluded from design boundary summary produced; see details or examples for a list of all possible output values. A value of NULL produces all available summaries.
r	See gsDesign . This is an integer used to control the degree of accuracy of group sequential calculations which will normally not be changed.
alpha	If used, a vector of alternate alpha-levels to print boundaries for. Only works with test.type 1, 4, 6, 7, and 8. If specified, efficacy bound columns are headed by individual alpha levels. The alpha level of the input design is always included as the first column.
include.rownames	indicator of whether or not to include row names in output.
hline.after	table lines after which horizontal separation lines should be set; default is to put lines between each analysis as well as at the top and bottom of the table.
row.names	indicator of whether or not to print row names
z	A vector of z-statistics
i	A vector containing the analysis for each element in z; each element must be in 1 to x\$k, inclusive
ylab	Used when functions are passed to plot.gsDesign to establish default y-axis labels
theta	A scalar value representing the standardized effect size used for conditional power calculations; see gsDesign; if NULL, conditional power is computed at the estimated interim treatment effect based on z

Value

gsBValue(), gsDelta(), gsHR() and gsCPz() each returns a vector containing the B-values, approximate treatment effect (see details), approximate hazard ratio and conditional power, respectively, for each value specified by the interim test statistics in `z` at interim analyses specified in `i`.

`summary` returns a text string summarizing the design at a high level. This may be used with `gsBoundSummary` for a nicely formatted, concise group sequential design description.

`gsBoundSummary` returns a table in a data frame providing a variety of boundary characteristics. The tabular format makes formatting particularly amenable to place in documents either through direct creation of readable by Word (see the `rtf` package) or to a csv format readable by spreadsheet software using `write.csv`.

`print.gsDesign` prints an overall summary a group sequential design. While the design description is complete, the format is not as ‘document friendly’ as `gsBoundSummary`.

`print.gsBoundSummary` is a simple extension of `print.data.frame` intended for objects created with `gsBoundSummary`. The only extension is to make the default to not print row names. This is probably ‘not good R style’ but may be helpful for many lazy R programmers like the author.

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

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References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

[gsDesign](#), [plot.gsDesign](#), [gsProbability](#), [xtable.gsSurv](#)

Examples

```
library(ggplot2)
# survival endpoint using gsSurv
# generally preferred over nSurv since time computations are shown
xgs <- gsSurv(lambdaC = .2, hr = .5, eta = .1, T = 2, minfup = 1.5)
gsBoundSummary(xgs, timename = "Year", tdigits = 1)
summary(xgs)

# survival endpoint using nSurvival
# NOTE: generally recommend gsSurv above for this!
ss <- nSurvival(
  lambda1 = .2, lambda2 = .1, eta = .1, Ts = 2, Tr = .5,
  sided = 1, alpha = .025, ratio = 2
)
```

```

xs <- gsDesign(nFixSurv = ss$n, n.fix = ss$nEvents, delta1 = log(ss$lambda2 / ss$lambda1))
gsBoundSummary(xs, logdelta = TRUE, ratio = ss$ratio)
# generate some of the above summary statistics for the upper bound
z <- xs$upper$bound
# B-values
gsBValue(z = z, i = 1:3, x = xs)
# hazard ratio
gsHR(z = z, i = 1:3, x = xs)
# conditional power at observed treatment effect
gsCPz(z = z[1:2], i = 1:2, x = xs)
# conditional power at H1 treatment effect
gsCPz(z = z[1:2], i = 1:2, x = xs, theta = xs$delta)

# information-based design
xinfo <- gsDesign(delta = .3, delta1 = .3)
gsBoundSummary(xinfo, Nname = "Information")

# show all available boundary descriptions
gsBoundSummary(xinfo, Nname = "Information", exclude = NULL)

# add intermediate parameter value
xinfo <- gsProbability(d = xinfo, theta = c(0, .15, .3))
class(xinfo) # note this is still as gsDesign class object
gsBoundSummary(xinfo, Nname = "Information")

# now look at a binomial endpoint; specify H0 treatment difference as p1-p2=.05
# now treatment effect at bound (say, thetahat) is transformed to
#  $xp\$delta0 + xp\$delta1 * (thetahat - xp\$delta0) / xp\$delta$ 
np <- nBinomial(p1 = .15, p2 = .10)
xp <- gsDesign(n.fix = np, endpoint = "Binomial", delta1 = .05)
summary(xp)
gsBoundSummary(xp, deltaname = "p[C]-p[E]")
# estimate treatment effect at lower bound
# by setting delta0=0 (default) and delta1 above in gsDesign
# treatment effect at bounds is scaled to these differences
# in this case, this is the difference in event rates
gsDelta(z = xp$lower$bound, i = 1:3, xp)

# binomial endpoint with risk ratio estimates
n.fix <- nBinomial(p1 = .3, p2 = .15, scale = "RR")
xrr <- gsDesign(k = 2, n.fix = n.fix, delta1 = log(.15 / .3), endpoint = "Binomial")
gsBoundSummary(xrr, deltaname = "RR", logdelta = TRUE)
gsRR(z = xp$lower$bound, i = 1:3, xrr)
plot(xrr, plottype = "RR")

# delta is odds-ratio: sample size slightly smaller than for relative risk or risk difference
n.fix <- nBinomial(p1 = .3, p2 = .15, scale = "OR")
xOR <- gsDesign(k = 2, n.fix = n.fix, delta1 = log(.15 / .3 / .85 * .7), endpoint = "Binomial")
gsBoundSummary(xOR, deltaname = "OR", logdelta = TRUE)

# for nice LaTeX table output, use xprint
xprint(xtable::xtable(gsBoundSummary(xOR, deltaname = "OR", logdelta = TRUE),
  caption = "Table caption.")

```

```
))
```

summary.spendfn	<i>Spending Function</i>
-----------------	--------------------------

Description

Spending Function

Usage

```
## S3 method for class 'spendfn'
summary(object, ...)

spendingFunction(alpha, t, param)
```

Arguments

object	A spendfn object to be summarized.
...	Not currently used.
alpha	Real value > 0 and no more than 1. Defaults in calls to <code>gsDesign()</code> are $\alpha=0.025$ for one-sided Type I error specification and $\alpha=0.1$ for Type II error specification. However, this could be set to 1 if, for descriptive purposes, you wish to see the proportion of spending as a function of the proportion of sample size/information.
t	A vector of points with increasing values from 0 to 1, inclusive. Values of the proportion of sample size/information for which the spending function will be computed.
param	A single real value or a vector of real values specifying the spending function parameter(s); this must be appropriately matched to the spending function specified.

Value

`spendingFunction` and spending functions in general produce an object of type `spendfn`.

name	A character string with the name of the spending function.
param	any parameters used for the spending function.
parname	a character string or strings with the name(s) of the parameter(s) in <code>param</code> .
sf	the spending function specified.
spend	a vector of cumulative spending values corresponding to the input values in <code>t</code> .
bound	this is null when returned from the spending function, but is set in <code>gsDesign()</code> if the spending function is called from there. Contains z-values for bounds of a design.
prob	this is null when returned from the spending function, but is set in <code>gsDesign()</code> if the spending function is called from there. Contains probabilities of boundary crossing at i-th analysis for j-th theta value input to <code>gsDesign()</code> in <code>prob[i, j]</code> .

Note

The gsDesign technical manual is available at <https://keaven.github.io/gsd-tech-manual/>.

Author(s)

Keaven Anderson <keaven_anderson@merck.com>

References

Jennison C and Turnbull BW (2000), *Group Sequential Methods with Applications to Clinical Trials*. Boca Raton: Chapman and Hall.

See Also

[gsDesign](#), [sfHSD](#), [sfPower](#), [sfLogistic](#), [sfExponential](#), [sfTruncated](#), [vignette\("gsDesignPackageOverview"\)](#)

Examples

```
# Example 1: simple example showing what most users need to know

# Design a 4-analysis trial using a Hwang-Shih-DeCani spending function
# for both lower and upper bounds
x <- gsDesign(k = 4, sfu = sfHSD, sfupar = -2, sfl = sfHSD, sflpar = 1)

# Print the design
x
# Summarize the spending functions
summary(x$upper)
summary(x$lower)

# Plot the alpha- and beta-spending functions
plot(x, plottype = 5)

# What happens to summary if we used a boundary function design
x <- gsDesign(test.type = 2, sfu = "OF")
y <- gsDesign(test.type = 1, sfu = "WT", sfupar = .25)
summary(x$upper)
summary(y$upper)

# Example 2: advanced example: writing a new spending function
# Most users may ignore this!

# Implementation of 2-parameter version of
# beta distribution spending function
# assumes t and alpha are appropriately specified (does not check!)
sfbdist <- function(alpha, t, param) {
  # Check inputs
  checkVector(param, "numeric", c(0, Inf), c(FALSE, TRUE))
  if (length(param) != 2) {
    stop(
      "b-dist example spending function parameter must be of length 2"
    )
  }
}
```

```

}

# Set spending using cumulative beta distribution and return
x <- list(
  name = "B-dist example", param = param, parname = c("a", "b"),
  sf = sfbdist, spend = alpha *
    pbeta(t, param[1], param[2]), bound = NULL, prob = NULL
)

class(x) <- "spendfn"

x
}

# Now try it out!
# Plot some example beta (lower bound) spending functions using
# the beta distribution spending function
t <- 0:100 / 100
plot(
  t, sfbdist(1, t, c(2, 1))$spend,
  type = "l",
  xlab = "Proportion of information",
  ylab = "Cumulative proportion of total spending",
  main = "Beta distribution Spending Function Example"
)
lines(t, sfbdist(1, t, c(6, 4))$spend, lty = 2)
lines(t, sfbdist(1, t, c(.5, .5))$spend, lty = 3)
lines(t, sfbdist(1, t, c(.6, 2))$spend, lty = 4)
legend(
  x = c(.65, 1), y = 1 * c(0, .25), lty = 1:4,
  legend = c("a=2, b=1", "a=6, b=4", "a=0.5, b=0.5", "a=0.6, b=2")
)

```

tEventsIA

Advanced time-to-event sample size calculation

Description

nSurv() is used to calculate the sample size for a two-arm clinical trial with a time-to-event endpoint under the assumption of proportional hazards. The default method assumes a fixed enrollment duration and fixed trial duration; in this case the required sample size is achieved by increasing enrollment rates. nSurv() implements the Lachin and Foulkes (1986) method as default. Schoenfeld (1981), Freedman (1982), and Bernstein and Lagakos (1989) methods are also supported; see Details. gsSurv() combines nSurv() with gsDesign() to derive a group sequential design for a study with a time-to-event endpoint.

Usage

```
tEventsIA(x, timing = 0.25, tol = .Machine$double.eps^0.25)
```

```

nEventsIA(tIA = 5, x = NULL, target = 0, simple = TRUE)

nSurv(
  lambdaC = log(2)/6,
  hr = 0.6,
  hr0 = 1,
  eta = 0,
  etaE = NULL,
  gamma = 1,
  R = 12,
  S = NULL,
  T = 18,
  minfup = 6,
  ratio = 1,
  alpha = 0.025,
  beta = 0.1,
  sided = 1,
  tol = .Machine$double.eps^0.25,
  method = c("LachinFoulkes", "Schoenfeld", "Freedman", "BernsteinLagakos")
)

## S3 method for class 'nSurv'
print(x, digits = 3, show_strata = TRUE, ...)

## S3 method for class 'gsSurv'
xtable(
  x,
  caption = NULL,
  label = NULL,
  align = NULL,
  digits = NULL,
  display = NULL,
  auto = FALSE,
  footnote = NULL,
  fnwid = "9cm",
  timename = "months",
  ...
)

gsSurv(
  k = 3,
  test.type = 4,
  alpha = 0.025,
  sided = 1,
  beta = 0.1,
  astar = 0,
  timing = 1,
  sfu = sfHSD,

```

```

sfupar = -4,
sfl = sfHSD,
sflpar = -2,
sfharm = sfHSD,
sfharmparam = -2,
r = 18,
lambdaC = log(2)/6,
hr = 0.6,
hr0 = 1,
eta = 0,
etaE = NULL,
gamma = 1,
R = 12,
S = NULL,
T = 18,
minfup = 6,
ratio = 1,
tol = .Machine$double.eps^0.25,
usTime = NULL,
lsTime = NULL,
testUpper = TRUE,
testLower = TRUE,
testHarm = TRUE,
method = c("LachinFoulkes", "Schoenfeld", "Freedman", "BernsteinLagakos")
)

## S3 method for class 'gsSurv'
print(x, digits = 3, show_gsDesign = FALSE, show_strata = TRUE, ...)

```

Arguments

x	An object of class nSurv or gsSurv. <code>print.nSurv()</code> is used for an object of class nSurv which will generally be output from <code>nSurv()</code> . For <code>print.gsSurv()</code> is used for an object of class gsSurv which will generally be output from <code>gsSurv()</code> . <code>nEventsIA</code> and <code>tEventsIA</code> operate on both the nSurv and gsSurv class.
timing	Sets relative timing of interim analyses in gsSurv. Default of 1 produces equally spaced analyses. Otherwise, this is a vector of length k or k-1. The values should satisfy $0 < \text{timing}[1] < \text{timing}[2] < \dots < \text{timing}[k-1] < \text{timing}[k]=1$. For <code>tEventsIA</code> , this is a scalar strictly between 0 and 1 that indicates the targeted proportion of final planned events available at an interim analysis.
tol	For cases when T or minfup values are derived through root finding (T or minfup input as NULL), tol provides the level of error input to the <code>uniroot()</code> root-finding function. The default is the same as for uniroot .
tIA	Timing of an interim analysis; should be between 0 and y\$T.
target	The targeted proportion of events at an interim analysis. This is used for root-finding will be 0 for normal use.
simple	See output specification for <code>nEventsIA()</code> .

lambdaC	Scalar, vector or matrix of event hazard rates for the control group; rows represent time periods while columns represent strata; a vector implies a single stratum. Note that rates corresponding the final time period are extended indefinitely.
hr	Hazard ratio (experimental/control) under the alternate hypothesis (scalar, > 0 , must differ from hr_0). Both $hr < hr_0$ (experimental is beneficial when lower hazard is better) and $hr > hr_0$ (e.g., time-to-response or safety designs) are supported.
hr0	Hazard ratio (experimental/control) under the null hypothesis (scalar, > 0 , must differ from hr).
eta	Scalar, vector or matrix of dropout hazard rates for the control group; rows represent time periods while columns represent strata; if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
etaE	Matrix dropout hazard rates for the experimental group specified in like form as eta; if NULL, this is set equal to eta.
gamma	A scalar, vector or matrix of rates of entry by time period (rows) and strata (columns); if entered as a scalar, rate is constant across strata and time periods; if entered as a vector, rates are constant across strata.
R	A scalar or vector of durations of time periods for recruitment rates specified in rows of gamma. Length is the same as number of rows in gamma. Note that when variable enrollment duration is specified (input $T=$ NULL), the final enrollment period is extended as long as needed; otherwise enrollment after $\text{sum}(R)$ is 0.
S	A scalar or vector of durations of piecewise constant event rates specified in rows of lambda, eta and etaE; this is NULL if there is a single event rate per stratum (exponential failure) or length of the number of rows in lambda minus 1, otherwise. The final time period is extended indefinitely for each stratum.
T	Study duration; if T is input as NULL, this will be computed on output; see details.
minfup	Follow-up of last patient enrolled; if minfup is input as NULL, this will be computed on output; see details.
ratio	Randomization ratio of experimental treatment divided by control; normally a scalar, but may be a vector with length equal to number of strata.
alpha	Type I error rate. Default is 0.025 since 1-sided testing is default.
beta	Type II error rate. Default is 0.10 (90% power); NULL if power is to be computed based on other input values.
sided	1 for 1-sided testing, 2 for 2-sided testing.
method	One of "LachinFoulkes" (default), "Schoenfeld", "Freedman", or "BernsteinLagakos". Note: "Schoenfeld" and "Freedman" methods only support superiority testing ($hr_0 = 1$). "Freedman" does not support stratified populations.
digits	Number of digits past the decimal place to print (<code>print.gsSurv()</code>); also a pass through to generic <code>xtable()</code> from <code>xtable.gsSurv()</code> .
show_strata	Logical; for <code>print.gsSurv()</code> , include strata summaries.
...	Other arguments that may be passed to generic functions underlying the methods here.

caption	Passed through to generic xtable().
label	Passed through to generic xtable().
align	Passed through to generic xtable().
display	Passed through to generic xtable().
auto	Passed through to generic xtable().
footnote	Footnote for xtable output; may be useful for describing some of the design parameters.
fnwid	A text string controlling the width of footnote text at the bottom of the xtable output.
timename	Character string with plural of time units (e.g., "months")
k	Number of analyses planned, including interim and final.
test.type	1=one-sided 2=two-sided symmetric 3=two-sided, asymmetric, beta-spending with binding lower bound 4=two-sided, asymmetric, beta-spending with non-binding lower bound 5=two-sided, asymmetric, lower bound spending under the null hypothesis with binding lower bound 6=two-sided, asymmetric, lower bound spending under the null hypothesis with non-binding lower bound 7=two-sided, asymmetric, with binding futility and binding harm bounds 8=two-sided, asymmetric, with non-binding futility and non-binding harm bounds. See details, examples and manual.
astar	Total spending for the lower (test.type 5 or 6) or harm (test.type 7 or 8) bound under the null hypothesis. Default is 0. For test.type 5 or 6, astar specifies the total probability of crossing a lower bound at all analyses combined. For test.type 7 or 8, astar specifies the total probability of crossing the harm bound at all analyses combined under the null hypothesis. If astar = 0, it will be changed to 1−alpha.
sfu	A spending function or a character string indicating a boundary type (that is, “WT” for Wang-Tsiatis bounds, “OF” for O’Brien-Fleming bounds and “Pocock” for Pocock bounds). For one-sided and symmetric two-sided testing is used to completely specify spending (test.type=1, 2), sfu. The default value is sfHSD which is a Hwang-Shih-DeCani spending function. See details, vignette("SpendingFunctionOverview") manual and examples.
sfupar	Real value, default is −4 which is an O’Brien-Fleming-like conservative bound when used with the default Hwang-Shih-DeCani spending function. This is a real-vector for many spending functions. The parameter sfupar specifies any parameters needed for the spending function specified by sfu; this is not needed for spending functions (sfLDOF, sfLDPocock) or bound types (“OF”, “Pocock”) that do not require parameters. Note that sfupar can be specified as a positive scalar for sfLDOF for a generalized O’Brien-Fleming spending function.
sf1	Specifies the spending function for lower boundary crossing probabilities when asymmetric, two-sided testing is performed (test.type = 3, 4, 5, or 6). Unlike the upper bound, only spending functions are used to specify the lower bound. The default value is sfHSD which is a Hwang-Shih-DeCani spending function.

	The parameter <code>sfl</code> is ignored for one-sided testing (<code>test.type=1</code>) or symmetric 2-sided testing (<code>test.type=2</code>). See details, spending functions, manual and examples.
<code>sflpar</code>	Real value, default is -2 , which, with the default Hwang-Shih-DeCani spending function, specifies a less conservative spending rate than the default for the upper bound.
<code>sfharm</code>	A spending function for the harm bound, used with <code>test.type = 7</code> or <code>test.type = 8</code> . Default is <code>sfHSD</code> . See spendingFunction for details.
<code>sfharmparam</code>	Real value, default is -2 . Parameter for the harm bound spending function <code>sfharm</code> .
<code>r</code>	Integer value (≥ 1 and ≤ 80) controlling the number of numerical integration grid points. Default is 18, as recommended by Jennison and Turnbull (2000). Grid points are spread out in the tails for accurate probability calculations. Larger values provide more grid points and greater accuracy but slow down computation. Jennison and Turnbull (p. 350) note an accuracy of 10^{-6} with $r = 16$. This parameter is normally not changed by users.
<code>usTime</code>	Default is NULL in which case upper bound spending time is determined by timing. Otherwise, this should be a vector of length k with the spending time at each analysis (see Details section of gsDesign).
<code>lsTime</code>	Default is NULL in which case lower bound spending time is determined by timing. Otherwise, this should be a vector of length k with the spending time at each analysis (see Details section of gsDesign).
<code>testUpper</code>	Indicator of which analyses should include an upper (efficacy) bound. A single value of TRUE (default) indicates all analyses have an efficacy bound. Otherwise, a logical vector of length k indicating which analyses will have an efficacy bound. Overridden to all TRUE for <code>test.type 1</code> and <code>2</code> . Must be TRUE at the final analysis to achieve targeted power. At each analysis, at least one of <code>testUpper</code> , <code>testLower</code> , or <code>testHarm</code> must be TRUE. Where <code>testUpper</code> is FALSE, the upper bound is set to $+20$ (effectively Inf) and displayed as NA in output.
<code>testLower</code>	Indicator of which analyses should include a lower (futility) bound. A single value of TRUE (default) indicates all analyses have a lower bound; FALSE indicates none. Otherwise, a logical vector of length k . Ignored for <code>test.type 1</code> (one-sided, no lower bound). Overridden to all TRUE for <code>test.type 2</code> (symmetric). For <code>test.type 3–8</code> , at least one analysis must be TRUE. Where <code>testLower</code> is FALSE, the lower bound is set to -20 (effectively $-\text{Inf}$) and displayed as NA in output.
<code>testHarm</code>	Indicator of which analyses should include a harm bound. A single value of TRUE (default) indicates all analyses have a harm bound; FALSE indicates none. Otherwise, a logical vector of length k . Only used for <code>test.type 7</code> or <code>8</code> ; at least one analysis must be TRUE for those types. Where <code>testHarm</code> is FALSE, the harm bound is set to -20 (effectively $-\text{Inf}$) and displayed as NA in output.
<code>show_gsDesign</code>	Logical; for <code>print.gsSurv()</code> , include <code>gsDesign</code> details.

Details

The Lachin and Foulkes method uses both null and alternate hypothesis variances to derive sample size and is extended here to support non-inferiority, super-superiority, and stratified designs. As an

alternative, the Kim and Tsiatis (1990) method can be used with fixed enrollment rates and either fixed enrollment duration or fixed minimum follow-up. The Schoenfeld approach uses the asymptotic distribution of the log-rank statistic under the assumption of proportional hazards and local alternatives (i.e., $\log(HR)$ is small). The Freedman approach uses the same asymptotic distribution and, like the Schoenfeld approach, uses just the null hypothesis variance to derive sample size. The Bernstein and Lagakos (1989) approach was derived to compute sample size for a stratified model with a common proportional hazards assumption across strata. Like the Lachin and Foulkes (1986) method, it uses both null and alternate hypothesis variances to compute sample size; however, the null hypothesis variance is computed differently. The Bernstein and Lagakos (1989) approach uses the alternate hypothesis failure rate assumptions for both the control and experimental groups, while the Lachin and Foulkes method uses null hypothesis rates that average the alternate hypothesis failure rates to get similar numbers of expected events under the null and alternate hypotheses. Since the Lachin and Foulkes method has fewer events under the null hypothesis (less statistical information), it calculates less power than the Bernstein and Lagakos method. Piecewise exponential survival is supported as well as piecewise constant enrollment and dropout rates. The methods are for a 2-arm trial with treatment groups referred to as experimental and control. A stratified population is allowed as in Lachin and Foulkes (1986); this method has been extended to derive non-inferiority as well as superiority trials. Stratification also allows power calculation for meta-analyses.

`print()`, `xtable()` and `summary()` methods are provided to operate on the returned value from `gsSurv()`, an object of class `gsSurv`. `print()` is also extended to `nSurv` objects. The functions `gsBoundSummary` (data frame for tabular output), `xprint` (application of `xtable` for tabular output) and `summary.gsSurv` (textual summary of `gsDesign` or `gsSurv` object) may be preferred summary functions; see example in vignettes. See also `gsBoundSummary` for output of tabular summaries of bounds for designs produced by `gsSurv()`.

Both `nEventsIA` and `tEventsIA` require a group sequential design for a time-to-event endpoint of class `gsSurv` as input. `nEventsIA` calculates the expected number of events under the alternate hypothesis at a given interim time. `tEventsIA` calculates the time that the expected number of events under the alternate hypothesis is a given proportion of the total events planned for the final analysis.

`nSurv()` produces an object of class `nSurv` with the number of subjects and events for a set of pre-specified trial parameters, such as accrual duration and follow-up period. The underlying power calculation is based on Lachin and Foulkes (1986) method for proportional hazards assuming a fixed underlying hazard ratio between 2 treatment groups. The method has been extended here to enable designs to test non-inferiority. Piecewise constant enrollment and failure rates are assumed and a stratified population is allowed. See also `nSurvival` for other Lachin and Foulkes (1986) methods assuming a constant hazard difference or exponential enrollment rate.

When study duration (T) and follow-up duration (minfup) are fixed, `nSurv` applies exactly the Lachin and Foulkes (1986) method of computing sample size under the proportional hazards assumption. For this computation, enrollment rates are altered proportionately to those input in `gamma` to achieve the power of interest.

Given the specified enrollment rate(s) input in `gamma`, `nSurv` may also be used to derive enrollment duration required for a trial to have defined power if T is input as `NULL`; in this case, both R (enrollment duration for each specified enrollment rate) and T (study duration) will be computed on output.

Alternatively and also using the fixed enrollment rate(s) in `gamma`, if minimum follow-up `minfup` is specified as `NULL`, then the enrollment duration(s) specified in R are considered fixed and `minfup` and T are computed to derive the desired power. This method will fail if the specified enrollment

rates and durations either over-powers the trial with no additional follow-up or underpowers the trial with infinite follow-up. This method produces a corresponding error message in such cases. For methods other than Lachin and Foulkes, these fixed-rate duration solves use the selected method for the fixed-design event calculation.

The input to `gsSurv` is a combination of the input to `nSurv()` and `gsDesign()`. When `T = NULL` and `minfup` is specified, `gsSurv()` preserves the input accrual rate and minimum follow-up, applies the group sequential design, and solves the accrual duration needed for the final planned number of events. When both `T` and `minfup` are `NULL`, `gsSurv()` preserves the input accrual rate and duration, applies the group sequential design, and solves the follow-up duration needed for the final planned number of events.

`nEventsIA()` is provided to compute the expected number of events at a given point in time given enrollment, event and censoring rates. The routine is used with a root finding routine to approximate the approximate timing of an interim analysis. It is also used to extend enrollment or follow-up of a fixed design to obtain a sufficient number of events to power a group sequential design.

Value

`nSurv()` returns an object of type `nSurv` with the following components:

<code>alpha</code>	As input.
<code>sided</code>	As input.
<code>beta</code>	Type II error; if missing, this is computed.
<code>power</code>	Power corresponding to input <code>beta</code> or computed if output <code>beta</code> is computed.
<code>lambdaC</code>	As input.
<code>etaC</code>	As input.
<code>etaE</code>	As input.
<code>gamma</code>	As input unless none of the following are <code>NULL</code> : <code>T</code> , <code>minfup</code> , <code>beta</code> ; otherwise, this is a constant times the input value required to power the trial given the other input variables.
<code>ratio</code>	As input.
<code>R</code>	As input unless <code>T</code> was <code>NULL</code> on input.
<code>S</code>	As input.
<code>T</code>	As input.
<code>minfup</code>	As input.
<code>hr</code>	As input.
<code>hr0</code>	As input.
<code>n</code>	Total expected sample size corresponding to output accrual rates and durations.
<code>d</code>	Total expected number of events under the alternate hypothesis.
<code>tol</code>	As input, except when not used in computations in which case this is returned as <code>NULL</code> . This and the remaining output below are not printed by the <code>print()</code> extension for the <code>nSurv</code> class.
<code>eDC</code>	A vector of expected number of events by stratum in the control group under the alternate hypothesis.

eDE	A vector of expected number of events by stratum in the experimental group under the alternate hypothesis.
eDC0	A vector of expected number of events by stratum in the control group under the null hypothesis.
eDE0	A vector of expected number of events by stratum in the experimental group under the null hypothesis.
eNC	A vector of the expected accrual in each stratum in the control group.
eNE	A vector of the expected accrual in each stratum in the experimental group.
variable	A text string equal to "Accrual rate" if a design was derived by varying the accrual rate, "Accrual duration" if a design was derived by varying the accrual duration, "Follow-up duration" if a design was derived by varying follow-up duration, or "Power" if accrual rates and duration as well as follow-up duration was specified and beta=NULL was input.

gsSurv() returns much of the above plus an object of class gsDesign in a variable named gs; see [gsDesign](#) for general documentation on what is returned in gs. The value of gs\$n.I represents the number of endpoints required at each analysis to adequately power the trial. Other items returned by gsSurv() are:

gs	A group sequential design (gsDesign) object.
lambdaC	As input.
etaC	As input.
etaE	As input.
gamma	As input unless none of the following are NULL: T, minfup, beta; otherwise, this is a constant times the input value required to power the trial given the other input variables.
ratio	As input.
R	As input unless T was NULL on input.
S	As input.
T	As input.
minfup	As input.
hr	As input.
hr0	As input.
eNC	Total expected sample size corresponding to output accrual rates and durations.
eNE	Total expected sample size corresponding to output accrual rates and durations.
eDC	Total expected number of events under the alternate hypothesis.
eDE	Total expected number of events under the alternate hypothesis.
tol	As input, except when not used in computations in which case this is returned as NULL. This and the remaining output below are not printed by the print() extension for the nSurv class.
eDC	A vector of expected number of events by stratum in the control group under the alternate hypothesis.

eDE	A vector of expected number of events by stratum in the experimental group under the alternate hypothesis.
eDC0	A vector of expected number of events by stratum in the control group under the null hypothesis.
eDE0	A vector of expected number of events by stratum in the experimental group under the null hypothesis.
eNC	A vector of the expected accrual in each stratum in the control group.
eNE	A vector of the expected accrual in each stratum in the experimental group.
variable	A text string equal to "Accrual rate" if a design was derived by varying the accrual rate, "Accrual duration" if a design was derived by varying the accrual duration, "Follow-up duration" if a design was derived by varying follow-up duration, or "Power" if accrual rates and duration as well as follow-up duration was specified and beta=NULL was input.

nEventsIA() returns the expected proportion of the final planned events observed at the input analysis time minus target when simple=TRUE. When simple=FALSE, nEventsIA returns a list with following components:

T	The input value tIA.
eDC	The expected number of events in the control group at time the output time T.
eDE	The expected number of events in the experimental group at the output time T.
eNC	The expected enrollment in the control group at the output time T.
eNE	The expected enrollment in the experimental group at the output time T.

tEventsIA() returns the same structure as nEventsIA(..., simple=TRUE) when

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References

- Kim KM and Tsiatis AA (1990), Study duration for clinical trials with survival response and early stopping rule. *Biometrics*, 46, 81-92
- Lachin JM and Foulkes MA (1986), Evaluation of Sample Size and Power for Analyses of Survival with Allowance for Nonuniform Patient Entry, Losses to Follow-Up, Noncompliance, and Stratification. *Biometrics*, 42, 507-519.
- Schoenfeld D (1981), The Asymptotic Properties of Nonparametric Tests for Comparing Survival Distributions. *Biometrika*, 68, 316-319.
- Freedman LS (1982), Tables of the Number of Patients Required in Clinical Trials Using the Log-rank Test. *Statistics in Medicine*, 1, 121-129.

See Also[uniroot](#)

[vignette\("gsSurvBasicExamples", package = "gsDesign"\)](#) for basic survival sample size examples, [vignette\("SurvivalOverview", package = "gsDesign"\)](#) for method background, and [vignette\("SeqDesignSurvival", package = "gsDesign"\)](#) for a SAS PROC SEQDESIGN translation example.

[gsBoundSummary](#), [xprint](#), [gsSurvCalendar](#), [gsSurvPower](#), [gsDesign-package](#), [plot.gsDesign](#), [gsDesign](#), [gsHR](#), [nSurvival](#)

[Normal xtable](#)**Examples**

```
# Vary accrual rate gamma to obtain power
# T, minfup and R all specified, although R will be adjusted on output
# gamma as input will be multiplied in output to achieve desired power
# Default method is Lachin and Foulkes
x_nsurv <- nSurv(
  lambdaC = log(2) / 6, R = 10, hr = .5, eta = .001, gamma = 1,
  alpha = 0.02, beta = .15, T = 36, minfup = 12, method = "LachinFoulkes"
)
# Demonstrate print method
print(x_nsurv)
# Same assumptions for group sequential design
x_gs <- gsSurv(
  k = 4, sfl = gsDesign::sfPower, sflpar = .5, lambdaC = log(2) / 6, hr = .5,
  eta = .001, gamma = 1, T = 36, minfup = 12, method = "LachinFoulkes"
)
print(x_gs)
# Demonstrate xtable method for gsSurv
print(xtable::xtable(x_gs,
  footnote = "This is a footnote; note that it can be wide.",
  caption = "Caption example for xtable output."
))
# Demonstrate nEventsIA method
# find expected number of events at time 12 in the above trial
nEventsIA(x = x_gs, tIA = 10)

# find time at which 1/4 of events are expected
tEventsIA(x = x_gs, timing = .25)

# Adjust accrual duration R to achieve desired power
# Trial duration T input as NULL and will be computed based on
# accrual duration R and minimum follow-up duration minfup
# Minimum follow-up duration minfup is specified
# We use the Schoenfeld method to compute accrual duration R
# Control median survival time is 6
nSurv(
  lambdaC = log(2) / 6, hr = .5, eta = .001, gamma = 6,
  alpha = .025, beta = .1, minfup = 12, T = NULL, method = "Schoenfeld"
)
```

```

# Same assumptions for group sequential design
gsSurv(
  k = 4, sfu = gsDesign::sfHSD, sfupar = -4, sfl = gsDesign::sfPower, sflpar = .5,
  lambdaC = log(2) / 6, hr = .5, eta = .001, gamma = 6,
  T = 36, minfup = 12, method = "Schoenfeld"
) |>
  print()

# Vary minimum follow-up duration minfup to obtain power
# Accrual duration R rate gamma are fixed and will not change on output.
# Trial duration T and minimum follow-up minfup are input as NULL
# and will be computed on output.
# We will use the Freedman method to compute sample size
# Control median survival time is 6
# Often this method will fail as the accrual duration and rate provide too
# little or too much sample size.
nSurv(
  lambdaC = log(2) / 6, hr = .5, eta = .001, gamma = 6, R = 25,
  alpha = .025, beta = .1, minfup = NULL, T = NULL, method = "Freedman"
)
# Same assumptions for group sequential design
gsSurv(
  k = 4, sfu = gsDesign::sfHSD, sfupar = -4, sfl = gsDesign::sfPower, sflpar = .5,
  lambdaC = log(2) / 6, hr = .5, eta = .001, gamma = 6,
  T = 36, minfup = 12, method = "Freedman"
) |>
  print()

# piecewise constant enrollment rates (vary accrual rate to achieve power)
# also piecewise constant failure rates
# will specify annualized enrollment and failure rates
nSurv(
  lambdaC = -log(c(.95, .97, .98)), # 5%, 3% and 2% annual failure rates
  S = c(1, 1), # 1 year duration for first 2 failure rates, 3rd continues indefinitely
  R = c(.25, .25, 1.5), # 2-year enrollment with ramp-up over first 1/2 year
  gamma = c(1, 3, 6), # 1, 3 and 6 annualized enrollment rates will be
  # multiplied by ratio to achieve desired power
  hr = .5, eta = -log(1 - .01), # 1% annual censoring rate
  minfup = 3, T = 5, # 5-year trial duration with 3-year minimum follow-up
  alpha = .025, beta = .1, method = "LachinFoulkes"
)
# Same assumptions for group sequential design
gsSurv(
  k = 4, sfu = gsDesign::sfHSD, sfupar = -4, sfl = gsDesign::sfPower, sflpar = .5,
  lambdaC = -log(c(.95, .97, .98)), # 5%, 3% and 2% annual failure rates
  S = c(1, 1), # 1 year duration for first 2 failure rates, 3rd continues indefinitely
  R = c(.25, .25, 1.5), # 2-year enrollment with ramp-up over first 1/2 year
  gamma = c(1, 3, 6), # 1, 3 and 6 annualized enrollment rates will be
  # multiplied by ratio to achieve desired power
  hr = .5, eta = -log(1 - .01), # 1% annual censoring rate
  minfup = 3, T = 5, # 5-year trial duration with 3-year minimum follow-up
  alpha = .025, beta = .1, method = "LachinFoulkes"
) |>

```

```

print()

# combine it all: 2 strata, 2 failure rate periods
# Note that method = "LachinFoulkes" may be preferred here
nSurv(
  lambdaC = matrix(log(2) / c(6, 12, 18, 24), ncol = 2), hr = .5,
  eta = matrix(log(2) / c(40, 50, 45, 55), ncol = 2), S = 3,
  gamma = matrix(c(3, 6, 5, 7), ncol = 2), R = c(5, 10), T = 27, minfup = 12,
  alpha = .025, beta = .1, method = "BernsteinLagakos"
)
# Same assumptions for group sequential design
gsSurv(
  k = 4, sfu = gsDesign::sfHSD, sfupar = -4, sfl = gsDesign::sfPower, sflpar = .5,
  lambdaC = matrix(log(2) / c(6, 12, 18, 24), ncol = 2), hr = .5,
  eta = matrix(log(2) / c(40, 50, 45, 55), ncol = 2), S = 3,
  gamma = matrix(c(3, 6, 5, 7), ncol = 2), R = c(5, 10), T = 27, minfup = 12,
  alpha = .025, beta = .1, method = "BernsteinLagakos"
) |>
print()

# Example to compute power for a fixed design.
# Trial duration T, minimum follow-up minfup and accrual duration R are all
# specified and will not change on output.
# beta=NULL will compute power and output will be the same as if beta were specified.
# This option is not available for group sequential designs.
nSurv(
  lambdaC = log(2) / 6, hr = .5, eta = .001, gamma = 6, R = 25,
  alpha = .025, beta = NULL, minfup = 12, T = 36, method = "LachinFoulkes"
) |>
print()

```

toBinomialExact

Translate survival design bounds to exact binomial bounds

Description

Translate survival design bounds to exact binomial bounds

Usage

```

toBinomialExact(
  x,
  observedEvents = NULL,
  alpha = NULL,
  usTime = NULL,
  lsTime = NULL,
  maxSpend = FALSE
)

```

Arguments

<code>x</code>	An object of class <code>gsSurv</code> ; i.e., an object generated by the <code>gsSurv()</code> function.
<code>observedEvents</code>	If <code>NULL</code> (default), targeted timing of analyses will come from <code>x\$n.I</code> . Otherwise, this should be vector of increasing positive integers with at most 1 value $\geq x\$n.Iplan$ and of length at least 2. Only one value can be greater than or equal to <code>x\$maxn.Iplan</code> . This determines the case count at each analysis performed. Primarily, this is used for updating a design at the time of analysis.
<code>alpha</code>	Optional alpha level for deriving updated exact efficacy bounds. If <code>NULL</code> , the alpha level from <code>x</code> is used.
<code>usTime</code>	Optional upper spending-time override (length <code>k</code> or <code>k - 1</code> , with final value appended as 1 if needed). If <code>NULL</code> , this defaults to <code>observedEvents / x\$maxn.Iplan</code> (capped at 1) when <code>observedEvents</code> is supplied, or to the planned design timing otherwise.
<code>lsTime</code>	Optional lower spending-time override for <code>test.type = 4</code> (same length and monotonicity requirements as <code>usTime</code>). If <code>NULL</code> , it defaults to <code>usTime</code> .
<code>maxSpend</code>	Logical scalar. If <code>'TRUE'</code> , force full alpha spending (and, for <code>'test.type = 4'</code> , full beta spending) at the final analysis even when <code>'observedEvents[k] < x\$maxn.Iplan'</code> . This keeps earlier analysis spending unchanged and applies the override only at the last look.

Details

Only `test.type 1` (one-sided) and `test.type 4` (non-binding futility) are supported. Other test types (including `test.type 7` and `8` with harm bounds) will produce an error.

The exact binomial routine `gsBinomialExact` has requirements that may not be satisfied by the initial asymptotic approximation. Thus, the approximations are updated to satisfy the following requirements of `gsBinomialExact`: `a` (the efficacy bound) must be positive, non-decreasing, and strictly less than `n.I`; `b` (the futility bound) must be positive, non-decreasing, strictly greater than `a`; `n.I - b` must be non-decreasing and ≥ 0 .

With `'observedEvents'`, spending times are based on `observedEvents / x$maxn.Iplan`. If `maxSpend = TRUE`, the final spending time is set to 1 so all remaining spending is used at the last look. If `x$testLower` is present (for example from `gsSurv()` with selective futility looks), futility spending is flattened at analyses where `testLower = FALSE`.

Value

An object of class `gsBinomialExact`.

See Also

[gsBinomialExact](#)

Examples

```
# The following code derives the group sequential design using the method
# of Lachin and Foulkes
```

```

x <- gsSurv(
  k = 3,                      # 3 analyses
  test.type = 4,              # Non-binding futility bound 1 (no futility bound) and 4 are allowable
  alpha = .025,               # 1-sided Type I error
  beta = .1,                  # Type II error (1 - power)
  timing = c(0.45, 0.7),      # Proportion of final planned events at interims
  sfu = sfHSD,                # Efficacy spending function
  sfupar = -4,                # Parameter for efficacy spending function
  sfl = sflDOF,               # Futility spending function; not needed for test.type = 1
  sflpar = 0,                 # Parameter for futility spending function
  lambdaC = .001,             # Exponential failure rate
  hr = 0.3,                   # Assumed proportional hazard ratio (1 - vaccine efficacy = 1 - VE)
  hr0 = 0.7,                  # Null hypothesis VE
  eta = 5e-04,                # Exponential dropout rate
  gamma = 10,                 # Piecewise exponential enrollment rates
  R = 16,                     # Time period durations for enrollment rates in gamma
  T = 24,                     # Planned trial duration
  minfup = 8,                 # Planned minimum follow-up
  ratio = 3                    # Randomization ratio (experimental:control)
)
# Convert bounds to exact binomial bounds
toBinomialExact(x)
# Update bounds at time of analysis
toBinomialExact(x, observedEvents = c(20,55,80))
# Update exact efficacy bounds using a different alpha level
toBinomialExact(x, observedEvents = c(20,55,80), alpha = 0.01)
# Explicit spending-time override
toBinomialExact(x, observedEvents = c(20, 55, 80), usTime = c(.25, .65, 1))
# Optionally force full spending at final look when final events are below plan
toBinomialExact(x, observedEvents = c(20, 55, 75), maxSpend = TRUE)

```

toInteger	<i>Translate group sequential design to integer events (survival designs) or sample size (other designs)</i>
-----------	--

Description

Translate group sequential design to integer events (survival designs) or sample size (other designs)

Usage

```
toInteger(x, ratio = x$ratio, roundUpFinal = TRUE)
```

Arguments

x	An object of class gsDesign or gsSurv.
ratio	Usually corresponds to experimental:control sample size ratio. If an integer is provided, rounding is done to a multiple of ratio + 1. See details. If input is non integer, rounding is done to the nearest integer or nearest larger integer depending on roundUpFinal.

`roundUpFinal` For non-survival designs, final sample size is rounded up to a multiple of `ratio + 1` with the default `roundUpFinal = TRUE` if `ratio` is a non-negative integer. For survival designs, the final event count is rounded up with `roundUpFinal = TRUE`. If `roundUpFinal = FALSE` and `ratio` is a non-negative integer, sample size is rounded to the nearest multiple of `ratio + 1`. See details.

Details

It is useful to explicitly provide the argument `ratio` when a `gsDesign` object is input since `gsDesign()` does not have a `ratio` in return. `ratio = 0`, `roundUpFinal = TRUE` will just round up the sample size for non-survival designs. Since `x <- gsSurv(ratio = M)` returns a value for `ratio`, `toInteger(x)` will round to a multiple of `M + 1` if `M` is a non-negative integer; otherwise, just rounding will occur. For 1:1 randomization, `ratio = 1` gives an even final sample size. For 2:1 randomization, `ratio = 2` gives a final sample size that is a multiple of 3. To just round without concern for randomization ratio, set `ratio = 0`. If `toInteger(x, ratio = 3)`, rounding for final sample size is done to a multiple of `3 + 1 = 4`; this could represent a 3:1 or 1:3 randomization ratio. For 3:2 randomization, `ratio = 4` would ensure rounding sample size to a multiple of 5.

For a `gsSurv` object, `x$n.I` is an event-count schedule. `toInteger()` rounds the final planned event count (up when `roundUpFinal = TRUE`; otherwise to nearest integer, with a 0.01 tolerance), then derives interim integer event targets from `x$timing * final_events`. Interim counts are constrained to be positive and strictly increasing. Group sequential boundaries and spending are recomputed with `gsDesign()` at the integer event counts.

Total sample size for a survival design is then updated under a fixed calendar plan (same enrollment periods, study duration, and minimum follow-up). Enrollment rates are scaled proportionally to the final-event inflation factor and rounded to the nearest allocation multiple `ratio + 1` (or rounded up when `roundUpFinal = TRUE`), with additional allocation-step adjustment only if needed to make the integer final event target achievable.

If fixed-calendar enrollment-rate inflation cannot make the integer final event target feasible, `toInteger()` falls back to a variable-duration solve and issues a warning. For a complete seasonal exact-binomial monitoring workflow, see `vignette("MultiSeasonRareEvents", package = "gsDesign")`.

Selective-bound settings (`testUpper`, `testLower`, `testHarm`, and `harm` spending for `test.type 7` or `8`) are carried from the input design into the internal `gsDesign()` recomputation so skipped looks stay skipped after integer rounding.

Value

Output is an object of the same class as input `x`; i.e., `gsDesign` with integer vector for `n.I` or `gsSurv` with integer vector `n.I` and integer total sample size. See details.

See Also

[gsSurv](#), [toBinomialExact](#), `vignette("MultiSeasonRareEvents", package = "gsDesign")`

Examples

```
# The following code derives the group sequential design using the method
# of Lachin and Foulkes
```

```
x <- gsSurv(
  k = 3,                      # 3 analyses
  test.type = 4,              # Non-binding futility bound 1 (no futility bound) and 4 are allowable
  alpha = .025,               # 1-sided Type I error
  beta = .1,                  # Type II error (1 - power)
  timing = c(0.45, 0.7),      # Proportion of final planned events at interims
  sfu = sfHSD,                # Efficacy spending function
  sfupar = -4,                # Parameter for efficacy spending function
  sfl = sfLDOF,               # Futility spending function; not needed for test.type = 1
  sflpar = 0,                 # Parameter for futility spending function
  lambdaC = .001,             # Exponential failure rate
  hr = 0.3,                   # Assumed proportional hazard ratio (1 - vaccine efficacy = 1 - VE)
  hr0 = 0.7,                  # Null hypothesis VE
  eta = 5e-04,                # Exponential dropout rate
  gamma = 10,                 # Piecewise exponential enrollment rates
  R = 16,                     # Time period durations for enrollment rates in gamma
  T = 24,                     # Planned trial duration
  minfup = 8,                 # Planned minimum follow-up
  ratio = 3                    # Randomization ratio (experimental:control)
)
# Convert sample size to multiple of ratio + 1 = 4,
# with final event count rounded up by default.
toInteger(x)
```

<i>xtable</i>	<i>xtable</i>
---------------	---------------

Description

xtable

Value

an object of class "xtable"

Index

- * **Bernstein**
 - tEventsIA, 116
- * **Foulkes**
 - tEventsIA, 116
- * **Freedman**
 - tEventsIA, 116
- * **Lachin**
 - tEventsIA, 116
- * **Lagakos**
 - tEventsIA, 116
- * **Schoenfeld**
 - tEventsIA, 116
- * **and**
 - tEventsIA, 116
- * **design**
 - ciBinomial, 12
 - condPower, 18
 - eEvents, 25
 - gsBinomialExact, 27
 - gsBound, 33
 - gsBoundCP, 35
 - gsCP, 37
 - gsDensity, 41
 - gsDesign, 44
 - gsProbability, 51
 - nNormal, 64
 - normalGrid, 67
 - plot.gsDesign, 69
 - print.nSurvival, 72
 - sfExponential, 82
 - sfHSD, 84
 - sfLDOF, 85
 - sfLinear, 88
 - sfLogistic, 91
 - sfPoints, 94
 - sfPower, 96
 - sfTDist, 98
 - sfTruncated, 100
 - sfXG1, 103
 - summary.gsDesign, 108
 - summary.spendfn, 114
 - tEventsIA, 116
- * **hazards**
 - tEventsIA, 116
- * **non-inferiority**
 - tEventsIA, 116
- * **power**
 - tEventsIA, 116
- * **programming**
 - checkLengths, 10
- * **proportional**
 - tEventsIA, 116
- * **sample**
 - tEventsIA, 116
- * **size**
 - tEventsIA, 116
- * **stratified**
 - tEventsIA, 116
- * **super-superiority**
 - tEventsIA, 116
- * **survival**
 - tEventsIA, 116
- as_gt, 3, 3
- as_rtf, 4
- as_table, 3, 4, 7
- binomialPowerTable, 8
- binomialsPRT (gsBinomialExact), 27
- checkLengths, 10, 10
- checkRange, 10, 11
- checkRange (checkLengths), 10
- checkScalar, 10
- checkScalar (checkLengths), 10
- checkVector, 10
- checkVector (checkLengths), 10
- ciBinomial, 12
- condPower, 18, 37, 40

- eEvents, 25
- gsBinomialExact, 3, 4, 6, 7, 27, 129
- gsBound, 33
- gsBound1 (gsBound), 33
- gsBoundCP, 35, 40, 42
- gsBoundSummary, 50, 58, 122, 126
- gsBoundSummary (summary.gsDesign), 108
- gsBValue (summary.gsDesign), 108
- gsCP, 36, 37
- gsCPOS (gsCP), 37
- gsCPz (summary.gsDesign), 108
- gsDelta (summary.gsDesign), 108
- gsDensity, 41
- gsDesign, 20, 21, 27, 34, 36, 40, 42, 44, 47, 52, 53, 57, 58, 64, 71, 75, 83, 85, 87, 89, 93, 95, 97, 99, 101, 104, 111, 112, 115, 121, 124, 126
- gsDesign-package, 126
- gsHR, 27, 75, 126
- gsHR (summary.gsDesign), 108
- gsPI (gsCP), 37
- gsPOS (gsCP), 37
- gsPosterior (gsCP), 37
- gsPP, 42
- gsPP (gsCP), 37
- gsProbability, 31, 34, 36, 40, 42, 50, 51, 64, 71, 112
- gsRR (summary.gsDesign), 108
- gsSurv, 57, 58, 64, 131
- gsSurv (tEventsIA), 116
- gsSurvCalendar, 54, 64, 126
- gsSurvPower, 58, 58, 126
- hrn2z (print.nSurvival), 72
- hrz2n (print.nSurvival), 72
- is.integer, 10
- isInteger (checkLengths), 10
- nBinomial, 9
- nBinomial (ciBinomial), 12
- nBinomial1Sample (gsBinomialExact), 27
- nEvents (print.nSurvival), 72
- nEventsIA (tEventsIA), 116
- nNormal, 64
- Normal, 16, 50, 126
- normalGrid, 37, 39, 40, 67
- nSurv (tEventsIA), 116
- nSurvival, 26, 27, 122, 126
- nSurvival (print.nSurvival), 72
- plot.binomialsPRT (gsBinomialExact), 27
- plot.gsBinomialExact (gsBinomialExact), 27
- plot.gsDesign, 27, 50, 53, 69, 75, 112, 126
- plot.gsProbability (plot.gsDesign), 69
- plot.ssrCP (condPower), 18
- Power.ssrCP (condPower), 18
- print.data.frame, 108
- print.eEvents (eEvents), 25
- print.gsBinomialExact (gsBinomialExact), 27
- print.gsBoundSummary (summary.gsDesign), 108
- print.gsDesign, 44
- print.gsDesign (summary.gsDesign), 108
- print.gsProbability (gsProbability), 51
- print.gsSurv (tEventsIA), 116
- print.nSurv (tEventsIA), 116
- print.nSurvival, 72
- print.xtable, 108
- repeatedPValueBinomialExact, 77
- sequentialPValue, 78
- sequentialPValueBinomialExact, 80
- sfBetaDist (sfLogistic), 91
- sfCauchy (sfLogistic), 91
- sfExponential, 82, 84, 86, 115
- sfExtremeValue (sfLogistic), 91
- sfExtremeValue2 (sfLogistic), 91
- sfGapped (sfTruncated), 100
- sfHSD, 84, 115
- sfLDOF, 85
- sfLDPocock (sfLDOF), 85
- sfLinear, 88
- sfLogistic, 91, 95, 115
- sfNormal (sfLogistic), 91
- sfPoints, 94
- sfPower, 96, 115
- sfStep (sfLinear), 88
- sfTDist, 98
- sfTrimmed (sfTruncated), 100
- sfTruncated, 100, 115
- sfXG (sfXG1), 103
- sfXG1, 103
- sfXG2 (sfXG1), 103

sfXG3 (sfXG1), [103](#)
simBinomial, [9](#)
simBinomial (ciBinomial), [12](#)
simBinomialSeasonalExact, [106](#)
spendingFunction, [46](#), [56](#), [121](#)
spendingFunction (summary.spendfn), [114](#)
ssrCP, [37](#), [40](#)
ssrCP (condPower), [18](#)
summary.gsDesign, [108](#)
summary.spendfn, [114](#)

testBinomial (ciBinomial), [12](#)
tEventsIA, [116](#)
toBinomialExact, [128](#), [131](#)
toInteger, [130](#)

uniroot, [16](#), [62](#), [118](#), [126](#)

varBinomial (ciBinomial), [12](#)

write.csv, [108](#)
write.csv2, [108](#)
write.table, [108](#)

xprint, [122](#), [126](#)
xprint (summary.gsDesign), [108](#)
xtable, [50](#), [126](#), [132](#)
xtable.gsDesign (gsDesign), [44](#)
xtable.gsSurv, [108](#), [112](#)
xtable.gsSurv (tEventsIA), [116](#)

z2Fisher (condPower), [18](#)
z2NC (condPower), [18](#)
z2Z (condPower), [18](#)
zn2hr (print.nSurvival), [72](#)